

# PATE Gene Clusters Code for Multiple, Secreted TFP/Ly-6/uPAR Proteins That Are Expressed in Reproductive and Neuron-rich Tissues and Possess Neuromodulatory Activity\*

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We report here syntenic loci in humans and mice incorporating gene clusters coding for secreted proteins each comprising 10 cysteine residues. These conform to three-fingered protein/Ly-6/urokinase-type plasminogen activator receptor (uPAR) domains that shape three-fingered proteins (TFPs). The founding gene is *PATE*, expressed primarily in prostate and less in testis. We have identified additional human *PATE*-like genes (*PATE-M*, *PATE-DJ*, and *PATE-B*) that co-localize with the *PATE* locus, code for novel secreted *PATE*-like proteins, and show selective expression in prostate and/or testis. Anti-*PATE-B*-specific antibodies demonstrated the presence of *PATE-B* in the region of the sperm acrosome and at high levels on malignant prostatic epithelial cells. The syntenic mouse *Pate*-like locus encompasses 14 active genes coding for secreted proteins, which are all, except for *Pate-P* and *Pate-Q*, expressed primarily in prostate and/or testis. *Pate-P* and *Pate-Q* are expressed solely in placental tissue. Castration up-regulates prostate expression of mouse *Pate-B* and *Pate-E*, whereas testosterone ablates this induced expression. The sequence similarity between TFP/Ly-6/uPAR proteins that modulate activity of nicotinic acetylcholine receptors and the *PATE* (*Pate*)-like proteins stimulated us to see whether these proteins possess analogous activity. Pharmacological studies showed significant modulation of the nicotinic acetylcholines by the *PATE-B*, *Pate-C*, and *Pate-P* proteins. In concert with these findings, certain *PATE* (*Pate*)-like genes were extensively expressed in neuron-rich tissues. Taken together, our findings indicate that in addition to par-

ticipation of the *PATE* (*Pate*)-like genes in functions related to fertility and reproduction, some of them likely act as important modulators of neural transmission.

A functional genomic approach has identified a gene, designated *PATE*, that codes for a secreted protein with preferential prostate and testis expression (1). *PATE* protein includes 10 cysteine residues, with the C-terminal cysteine residue positioned within a cysteine-asparagine (CN) dipeptide sequence. The distribution of cysteine residues conforms to a consensus cysteine pattern found in a large family of three-fingered proteins (TFPs),<sup>3</sup> characterized by a distinct disulfide bonding pattern between 8 and 10 cysteine residues (2, 3). This domain is additionally found in uPAR and murine Ly-6 GPI-anchored proteins, and is also called an Ly-6/uPAR domain (3). Interestingly, the TFP architecture is seen also in the transforming growth factor- $\beta$  receptor family of proteins, including BMP2 and activin receptors (4).

A large protein family encompassing an extensive group of GPI-anchored, transmembrane, and secreted proteins contains this domain. The prototype-secreted protein members of this family include short chain snake and frog toxins, which in many cases bind with high affinity to neuronal receptors and block their activity (5–7).

Until recently, the only recognized secreted mammalian TFP/Ly-6/uPAR proteins were SLURP-1 (secreted mammalian Ly-6/uPAR-related protein) (8) and SLURP-2 (9), which are located on human chromosome 8q24.3 within a cluster of Ly-6-like human genes that otherwise code for GPI-linked proteins. Consistent with the putative ligand function of some secreted TFP/Ly-6/uPAR proteins, SLURP-1 was recently identified as a neuromodulator of the  $\alpha 7$  nicotinic receptor ( $\alpha 7$  nAChR), suggesting that it may regulate calcium homeostasis (10). It is clear that members of the secreted TFP/Ly-6/uPAR protein family,

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The nucleotide sequence(s) reported in this paper has been submitted to the GenBank™/EBI Data Bank with accession number(s) EU703625, EU703626, EU703627, EU703628, and EU703629.

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<sup>3</sup> The abbreviations used are: TFP, three-fingered protein; uPAR, urokinase-type plasminogen activator receptor; ACh, acetylcholine; nAChR, nicotinic acetylcholine receptor; RT, reverse transcription; GPI, glycosylphosphatidylinositol; h, human; m, mouse; d, dog; TEV, tomato etch virus; PNGase F, peptide:N-glycosidase F; DHT, dihydroxytestosterone.

to which the PATE protein belongs, interact with partner proteins that, in many cases, are membrane-tethered receptors.

The human *PATE* gene is telomerically juxtaposed to the gene encoding acrosomal vesicle protein 1 (*ACRV1*), also known as the *SP10* gene (11). Interestingly, the *ACRV1* protein also contains 10 cysteine residues that conform to the TFP/Ly-6/uPAR domain, suggesting that the two genes *ACRV1* and *PATE* may be part of a single chromosomal locus comprising TFP/Ly-6/uPAR genes.

In this study, we report the identification and expression patterns of three additional human *PATE*-like genes (*PATE-M*, *PATE-DJ*, and *PATE-B*) that co-localize with the *ACRV1* and *PATE* genomic locus. These novel *PATE*-like genes code for secreted proteins containing the typical TFP/Ly-6/uPAR domain. Significantly, all show selective expression in prostate and/or testis. We have identified the orthologous murine *Pate*, *Pate-M*, *Pate-DJ*, and *Pate-B* genes that localize centromerically to the mouse *Acrv1/sp10* gene. Remarkably, the mouse *Pate*-like genomic locus includes an additional nine transcriptionally active *Pate*-like genes, which all encode secreted TFP/Ly-6/uPAR-domain-containing proteins, whereas in the human genome these mouse *Pate*-like genes are either inactive (two genes) or completely absent (the remaining seven genes). Selective expression of the mouse *Pate*-like genes in prostate, testis, placenta as well as specific effects of castration and subsequent testosterone administration on their expression in prostate all indicate that these genes function in both male- and female-related reproductive activities and are likely hormonally regulated. Furthermore, because of the sequence similarity between TFP/Ly-6/uPAR proteins that modulate activity of nicotinic acetylcholine receptors (nAChRs) and the *PATE* (*Pate*)-like proteins, we conducted experiments to see whether these proteins also possess comparable activity. These analyses showed that certain *PATE* (*Pate*)-like proteins modulate the activity of nAChRs. Taken together, our findings indicate that in addition to participation of the *PATE* (*Pate*)-like genes in functions related to fertility and reproduction, they are likely to function as important modulators of neural activity.

## EXPERIMENTAL PROCEDURES

**Materials and Antibodies**—Unless otherwise specified, chemicals and reagents were obtained from Sigma. The anti-FLAG antibodies were affinity-purified rabbit polyclonal antibodies. Fresh ACh stock solutions were made daily in Ringer's solution and diluted.

**RT-PCR Analyses of the Human and Mouse *PATE* (*Pate*)-like Genes**—Forward and reverse oligonucleotide primers were synthesized using the DNA sequences obtained from the *PATE* (*Pate*)-like sequences (see under "Bioinformatic Strategies for Identification of *PATE* (*Pate*)-like Genes"). RT-PCR analysis of the human *PATE*-like genes (and flanking genes) as well as the mouse *Pate*-like genes was performed with cDNAs obtained from different human or mouse tissues (Clontech) as indicated. Forward and reverse primers were chosen such that they always spanned an intron, and the observed RT-PCR product at all times corresponded to the size expected of a spliced mRNA.

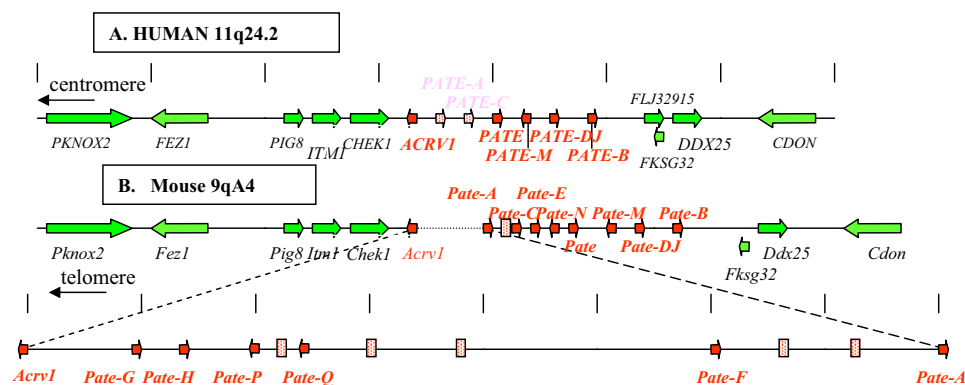
**Sequencing of *PATE* (*Pate*)-like cDNAs**—All human *PATE*-like cDNAs were either directly sequenced from gel-purified

RT-PCR DNAs or alternatively the gel-purified cDNAs were cloned into TOPO4 (Invitrogen) plasmids and then sequenced. For the mouse *Pate*-like cDNAs, RT-PCR generated DNAs were gel-purified and directly sequenced.

**Bioinformatic Strategies for Identification of *PATE* (*Pate*)-like Genes**—The sequence homology among the TFP family of proteins is generally low except for the common pattern of cysteines in the sequence, and an initial attempt to identify *PATE*-like genes by using conventional protein homolog search programs such as BLAST or BLAT was not successful. However, a careful inspection of protein sequences and exon structures of initially identified *PATE* and *PATE*-like genes enabled us to devise a method to detect *PATE*-like genes in the genomic sequences. We developed two protein sequence patterns, P1 and P2, to represent C1–C5 and C6–C10N patterns, respectively, found in *PATE*-like proteins. The two patterns are "CX(2)CX(5,10)CX(3,8)CX(4,9)C" for P1 and "CX(2,4)CX(11,20)CCX(2,7)C" for P2, where  $X(n,m)$  denotes a stretch of any amino acids ranging in length from  $n$  to  $m$ . The patterns were reverse-translated and transformed to Perl regular expressions. We searched the genomic locus bounded by *PKNOX2* and *CDON* genes in the human genome (May 2004 freeze) and orthologous loci in the mouse genome (May 2004 freeze), in the rat genome (June 2003 freeze), and in the dog genome (July 2004 freeze). Two exons bearing P1 and P2, respectively, were separately searched. Then, closely (less than 10 kb) located P1 and P2 pairs were joined to form single genes. In human and mouse, the precise exon boundaries were predicted by manually inspecting all possible intron-exon boundaries for those that will maintain the open reading frame and the experimentally determined protein sequence. The corresponding signal peptide-bearing exon was located in the 5'-flanking region of each gene. The putative gene structure was verified by expressed sequence tag search, by comparing each gene with a respective ortholog in human or mouse, or experimentally by RT-PCR, cloning, and sequencing. Possible orthologous relationships among *PATE*-like genes from human, mouse, rat, and dog were inferred by a phylogenetic analysis using MEGA3 program (12) based on a multiple sequence alignment of P1 and P2 sequences obtained by using T-Coffee program (13).

**Generation of Eucaryotic Expression Constructs and Fusion Proteins**—Cloning was conducted with the eucaryotic expression vector pCMV3 (Sigma) via selected restriction sites. This vector codes for the preprotrypsin signal peptide followed by sequences coding for the FLAG epitope. DNA coding for the human Fc fragment (hFc) was inserted 3' to the FLAG epitope. A cleavage site for the highly specific tomato etch virus (TEV) protease was also introduced between the C terminus of the *Pate*-like proteins and the hFc segment. cDNA fragments encoding the *Pate*-like proteins were subcloned in-frame into the pCMV3 (5'FLAG—hFc3') vector to render pCMV3 as 5'FLAG-*Pate*-like-TEV-hFc3'.

**Generation of HEK293 Transfectants Expressing FLAG-*Pate*-TEV-hFc Proteins**—HEK293 (human kidney) cells were transiently transfected with the eucaryotic pCMV3 expression vectors (6  $\mu$ g of DNA/25-cm<sup>2</sup> flask) coding for the FLAG-(*Pate*-like)-TEV-hFc fusion proteins. The secreted C-terminally hFc



**FIGURE 1. Arrangement of genes within the locus comprising the human *PATE*-like genes and the corresponding syntenic murine genomic locus.** *A*, known human genes lie within a genomic segment on chromosome 11q24 initiating at the centromeric side with *PKNOX2* at nucleotide 124,726,419 (numbering as in the Genome Browser May 2004 release), and terminating at the telomeric side with *CDON* at nucleotide 125,438,397. Arrows indicate direction of transcription. The genes coding for proteins containing the distinctive 10-cysteine motif are shown in red, and inactive pseudogenes are *stippled*. Accession numbers are as follows: *ACRV1*, NM\_001612; *PATE*, NM\_138294; *PATE-M*, NM\_212555; *PATE-B*, AK123042. Accession number for *PATE-DJ* is EU703625. *B*, syntenic murine (chromosome 9qA4) genomic locus. Note the 0.8-Mbp insertion in the mouse genome between *Acrv1* and *Pate-A*. Accession numbers are as follows: *Acrv1*, NM\_007391; *Pate-H*, BY721155; *Pate-Q*, BQ032923; *Pate-F*, BY721010; *Pate-A*, AK020329; *Pate-C*, NM\_026593; *Pate-E*, AV379335; *Pate*, AK033745; *Pate-M*, BY721028; *Pate-B* (*svs7*), NM\_020264. Accession numbers for *Pate-G*, *Pate-P*, *Pate-N*, and *Pate-DJ* are EU703627, EU703629, EU703628, and EU703626, respectively.

tagged FLAG-Pate-like-TEV-hFc proteins contained in the conditioned media (CM) were collected on protein A-Sepharose 4 Fast Flow resin (Amersham Biosciences), and the N-terminal Pate-like proteins were released by incubating the protein A beads with TEV protease.

**Preparation of RNA**—Human nAChR clones were obtained from Dr. Jon Lindstrom. After linearization and purification of cloned cDNAs, RNA transcripts were prepared *in vitro* using the appropriate mMessage mMachine kit from Ambion Inc. (Austin, TX).

**Expression in *Xenopus* Oocytes**—Mature (>9 cm) female *Xenopus laevis* African toads (Nasco, Ft. Atkinson, WI) were used as a source of oocytes. Prior to surgery, frogs were anesthetized by placing the animal in a 1.5 g/liter solution of MS222 (3-aminobenzoic acid ethyl ester; Sigma) for 30 min. Oocytes were removed from an incision made in the abdomen.

To remove the follicular cell layer, harvested oocytes were treated with 1.25 mg/ml collagenase (Worthington) for 2 h at room temperature in calcium-free Barth's solution (88 mM NaCl, 1 mM KCl, 2.38 mM NaHCO<sub>3</sub>, 0.82 mM MgSO<sub>4</sub>, 15 mM HEPES (pH 7.6), 0.1 mg/ml gentamicin sulfate). Subsequently, stage 5 oocytes were isolated and injected with 50 nl (5–20 ng) each of the appropriate subunit cRNAs. Recordings were made 3–5 days after injection.

**Electrophysiology**—Experiments were conducted using OpusXpress 6000A (Axon Instruments, Union City, CA) as reported previously (14). OpusXpress is an integrated system that provides automated impalement and voltage clamp of up to eight oocytes in parallel. Cells were automatically perfused with bath solution, and agonist solutions were delivered from a 96-well plate. Both the voltage and current electrodes were filled with 3 M KCl. The agonist solutions were applied via disposable tips, which eliminated any possibility of cross-contamination. Cells were voltage-clamped at a holding potential of –60 mV. Data were collected at 50 Hz and filtered at 20 Hz. ACh applications were 8 s with 241-s wash periods.

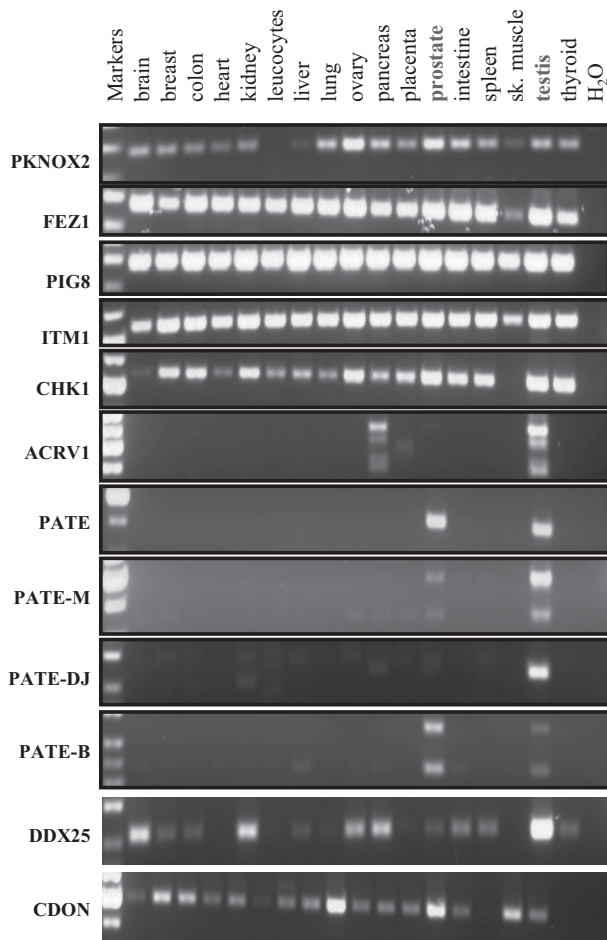
Each oocyte received two initial control applications of ACh, and then the PATE-like peptides were pre-applied for 241 s at the indicated concentrations through an alternative supply of bath solution. Subsequently the PATEs were co-applied with ACh at the control concentration. The control ACh concentrations for a7 and a4b2 receptors were 60 and 30 μM, respectively. Responses to the ACh PATE co-application were calculated relative to the preceding ACh control responses based on net charge (15). Net charge was integrated for the entire response, *i.e.* until the currents return to base line, such that the total time window for the net charge measurement was 120 s, beginning 2 s prior to the ACh delivery. Responses of at least four

oocytes were measured for each experimental concentration. Statistical analyses of PATE effects were based on pairwise *t* test between the responses of each oocyte to ACh alone or ACh plus PATE peptide, following preincubation with the PATEs. All of the statistics were based on pairwise *t* tests, where responses of cells recorded under control conditions were compared on a cell by cell basis with those obtained after PATE treatment. The calculations of *p* values were made with Stateview version 4.01, (ABACUS Concepts, Berkeley, CA).

**SDS-PAGE**—SDS-polyacrylamide gel for protein separation was performed as described previously (16), and blots were reacted with polyclonal rabbit anti-FLAG primary antibody. For glycosidase treatment, protein A-purified proteins were incubated with 1 unit of PNGase F (New England Biolabs).

## RESULTS

**Identification of a Human *PATE*-like Gene Cluster**—The *PATE* gene codes for a small, cysteine-rich protein selectively expressed in human male reproductive tissues, including prostate, testis, epididymis, and seminal vesicle (1, 17). Pattern-search techniques (see “Experimental Procedures”) revealed three additional *PATE*-like genes, designated *PATE-B*, *PATE-M*, and *PATE-DJ*, which localized to the same 11q24 genomic locus as the *PATE* gene (Fig. 1). Expression analyses of these genes by RT-PCR in 17 different human tissues demonstrated selective expression in prostatic and/or testicular tissue with negligible expression in all other tissues (Fig. 2). The *PATE-B* gene was expressed primarily in prostate with lesser expression in testis, whereas the *PATE-M* and *PATE-DJ* genes showed a reverse pattern of expression (Fig. 2). RT-PCR analyses of all known genes within a stretch of 700 kbp comprising the human *PATE*-like genes demonstrated that the non-*PATE*-like genes did not show preferential expression in reproductive tissues (Fig. 2). Consistent with previously published data, *ACRV1* (acrosomal vesicle protein 1 gene also known as *SP10*)



**FIGURE 2. RT-PCR expression analysis of human *PATE*-like genes and flanking genes.** RT-PCR analysis of the human *PATE*-like genes and flanking genes was performed with cDNAs obtained from the indicated human tissues. Forward and reverse primers were chosen such that they always spanned an intron, and the observed RT-PCR product at all times corresponded to the size expected of a spliced mRNA. For the *ACRV1*, *PATE*, *PATE-M*, *PATE-DJ*, and *PATE-B* genes, the forward and reverse primers were located in the first and third exons (coding for the signal peptide and cysteines 6–10, respectively). PCR was performed for 35 cycles. Note that *PIG8* (*p53* induced gene 8) is ubiquitously expressed in all tissues, serving as a convenient internal control for cDNA integrity.

demonstrated multiple splice isoforms, expressed primarily in testicular tissue (11).

Sequencing full-length cDNAs showed that the human *PATE*-like genes code for similar proteins that all include a putative N-terminal signal peptide (see below) and 10 conserved cysteine residues (Table 1). Comparison of genomic and cDNA sequences showed that in all *PATE*-like proteins the N-terminal signal peptide is encoded by the first exon (exon 1), whereas protein domains containing cysteines 1–5 and cysteines 6–10 are encoded by two separate 3' exons (exons 2 and 3, respectively, shown in Fig. 3). For *PATE* and *PATE-M* genes, 1–2 additional exons, designated 1a and 1b, are present between exons 1 and 2 and code for a small number of amino acids. The upstream *ACRV1* gene shows a similar exon structure to the *PATE*-like genes.

All splice events in the *PATE*-like genes use a +1 phase (Fig. 3); thus alternative splicing involving exon skipping will not alter downstream reading frames. Interestingly, *PATE-B* and

*PATE-M* generate two splice isoforms (Fig. 2). Cloning and sequencing of the smaller transcripts confirmed that they derive from exon 2 skipping. Both isoforms include the putative N-terminal signal peptide, but although the larger transcript codes for proteins comprising all 10 cysteine residues, the smaller transcript codes only for cysteines 6–10.

**Prediction of Signal Peptides at the N Terminus of All Human *PATE*-like Genes**—As expected of signal peptide sequences, analysis of all human *PATE*-like genes reveals clustering of hydrophobic amino acid residues just distal to the initiating methionine (Table 1). Analysis by the SignalP (signal peptide prediction) algorithm predicts with very high probability a signal peptide and accompanying cleavage site for each *PATE*-like protein (Table 1). That this N-terminal region is in fact a signal peptide directing protein for secretion has been functionally demonstrated for *PATE* (17) and *ACRV1* (*SP10*) (18), and it is very likely that these regions serve the same function in the *PATE-B*, *PATE-DJ*, and *PATE-M* proteins. By transfecting human HEK293 cells with cDNA coding for the native *PATE-B* protein and analyzing spent culture medium for the presence of *PATE-B* protein, we have experimentally confirmed (data not shown) that the native *PATE-B* protein is indeed secreted from the cell.

**Northern Blot Analysis of *PATE-B* and *PATE-M* Expression**—To extend and confirm the RT-PCR analyses, Northern blot analyses of *PATE-B* and *PATE-DJ* were performed (Fig. 4). This demonstrated exclusive expression, albeit at low levels, of *PATE-B* and *PATE-DJ* in prostate and testis, respectively, confirming the RT-PCR analyses; no other tissues expressed these genes.

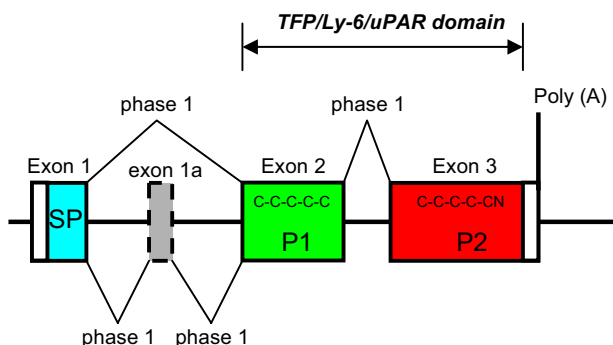
**Additional Vestigial Inactive *PATE*-like Genes in the Human *PATE*-like Gene Cluster**—Searches for additional human *PATE*-like genes within the human gene cluster revealed two potential *PATE*-like genes, designated *PATE-A* and *PATE-C* (Fig. 1). Extensive RT-PCR analyses using a number of forward and reverse primers based on these genomic sequences failed, in all tissues examined, to reveal expression of these genes.

**Identification of 14 Active Mouse *Pate*-like Genes within the Mouse Genome Syntenic to the Human *PATE*-like Genomic Locus**—The human *PATE*-like gene locus extends from the *ACRV1* gene to the *PATE-B* gene, encompassing about 180 kbp. The human genes adjacent to the *PATE*-like gene locus are (centromeric → telomeric, Fig. 1A) as follows: *PKNOX2*, *FEZ1*, *PIG8*, *ITM1*, *CHEK1*, *ACRV1*, (*PATE-A*), (*PATE-C*), *PATE*, *PATE-M*, *PATE-DJ*, *PATE-B*, *DDX25* and *CDON*, and this region stretches for about 700 kbp. The syntenic mouse genomic segment was identified and demonstrated the following arrangement (telomeric → centromeric, Fig. 1B): *Pknox2*, *Fez1*, *Pig8*, *Itm1*, *Chk1*, *Acrv1* (*sp10*), *Svs7* (seminal vesicle secretion protein 7 (19) also called caltrin (20), identified here as the mouse *Pate-B* ortholog), *Ddx25* and *Cdon* (Fig. 1). Further analyses designed to identify mouse orthologs revealed the mouse *Pate-A*, *Pate-C*, *Pate*, *Pate-M*, *Pate-DJ*, and *Pate-B* (*svs7*) genes (Fig. 1B). An additional two potential mouse *Pate*-like genes, designated *Pate-E* and *Pate-N*, were identified between *Pate-C* and *Pate*. Unexpectedly, inspection of the mouse genomic *Pate*-like gene locus revealed 0.8 Mbp of genomic DNA situated

**TABLE 1**
**Sequences of the human PATE-like and mouse Pate-like proteins and signal peptide prediction**

The amino acid sequences of the human PATE-like (and mouse Pate-like) proteins are presented starting with the initiating methionine. Upper panel, sequences were subjected to SignalP analysis-signal peptide probabilities, and the predicted signal peptide cleavage sites are presented. The upward facing arrow designates the predicted cleavage site of the signal peptide, and the red dots indicate the exon boundaries. Hydrophobic amino acid residues downstream to the initiating methionine, and likely comprising the signal peptide, are highlighted. Lower panel, exons 2 and 3 include cysteines 1–5 and cysteines 6–10, respectively.

PATE-like Protein	Exon 1. (signal peptide)	HMM/SigP	Exon 1a.	Exon 1b.	Tissue Expression
PATE	MDKSLLELIPILLCCFR.ALSG↑SL	YES	.ALSGSLSMRND.	.VNEIVAVKNNF.	Pro>Tes
PATE-M	MLVLFLLGTVFLFCPYW.G↑EL	YES	.GELHDPKIG.		Tes>Pro
PATE-DJ	MNKHFLFLFLLYCLIV.AVTSL↑QC	YES			Tes
PATE-B	MNTLLVLSISFLYLKE.VMG↑LK	YES			Pro>Tes
Pate-G	MYAAFCSSLLGSHC↑NWKIPLDTPAWHLLA.TL	YES			Pro>Tes>1 other
Pate-H	MNPVTKISTLLIVTLPFICFAE.A↑LKCF	YES			Pro
Pate-P	MGKHLVQLLLVLSLLVMSLQ.A↑LT	YES			Pla
Pate-Q	MGKHILQLLLVLSLLVMSSQ.A↑LT	YES			Pla
Pate-F	MSKLLLLLLGAFVVLVIQ.AQA↑IIC	YES			Tes
Pate-A	MGKLLLLLHFLMQASFALVFIQ.VQA↑TV	YES			Pro, Tes
Pate-C	MKNFLRLCLFLLCFET.GFP↑LQ	YES			Pro
Pate-E	MGKVQELSIVLLCMQT.ALA↑LM	YES			Pro, Tes
Pate-N	MDWLLFLLPGVLILYKSHM.G↑EP	YES			Tes
Pate	MCKSQLRLCLIFLCYIK.VFSG↑SL	YES	.VFSGSLPGDANK.	.PGKVLVHENNN.	Pro, Tes
Pate-M	MEVLVMICLFCQYW.VLN↑ELE	YES	.VLNELEE.		Tes>Pro>2 others
Pate-DJ	MNKHFLLLSFLYFIV.EATS↑LKCV	YES			Tes
Pate-B	MNSVTKISTLLIVLSFLCFVE.G↑LI	YES			Pro>3 others
PATE-like Protein	Exon 2 (cysteines #1-#5)	Exon 3 (cysteines #6-#10)			
PATE	.PVIEIVQCRMCCHLQFPPEKCSRRICTATTE.ACMVIRMKRD.	.NPWITFMCLKNCADVKIRWSVYLVNFRCCRSDDLQEDL*			
PATE-M	.ATEIMCYECKYHL.LCYVMYSCSLKHK.SCAVENFYLTRK.	.QSMYHYSKLSGMTSCEDI.NFLFTKRVELICCDHSNYCNLPEIV*			
PATE-DJ	.AVTSLQCITCHLRRTDRRRFVCTAQK.ACMLLRIYQ.	.RNTLQISYMVCCQKCRDM.TFDLRNRTYVHTCCN.YNYCNFKL*			
PATE-B	.VMILKNCICYTEWK.CMARRTCIAKEN.LCSTTAYFR.	.DKHMYSHMCKYKREE.ESSKRILRVLLCCD.RNFCNVF*			
Pate-G	.TLLCYKCKHFNSTKNCLTQSVCVARRD.KCCL.WTVTS	.DDFLSYAATCWTCHVNVK.YTIRSVRSEHKCCNSSSLCNQF*			
Pate-H	.ALKCFQCTLFNSK.KCLFQEPPEQNN.VCVL.WAKEE.	.GRFMYFTECSHTCVNQ.TLNLRNKRTEMKCCNDKSFCKNF*			
Pate-P	.AITCTCDRMNSRRICEKECCQARPE.KCAS.FILK.	.DKIQFSSRCADLCFT.TVMTDKTVKMNCCNNKSFCKNL*			
Pate-Q	.AITCTCDRINSQICESKECCQAKPE.KCAS.LILK.	.DKIQFSSRCANICFT.TVQT.DQTVKMKCCCKRSFCNEL*			
Pate-F	.AQAICMCSNMFV.NKCIETE.KCTMEEACACRT.DIYHFHPR.	.DFLYNHTILECSKSCAS.EESYFHLKILFCCKSQDFCKNYKIK*			
Pate-A	.VQAVCMVCKSPK.SHCLVKNNTRYKP.CRTRNYFLSHT.	.KVVHNHTELDCKACMAE.NMYLAKKISFCCKEDFCNKYHQVNNKNIY*			
Pate-C	.FPIQCQCQSYK.NECATKKTCTKPE.TCMIRRTWYANE.	.IHNLDQAEKTCNSCKFE.EKTSYLTTHYCCSHDFCNDINLPIVMT*			
Pate-E	.ALALMCRECKSY.HQKCIHEMRTCTAKD.SCLT.VRLWISPS.	.SVHTPNDAYSRCQENCTD.EYYYDYITLICKCKKFDENDIAPVSDWS*			
Pate-N	.EPTFCTSCDEYI.DKCTRNLVCHARYPFACQTEVYIQLNT.	.EYLYKYSVLCPRRCEVYVRFIRFEKNI.FSCCN.ESYCNSFSKSDTQFKER*			
Pate	.VVEIVQCRMCCHLQFPENCTRRICTATAE.ACMAKVEKRD.	.ITWVNFMCCLKNCANVKKKWVPLVDVFCRRQDMCNENF*			
Pate-M	.EDRILCYKCKKYH.LCYIMTSCVPNHR.TCAENFYILTCK.	.QSMYHYSRLSCMTNCEDI.NELSFERTEELICCKHSNYCNLPMIL*			
Pate-DJ	.EATSCKVCTCHLRQSDHRRFVCLAQKH.ICMSLRIYES.	.SLQISYMVCCQKCRK.NLTYIFNNRTYTHKCCN.SDFCNFRL*			
Pate-B	.ILICNSCEKSR.DSRCTMPQSRCAKPE.SCSTVSHFV.	.TKHVYSKMCSPQCKEKQLNTKKLIYMCCE.KNLCSNF*			



**FIGURE 3. Exon structure of PATE (Pate)-like genes.** The canonical exon structure of the PATE (Pate)-like genes is presented. Optional extra exons are represented as dotted boxes. The coding region is in color: blue, signal peptide (SP); green, pattern 1 (P1) containing C1 to C5; red, pattern 2 (P2) containing C6 to C10.

between the mouse *Acrv1* and *Pate-A* genes. Analysis of this segment revealed several additional *Pate*-like genes (Fig. 1 and Table 1). The expected human equivalent to the 0.8-Mbp mouse fragment was completely absent.

All mouse *Pate*-like genes code for putative *Pate*-like proteins comprising a hydrophobic N-terminal signal peptide (Table 1). RT-PCR analyses (Fig. 5) utilizing oligonucleotide primers located in exons 2 and 3 revealed that the mouse *Pate-A* and *Pate-C* mouse genes (corresponding to the inactive human *PATE-C* and *PATE-A* genes) were clearly expressed, in addition to expression of the mouse orthologs *Pate-B*, *Pate-DJ*, *Pate-M*, and *Pate*. Even more striking was the discovery of an additional seven transcriptionally active mouse *Pate*-like genes, designated *Pate-E*, *Pate-H*, *Pate-N*, *Pate-F*, *Pate-P*, *Pate-Q*, and *Pate-G* (Figs. 1 and 5 and Table 1).

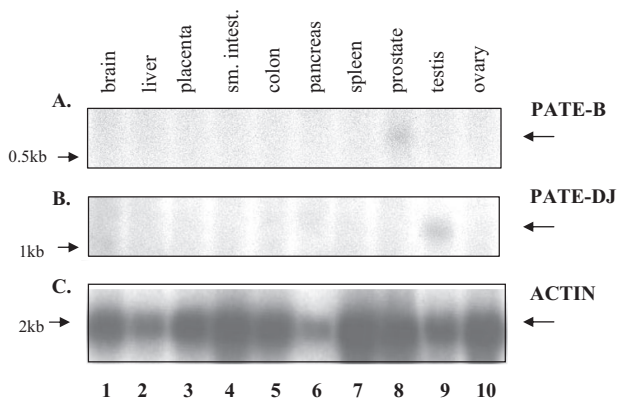


FIGURE 4. Northern blot analyses of *PATE-B* and *PATE/DJ* expression. *PATE-B*, *PATE/DJ*, and actin cDNAs were radioactively ( $^{32}\text{P}$ ) labeled and each used to sequentially probe, under stringent wash conditions, a Northern blot (Clontech) of total RNA derived from the indicated human tissues. The blot was stripped between sequential probing. Expression of *PATE-B* and *PATE/DJ* mRNAs is clearly visible in prostatic and testicular tissues (lanes 8 and 9, respectively); no signal was seen in any other tissues examined.

We do not know why the syntenic mouse locus harbors so many more active *Pate*-like genes as compared with the human genome, but this does suggest a significantly more complex role for the *Pate*-like proteins in rodents.

Constituent *Pate*-like genes could be segregated according to their tissue expression profiles (Fig. 5). Genes *Pate-E*, *Pate-A*, and *Pate* were predominantly expressed in both prostate and testis, whereas *Pate-C* and *Pate-H* expression was limited to prostatic tissue. *Pate-N*, *Pate-F*, and *Pate-DJ* showed almost exclusive expression in testis. *Pate-G*, *Pate-B*, and *Pate-M* all showed major expression in either prostate or testis but, in addition, showed significant expression levels in skeletal muscle (*Pate-G*), eye, kidney, skeletal muscle (*Pate-B*), and brain and lung (*Pate-M*). Several mouse *Pate*-like pseudogenes were scattered among the active mouse *Pate*-like genes (Fig. 1).

**Expression in Placental Tissue of Two Mouse *Pate*-like Genes—**Two murine *Pate*-like genes, *Pate-P* and *Pate-Q*, were expressed exclusively in the female-restricted organ, mouse placenta (Fig. 5). Notably, these genes (a) are genomically adjacent to each other (Fig. 1), (b) code for highly similar proteins (Table 1), and (c) both include an 11th cysteine residue in addition to the consensus *Pate*-like 10 cysteine residues. Supporting placental expression reported here is a trophoblast cDNA library EST (BQ032923) representing a partial *Pate-Q* sequence.

**Castration Induces *Pate*-like Gene Expression in the Ventral Prostate That Is Ablated by Subsequent Dihydroxytestosterone Administration—**The selective tissue distribution of *Pate*-like gene expression in the testis, prostate, and placenta indicated that the *Pate*-like proteins are likely involved in reproductive-related behavior.

To see whether *in vivo* hormonal changes may affect *Pate*-like gene expression, we investigated effects of castration and subsequent dihydroxytestosterone (DHT) administration on prostate expression of the *Pate*-like genes. As differential gene expression has been documented in the anatomically discrete dorsal and ventral prostate lobes, we investigated *Pate*-like gene expression in each separate lobe. The dorsal lobe in uncastrated mice clearly expressed the two *Pate*-like genes, *Pate-B* and

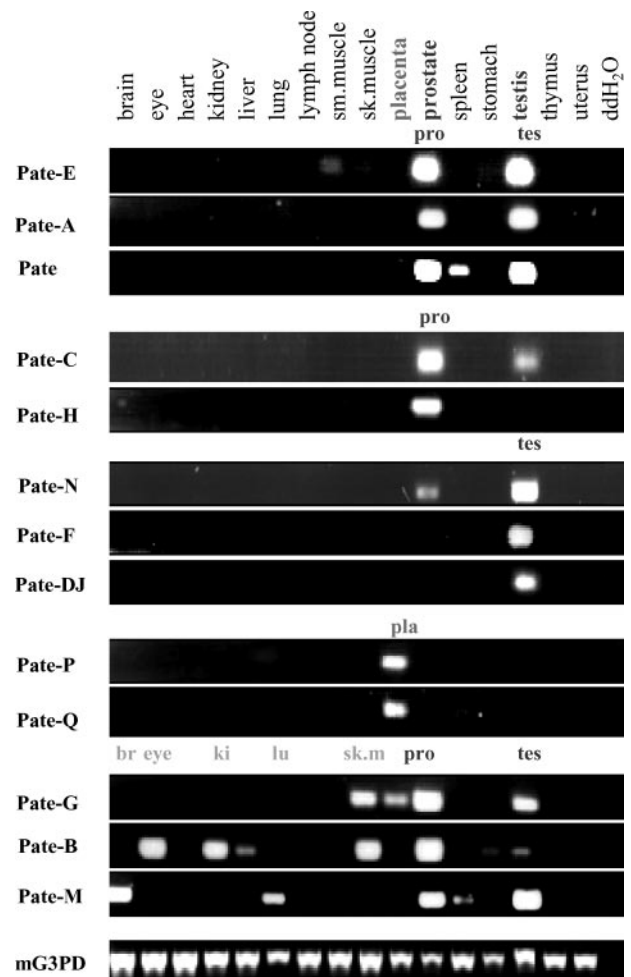


FIGURE 5. RT-PCR expression analyses of mouse *Pate*-like genes. RT-PCR analyses of the mouse *Pate*-like genes were performed with cDNAs (Clontech) obtained from different mouse tissues as indicated. Forward and reverse primers were chosen such that they always spanned an intron, and all RT-PCR products corresponded to the sizes expected of spliced mRNA. Results presented here used forward and reverse primers located in the second and third exons (coding for cysteines 1–5 and cysteines 6–10, respectively); similar results were obtained when the analysis was repeated with forward and reverse primers located in the first and third exons (data not shown). PCR was performed for 35 cycles, and the PCR products were analyzed as described under "Experimental Procedures." The ubiquitously expressed mouse glyceraldehyde-3-phosphate dehydrogenase (*mG3PDH*) served as a control for cDNA integrity.

*Pate-E* (Fig. 6, DP). Dorsal lobe expression remained high, irrespective of castration and subsequent DHT administration (data not shown). In contrast, the *Pate-B* and *Pate-E* genes were not expressed in the uncastrated ventral lobe (Fig. 6, lane 9). Following castration, however, both *Pate-B* and *Pate-E* were clearly expressed in the ventral lobe (Fig. 6, lanes 1–4). DHT administration subsequent to castration ablated this expression (Fig. 6, lanes 5–8). The DHT-mediated suppression of castration-induced ventral lobe expression occurred swiftly (Fig. 6, lane 5); 0.2 h of DHT administration completely extinguished *Pate-E* expression and led to the partial suppression of *Pate-B* expression that was complete following 24 h of DHT treatment. In contrast to the differential dorsal and ventral lobe expression of *Pate-B* and *Pate-E*, the *Pate-H* gene demonstrated high expression in both ventral and dorsal lobes (Fig. 6, lanes 9 and 10, respectively). Neither castration nor subsequent DHT treat-

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ment altered this expression (Fig. 6, lanes 1–8). In fact, the universal expression of the *Pate-H* gene served as a convenient internal control for integrity of prostate cDNAs as did the housekeeping *L19* gene (Fig. 6, lanes 1–10).

**The PATE-B Protein Localizes to the Acrosomal Region of Human Sperm and Is Expressed in Discrete Normal and Malignant Epithelial Prostate Cells**—The discovery of clusters of human and mouse *PATE* (*Pate*)-like genes expressed in reproductive tissues suggested involvement in functions related to reproductive activities. To further understand their function, we generated antibodies directed against the human PATE-B protein. With these anti-PATE-B reagents we investigated the following: (a) whether the PATE-B protein localizes to sperm

cells and (b) whether the PATE-B protein product can be detected in normal and/or malignant prostate tissue.

Incubation with anti-PATE-B antibodies demonstrated intense staining of sperm cells primarily in the acrosomal region (Fig. 7, panel 1). Specificity of staining was confirmed by use of preimmune serum (Fig. 7, panel B1, no staining) addition to the anti-PATE-B antibodies of either a nonspecific protein or competing PATE-B protein (staining retained and absence of staining, Fig. 7, panels C and D, respectively).

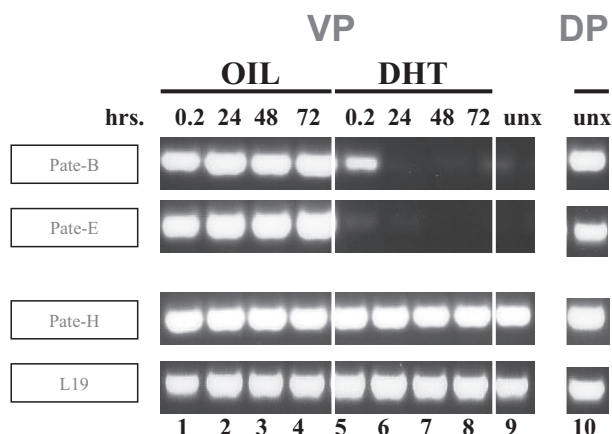
To investigate PATE-B protein expression in the prostate, we performed immunohistochemical analyses of prostatic tissue sections (Fig. 8). Analyzed samples include both normal ducts as well as clusters of malignant epithelial cells that had spread into prostate stroma. Substantial anti-PATE-B immunoreactivity localized in the normal ducts to the cytoplasm of apical epithelial cells (Fig. 8A). Surprisingly, only a discrete subpopulation of apical epithelial cells demonstrated the PATE-B protein (Fig. 8A). Furthermore, PATE-B staining was granular, and the select anti-PATE-B positive cells included distinctive dendritic-like projections.

Malignant epithelial cells forming cell clusters in the prostatic stroma (Fig. 8B) showed a more intense PATE-B immunoreactivity as compared with that seen in the normal, apical epithelial cells (compare Fig. 8, A and B). As in the case of normal prostatic epithelial cells, only a discrete subpopulation of malignant cells demonstrated PATE-B expression (Fig. 8B). Specificity of staining was confirmed by use of preimmune serum and addition of competing specific PATE-B protein, in both of these controls no staining was observed, as expected (data not shown).

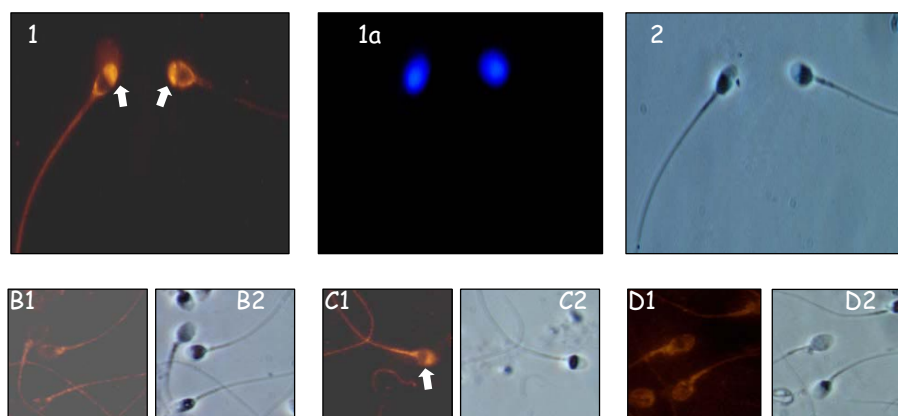
**Presence in Rat and Dog Genomes of Pate-like Genes**—Investigation of the rat syntenic locus revealed a 2.5-Mbp insertion, corresponding to that of the mouse 0.8-Mbp insert, located between the rat *Acrv1* (*sp10*) and *Pate-A* genes. This analysis showed that the mouse *Pate*-like genes *Acrv1*, *Pate-P*, *Pate-Q*, *Pate-F*, *Pate-A*, *Pate-C*, *Pate-E*, *Pate-N*, *Pate*, *Pate-M*, *Pate-DJ*, and *Pate-B* all have rat orthologs (Fig. 9). We could not assign

rat orthologs to the mouse *Pate-G* and *Pate-H* genes. The rat locus includes several *Pate*-like genes that code for proteins that appear rat-specific, including *RSP1* (rat spleen protein 1), *RUP2* (rat urinary protein 2), *RUP3* (rat urinary protein 3) (21), *Suc1g1*, and an additional five rat *Pate*-like genes that could not be assigned a mouse ortholog. Analysis of the dog *Pate*-like gene locus revealed dog (d)*Acrv1*, *dPate*, *dPate-M*, and *dPate-DJ*. Four additional dog *Pate*-like genes located between *dAcrv1* and *dPate* (Fig. 9) could not be clearly assigned to any human, mouse, or rat orthologs.

**Analysis by SDS-PAGE of Recombinant Pate-like Proteins**—To see whether we could synthetically produce recombinant Pate-like pro-



**FIGURE 6. Effect of castration and subsequent DHT administration on Pate-like gene expression in the ventral and dorsal lobes of the mouse prostate.** Mice were castrated and 14 days later injected (subcutaneously) at time 0, 24, and 48 h either with oil or with DHT dissolved in the oil. Mice were sacrificed 12 min and 24, 48, and 72 h, respectively, and the ventral and dorsal prostate lobes were isolated, followed by RNA isolation and cDNA preparation. RT-PCR analyses were performed using forward and reverse primers located in the first and third exons. PCR was performed for 35 cycles, and the PCR products were analyzed as described under "Experimental Procedures." The ubiquitously expressed mouse *L19* gene served as a control for cDNA integrity. *unx*, uncastrated.

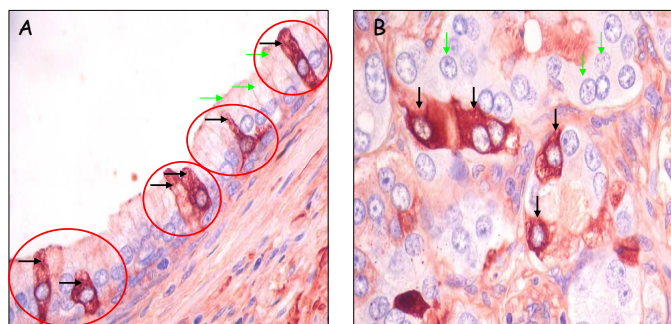


**FIGURE 7. Localization of the PATE-B protein to the acrosomal region of human sperm.** Panels 1, 1a, and 2, human sperm was incubated with anti-PATE-B antisera followed by fluorescently labeled anti-rabbit antibodies. Specific staining was observed in the region of the sperm acrosome (panel 1). Staining with 4',6-diamidino-2-phenylindole demonstrated the sperm nucleus (panel 1a), and Nomarski optics showed the whole sperm (panel 2). Panels B–D, human sperm similarly investigated with preimmune serum (panel B1) or with immune anti-PATE-B antisera plus a nonrelevant protein (panel C1) or competing soluble PATE-B protein (panel D1) attested to the specificity of staining, indicated by the white arrow. Nomarski optics demonstrated the sperm cells (panels B2, C2, and D2).

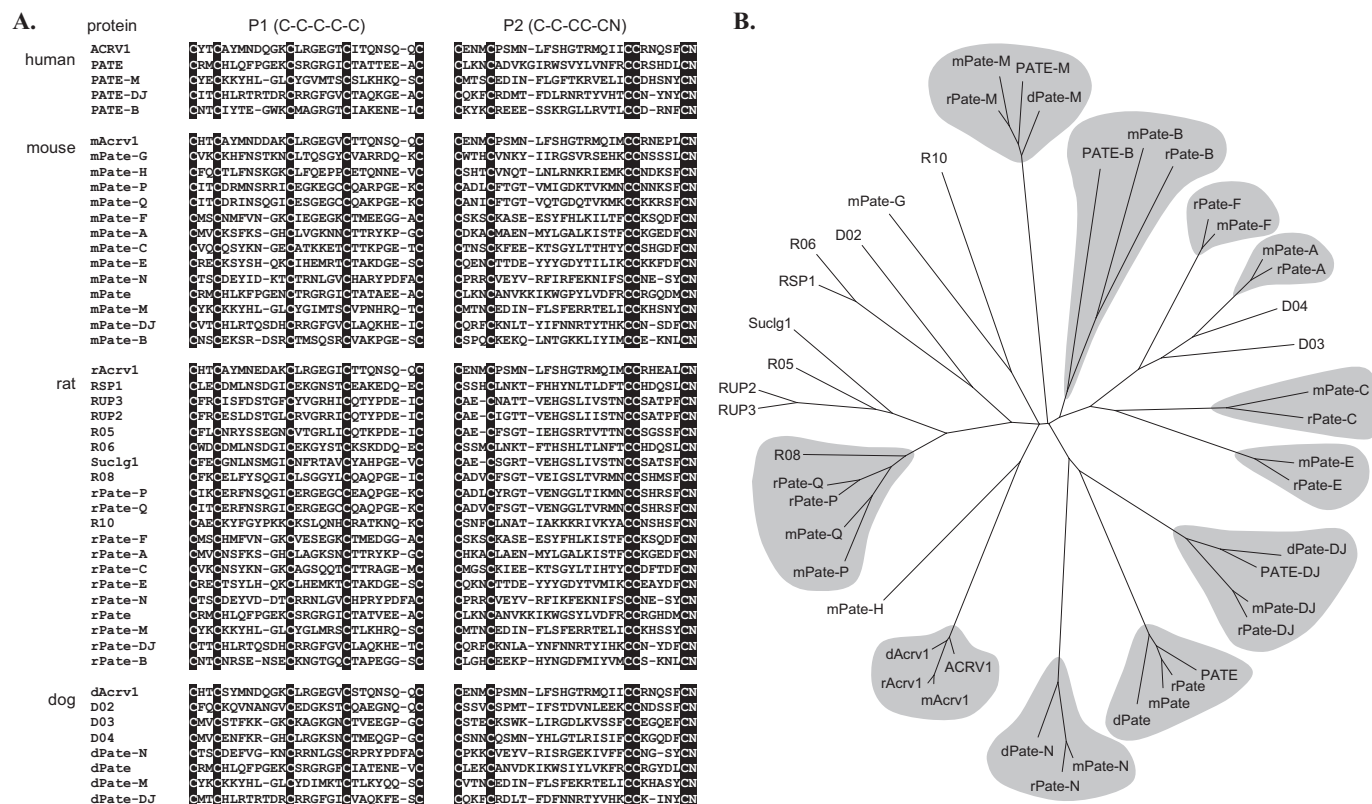
teins in mammalian cells and to investigate whether these proteins undergo post-translational modifications, we expressed mouse Pate, Pate-C, and Pate-P proteins in a mammalian cell expression system. Immunoblot analyses revealed (Fig. 10) that whereas Pate-P protein migrated with its expected mobility

(Fig. 10A, lane 2), the Pate and Pate-C proteins each displayed two bands (Fig. 10A, lanes 1 and 3, respectively). Treatment with PNGase F, resulted in increased mobility of the Pate and Pate-C proteins (Fig. 10B, compare 1st and 2nd, and 3rd and 4th lanes, respectively), demonstrating that both these proteins are modified with N-linked sugars. The Pate protein showed a single consensus N-glycosylation site, NCT, whereas Pate-C harbors an NSC sequence, an alternative N-linked glycosylation site (22).

*Human PATE (Pate)-like Proteins Modulate Activity of nAChRs*—Having established expression of the PATE (Pate)-like genes and their hormonal regulation in reproductive tissues, having identified the homologous clusters of human, mouse, rat, and dog genes, and following the successful synthesis of the PATE (Pate)-like proteins in mammalian cells, we wished to understand what functions they may perform. The striking similarity in molecular makeup between snake toxins that bind to and modulate the activity of nAChRs, the recently discovered mammalian TFP/Ly-6/uPAR-like proteins SLURP1, SLURP2 (23), and Lynx1 all experimentally shown to modulate nAChR activity and the PATE (Pate)-like proteins prompted us to investigate whether these latter proteins possess similar activities. To test for this, we expressed various nAChRs using the *Xenopus* oocyte cell expression system, and we assessed evoked changes in channel activation that occur in response to application of the recombinant PATE (Pate)-like



**FIGURE 8. Human PATE-B protein expression in discrete prostatic apical epithelial cells and in malignant epithelial cells.** A, tissue sections of paraffin-embedded prostate tissue containing both normal and malignant prostate cells were reacted with anti-PATE-B rabbit polyclonal antisera, and antibody binding was detected with polymer-horseradish peroxidase-conjugated anti-rabbit antibodies followed by 3-amino-9-ethylcarbazole chromagen. Staining is seen in isolated cells (black arrows) suggesting that a specific subpopulation of the ductal epithelial cells express PATE-B; other cells in this duct (green arrows indicating selected cells) are barely stained indicating very low PATE-B expression. B, malignant prostate cells from a different region but contained in the section as in A. Discrete malignant cells are very intensely stained (black arrows) indicating high PATE-B expression, whereas other malignant cells (green arrows indicating selected cells) show very little staining.



**FIGURE 9. Multiple sequence alignment of C<sub>10</sub>N motif and a phylogenetic tree of PATE (Pate)-like proteins from human, mouse, rat, and dog.** A, the amino acid sequences of the human, mouse, rat, and dog PATE (Pate)-like proteins are presented. The first (P1) and second (P2) blocks show sequences extending from C1 to C5 and C6 to C10N, respectively. The C<sub>10</sub>N motifs are highlighted. Where possible Pate-like nomenclature has been used for the interspecies orthologs. Otherwise arbitrary designations, such as R05, were used for illustration. Expression of all human and mouse PATE (Pate)-like genes presented have been demonstrated experimentally by RT-PCR analyses. B, same multiple sequence alignment was used to build a phylogenetic tree. The tree is a Neighbor-joining consensus tree based on 1000 replicates. Apparent orthologous groups are marked with a gray background.



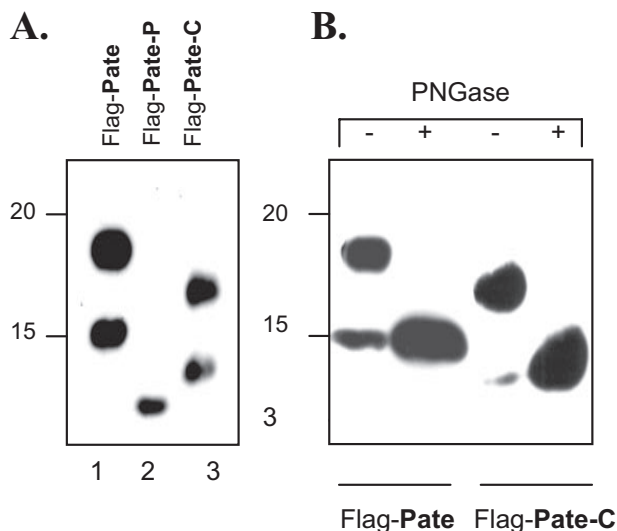
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proteins. In this study we evaluated measurement of net charge accumulation over the entire period of drug administration. This analysis produces results essentially identical to those obtained by more complicated concentration-correction methods. The net charge is a particularly attractive parameter (15) as it represents the time integration of all activated channels

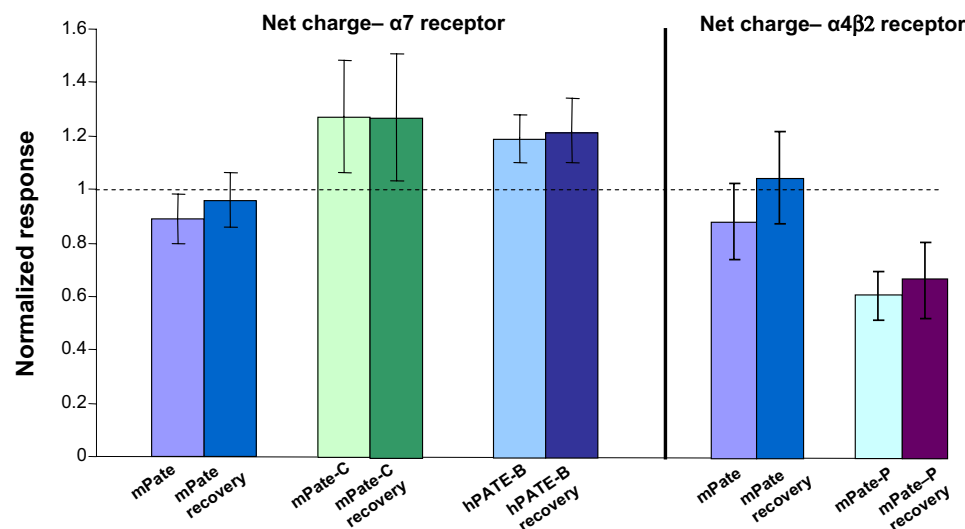
responding to drug administration. This analysis (Fig. 11) showed that application of mouse Pate (mPate) to both  $\alpha 4\beta 2$  and  $\alpha 7$  nAChRs or human PATE-B (hPATE-B) to the  $\alpha 4\beta 2$  nAChR did not evoke any changes in net charge, based on pairwise *t* tests of control ACh-evoked responses recorded prior to PATE application to those responses evoked by the application of ACh in the continued presence of the PATE. In contrast, an increase in the net charge was measured after application of hPATE-B to the  $\alpha 7$  nAChR, a change that was statistically significant ( $p < 0.05$ , based on pairwise *t* test analysis of responses obtained from the same cells in the absence and presence of hPATE-B) and persisted even after a 4-min wash (Fig. 11). Additionally, mPate-C increased the net charge when applied to human  $\alpha 7$  nAChRs, a statistically significant change ( $p < 0.05$  based on pairwise *t* tests) that also persisted after a 4-min wash (Fig. 11). Interestingly, the mPate-P protein did not elicit any changes when applied to the human  $\alpha 7$  nAChRs but did evoke a statistically significant decrease in net charge when added to the  $\alpha 4\beta 2$  nAChR ( $p < 0.05$  based on pairwise *t* tests), a change also persisting after a 4-min wash (Fig. 11). In summary, of the four recombinant PATE (Pate)-like proteins we synthesized and assayed, three (hPATE-B, mPate-C, and mPate-P) showed modulatory activity of the nAChRs.

**Expression of PATE (Pate)-like Proteins in Neuron-rich Tissues**—As shown above certain PATE (Pate)-like proteins clearly modulate the activity of nicotinic AChRs. This matches well with the similarity between the PATE-like protein family and the nAChR neuromodulators, the secreted, mammalian TFP/Ly-6/uPAR SLURP1 and SLURP2 proteins. Functioning as neuromodulators, one would expect that neuron-rich tissues should also express PATE (Pate)-like genes. Consistent with this, analyses of mouse Pate-like gene expression indeed show that mouse Pate-B, in addition to its extensive expression in

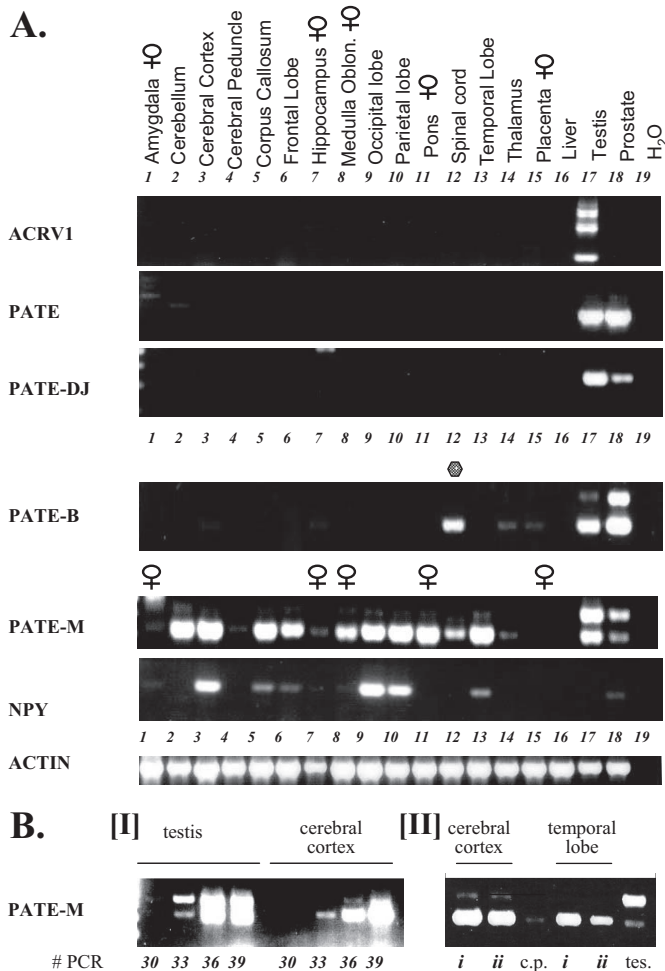
prostate tissue, is strongly expressed in the eye and skeletal muscle both heavily innervated tissues (Fig. 5). Additionally, besides its expression in prostate and testis, mPate-M is extensively expressed in brain (Fig. 5). Viewed in this context, it was surprising to note the lack of human PATE-like gene expression in neuron-rich tissues, such as brain. This could be due to the following: (a) low expression levels in brain requiring increased sensitivity for detection, and/or (b) expression only in discrete regions of the brain such that specific PATE-like cDNAs are diluted out in total brain cDNAs, proscribing their detection. To test these options and to enhance sensitivity, we repeated the RT-PCR analyses with an increased number of PCR cycles (40 instead of 35). This clearly showed low levels of PATE-M expression in the brain (data not shown). A subsequent



**FIGURE 10. Expression of Pate, Pate-P, and Pate-C proteins, N-glycosylation of Pate and Pate-C in HEK293 (human kidney) cells.** A, HEK293 cells were transfected with constructs encoding the proteins that comprised the Pate, Pate-P, or Pate-C sequences, tagged at their N termini with a FLAG epitope, and the secreted Pate-like proteins purified as described under "Experimental Procedures." The purified proteins were analyzed by 11% SDS-PAGE, Western-blotted, and probed with anti-FLAG antibodies (A and B). B, Western blot analysis of FLAG-tagged Pate-like proteins treated with PNGase F. Proteins were treated as described in A, with the difference that the proteins were treated with PNGase F to remove N-linked sugars (lanes indicated by +).



**FIGURE 11. Modulation of the activity of nAChRs by PATE (Pate)-like proteins.** The indicated nAChRs were expressed in *Xenopus* oocytes as described under "Experimental Procedures." After obtaining initial control responses to 8-s applications of ACh, cells were washed for 4 min with either control Ringer or Ringer's plus the PATE-like proteins (60 or 200 nM) and then challenged with ACh or ACh plus the PATE-like proteins, respectively. Responses to ACh plus the PATE-like proteins were normalized to the net charge responses to ACh alone for each oocyte. ACh concentrations were 30  $\mu\text{M}$  for  $\alpha 4\beta 2$ -expressing cells and 60  $\mu\text{M}$  for  $\alpha 7$ -expressing cells. To test for recovery, oocytes were then washed for an additional 4 min in the absence of acetylcholine and the PATE-like proteins, and then acetylcholine (30 or 60  $\mu\text{M}$ ) was applied for 8 s and the net charge calculated to give the recovery values.



**FIGURE 12. Expression of human PATE-like genes in different regions of the brain.** *A, upper panel*, cDNAs prepared from different individual regions of the brain as indicated, were subjected to RT-PCR analyses. Expression analysis of the human PATE-like genes was performed with forward and reverse primers that always spanned an intron and the observed RT-PCR product at all times corresponded to the size expected of a spliced mRNA. For the *ACRV1*, *PATE*, *PATE-M*, *PATE-DJ*, and *PATE-B* genes, the forward and reverse primers were located in the first and third exons (coding for the signal peptide and cysteines 6–10, respectively). In addition, cDNAs derived from testes and prostate on the one hand and liver and placenta on the other hand were also analyzed for expression of the PATE-like genes and served as positive and negative controls, respectively. PCR was performed for 35 cycles. Control for cDNA integrity was provided by the actin primers, as indicated, and expression analysis of the brain neuropeptide NPY served as a positive control for expression of a brain neuropeptide in specific regions of the brain. *B, panel I*, to assess the relative PATE-M expression levels in testis and cerebral cortex, a semiquantitative RT-PCR analysis was performed wherein samples were taken following 30, 33, 36, and 39 PCR cycles and subjected to agarose gel analysis. The fact that there are two differently sized PATE-M isoforms, each possessing a distinct duplex melting temperature, precluded real time PCR analysis for this gene. *Panel II*, to assess the generality of PATE-M gene expression in the brain, cDNA samples deriving from cerebral cortex and temporal lobe isolated from different individuals (*i* and *ii*) were subjected to PATE-M RT-PCR analysis. cDNAs prepared from testis (*tes*) and cerebral peduncle (*c.p.*) served as positive and negative controls, respectively.

analysis of cDNA prepared not from total brain but from different individual regions of brain unambiguously resolved this issue (Fig. 12). Whereas PATE and PATE-DJ were not expressed in any of the tested regions, PATE-B was expressed in spinal cord tissue. Particularly striking was the extensive expression of PATE-M. This was seen in the cerebellum, cerebral cortex, corpus callosum, occipital, parietal and temporal

lobes, and pons (Fig. 12, lanes 2, 3, 5, 9, 10, 13, and 11, respectively). Lesser expression levels were observed in the frontal lobe and medulla oblongata with low levels in the spinal cord (Fig. 12, lanes 6, 8, and 12). PATE-M was not expressed in the amygdala, cerebral peduncle, hippocampus, and thalamus (Fig. 12, lanes 1, 4, 7, and 12). Remarkably, PATE-M expression in the various central nervous system locations was exclusively restricted to the exon 2-deleted PATE-M isoform. This stands in contrast to the predominant expression of the exons 1, 2, and 3 isoform in the testis (Fig. 12*B, panel I, lower panel*, compare lanes labeled *testis* and *cerebral cortex*). Brain samples derived from female donors also clearly expressed PATE-M (Fig. 12*A, lanes 8 and 11*), indicating that its expression in neuronal tissues is not gender-specific.

A semi-quantitative analysis comparing testis with cerebral cortex showed comparable levels of PATE-M expression in both tissues (Fig. 12*B*). This is significant because it means that the level of PATE-M expression in the cerebral cortex is at least equivalent to that seen in testis which, of all non-central nervous system tissues examined (Fig. 2) is by far the highest PATE-M expressing tissue.

**DISCUSSION**

We report here syntenic human and mouse genomic loci that contain clusters of genes coding for secreted proteins each comprising a typical TFP/Ly-6/uPAR domain. Their unique expression patterns both in humans and mice provide compelling evidence for critical roles of the PATE (Pate)-like proteins in activities related not only to fertility and reproduction but also to neuronal activity.

The mouse and human PATE (Pate)-like genes all code for proteins that harbor N-terminal sequences conforming to signal peptides that direct protein for secretion. This is consistent with the functionally demonstrated secretion of ACRV1 (SP10) (18), PATE (17), and Pate-B (svs7, caltrin (19)). No other hydrophobic regions corresponding either to transmembrane domains or to signals for addition of GPI tails are present in the PATE (Pate)-like proteins.

Similar to the PATE-like gene locus, a cluster of human Ly-6 orthologous genes resides on chromosome 8q24, and each of these genes codes for a protein that includes the distinctive 10-cysteine pattern seen in the PATE (Pate)-like proteins. With the exception of the secreted proteins SLURP-1 (8) and SLURP-2 (9), this 8q24 locus codes only for GPI-linked, membrane-tethered proteins. In contrast, all active genes residing in both the human (11q24) and mouse (9qA4) PATE (Pate)-like loci described here, code exclusively for secreted TFP/Ly-6/uPAR proteins. They are thus similar to the secreted SLURP-1 and SLURP-2 proteins and to the extensive group of secreted snake and frog toxins bearing the TFP/Ly-6/uPAR domain.

As such, the PATE (Pate)-like proteins are likely to be signaling molecules that bind to target cells, thereby modulating their activity. Indeed, SLURP-1 and SLURP-2, both secreted TFP/Ly-6/uPAR-like proteins similar in structure to the PATE-(Pate)-like proteins, bind to and modulate the activity of nicotinic acetylcholine receptors (10, 23) present on the membrane of target cells. Just as these TFP/Ly-6/uPAR-like proteins modulate nAChR activity, we now demonstrate that in fact the

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PATE (Pate)-like proteins possess similar activities and also modulate the activity of nAChRs.

Our investigations demonstrate that all human PATE-like genes are expressed in male reproductive tissues such as prostate and testis. This does not mean that the PATE (Pate)-like genes function solely in male-restricted reproductive activities. Indeed the mouse *Pate-Q* and *Pate-P* genes show very selective expression in placenta, an exclusively female-specific tissue. Furthermore, expression of a discrete group of Pate-like genes in the mammary glands of pregnant and/or lactating mice (data not shown) also suggests that the Pate-like proteins are likely to function in reproductive activities that are not exclusively male-limited.

Hormonal governance of expression is sustained by the unambiguous castration-induced expression in the ventral prostate of *Pate-B* and *Pate-E*, both of which undergo swift down-regulation following DHT administration. Additionally, expression of certain Pate-like genes was observed only in mammary glands from pregnant or lactating mice and not from virgin mice (data not shown). Taken as a whole these results clearly implicate hormonal participation in the regulation of Pate-like gene expression.

Immunostaining with antibodies demonstrated human PATE-B protein localizing at the sperm acrosomal region. The secreted TFP/Ly-6/uPAR domain-containing proteins Pate-B, PATE, and ACRV1 (*Acrv1*), all bind to sites on the sperm cell (17, 19, 24), suggesting specific receptors for these proteins present on the sperm membrane. These results also tie in with what is known about the mouse Pate-B homolog (*caltrin* or *svs7*), which has been postulated to modulate calcium permeability in sperm cells (20). It is thus not unreasonable to assume that specific receptors for the PATE-B protein reside on human sperm.

Our analyses also showed that the human PATE-B protein localized to both normal and malignant prostatic epithelial cells. Both their morphology and the fact that only discrete, ductal, apical cells stained positive suggest that these PATE-B-positive cells may in fact represent neuroendocrine cells, a specialized subset of epithelial cells. Prostatic neuroendocrine cells (i) are intraglandular and intraductal hybrid epithelial/neuroendocrine cells, (ii) are generally widely scattered throughout the prostate with only an occasional cell per gland/duct, and (iii) express and secrete serotonin and numerous neuropeptides. Both afferent and efferent nerves innervate prostatic neuroendocrine cells. That the PATE-B positive cells are of the neuroendocrine cell type is especially appealing in light of our findings on the neuromodulatory function of the PATE-like proteins (below).

It was also striking that malignant prostatic epithelial cells showed PATE-B expression. As seen with normal epithelium, only certain cancer cells stained PATE-B-positive. This expression was more intense than that seen in the normal cell counterparts. Further investigations will be required to elucidate whether PATE-B expression in these malignant prostate cells actually contributes to their transformed phenotype. Whatever the case may be, localization of PATE-B protein in both sperm and prostatic cells underscores its likely participation in reproductive functions.

All PATE (Pate)-like proteins probably adopt a “three-fingered” fold similar to that of the snake neuro- and cytotoxins (25, 26). There are extensively documented, highly specific interactions between the TFP/Ly-6/uPAR domain-containing snake toxins and the nAChR (5), a ligand-gated five-subunit ion channel. Interestingly, mammalian *Lynx1*, a GPI-anchored TFP/Ly-6/uPAR domain-containing protein, modulates cellular calcium permeability by interacting with neuronal nicotinic acetylcholine receptors (27). More pertinent to the secreted PATE (Pate)-like proteins, the secreted SLURP-1 protein potentiates the human  $\alpha 7$  nicotinic AChRs (10). Thus there is not only structural likeness between snake neurotoxins and the mammalian TFP/Ly-6/uPAR domain-containing proteins SLURP-1 and *Lynx1* but also a functional similarity. Experiments designed to assess whether PATE (Pate)-like proteins possess similar functions, demonstrated that certain PATE (Pate)-like proteins do in fact modulate the activity of nicotinic acetylcholine receptors. Three of the four recombinant PATE (Pate)-like proteins we synthesized and assayed modulated the activity of nAChRs. Two (hPATE-B and mPate-C) up-regulated the homomeric  $\alpha 7$  nAChR and one (mPate-P) down-regulated the  $\alpha 4\beta 2$  heteromeric nAChR. Because of the multitude of the PATE (Pate)-like proteins, to date only four have been investigated for their neuromodulatory activity. We anticipate that additional PATE (Pate)-like proteins also possess neuromodulatory activity, and experiments are in progress to check for this.

The diversity of nAChRs engendered by the combinatorial assembly of various nicotinic subunits ( $\alpha 1$ - $\alpha 10$ ,  $\beta 1$ - $\beta 4$ ,  $\delta$ ,  $\gamma$ , and  $\epsilon$ ) indicates different electrical and binding properties for each combinatorial subtype (28). One would thus naturally expect that modulations of their activity would necessitate a diversity of interacting modulators. For the diversity that we have observed for the PATE (Pate)-like proteins, wherein each particular protein interacts with and modulates in a different manner, specific nAChRs may provide this multiplicity of interacting modulators. Support for such a scenario is provided by our findings with human PATE-B, which on the one hand up-regulates activity of a homomeric  $\alpha 7$  nAChR but is without effect on an  $\alpha 4\beta 2$  combination. In a similar manner, Pate-P down-regulates the  $\alpha 4\beta 2$  configuration but does not affect a homomeric  $\alpha 7$  nAChR. Gross nAChR inactivation as mediated by snake toxins could not be tolerated by the organism, and one would thus further predict that naturally occurring endogenous proteins modulating nAChR activity will do so only moderately. That the PATE (Pate)-like proteins mediate modest, yet statistically significant, alterations in nAChR activity fits in well with this postulated fine-tuning.

Taken together our data are consistent with a neuromodulatory function for the PATE (Pate)-like proteins. This firmly places them in the family of the neuromodulatory TFP/Ly-6/uPAR domain-containing proteins SLURP-1 and *Lynx1*.

How are the neuromodulatory functions of the PATE (Pate)-like proteins to be reconciled with their extensive expression in reproductive tissues? It is well known that neurons and sperm share many common “neuron-specific” receptors. For example, the nAChR subunits  $\alpha 7$ ,  $\alpha 9$ ,  $\alpha 3$ ,  $\alpha 5$ , and  $\beta 4$  have all been demonstrated to be present in human sperm, and the  $\alpha 7$  subunit

likely exists as a homomer in the posterior post-acrosomal and neck regions of sperm (29, 30). Furthermore, the human acrosome reaction initiated by acetylcholine or recombinant egg zona pellucida protein ZP3 involves the  $\alpha 7$  nAChR (31, 32), and it has been proposed that the sperm nAChRs may be key regulators of signaling pathways important to the acrosome reaction and sperm motility (29, 33). This nAChR commonality in both sperm and neurons has even led to the classification of a sperm cell as “a neuron with a tail” (34). Although provocative, this definition does attest to the unequivocal participation of what was previously thought to be neuron-specific receptors and ligands in the functioning of cells pertaining to the reproductive system. This common functionality of nAChRs in both reproductive and neuronal tissues thus reconciles the neuromodulatory activity of the PATE (Pate)-like proteins and rationalizes their expression in both neuronal and reproductive tissues.

In summary, we have identified a new family of secreted TFP/Ly-6/uPAR domain-containing proteins, designated the PATE (Pate)-like proteins, that likely play important roles in reproductive and neuronal physiology. Further clarification of their normal functions will likely contribute to new therapeutics for pathologies of both the central nervous system and the reproductive system.

## REFERENCES

- Bera, T. K., Maitra, R., Iavarone, C., Salvatore, G., Kumar, V., Vincent, J. J., Sathyanarayana, B. K., Duray, P., Lee, B. K., and Pastan, I. (2002) *Proc. Natl. Acad. Sci. U. S. A.* **99**, 3058–3063
- Fry, B. G., Wuster, W., Kini, R. M., Brusic, V., Khan, A., Venkataraman, D., and Rooney, A. P. (2003) *J. Mol. Evol.* **57**, 110–129
- Ploug, M., and Ellis, V. (1994) *FEBS Lett.* **349**, 163–168
- Ploug, M. (2003) *Curr. Pharm. Des.* **9**, 1499–1528
- Tsetlin, V. I., and Hucho, F. (2004) *FEBS Lett.* **557**, 9–13
- Tremeau, O., Lemaire, C., Drevet, P., Pinkasfeld, S., Ducancel, F., Boulain, J. C., and Menez, A. (1995) *J. Biol. Chem.* **270**, 9362–9369
- Kolbe, H. V., Huber, A., Cordier, P., Rasmussen, U. B., Bouchon, B., Jaquinod, M., Vlasak, R., Delot, E. C., and Kreil, G. (1993) *J. Biol. Chem.* **268**, 16458–16464
- Adermann, K., Wattler, F., Wattler, S., Heine, G., Meyer, M., Forssmann, W. G., and Nehls, M. (1999) *Protein Sci.* **8**, 810–819
- Tsuji, H., Okamoto, K., Matsuzaka, Y., Iizuka, H., Tamiya, G., and Inoko, H. (2003) *Genomics* **81**, 26–33
- Chimienti, F., Hogg, R. C., Plantard, L., Lehmann, C., Brakch, N., Fischer, J., Huber, M., Bertrand, D., and Hohl, D. (2003) *Hum. Mol. Genet.* **12**, 3017–3024
- Freemerman, A. J., Flickinger, C. J., and Herr, J. C. (1995) *Mol. Reprod. Dev.* **41**, 100–108
- Kumar, S., Tamura, K., and Nei, M. (2004) *Brief. Bioinform.* **5**, 150–163
- Notredame, C., Higgins, D. G., and Heringa, J. (2000) *J. Mol. Biol.* **302**, 205–217
- Stokes, C., Papke, J. K. P., McCormack, T., Kem, W. R., Horenstein, N. A., and Papke, R. L. (2004) *Mol. Pharmacol.* **66**, 14–24
- Papke, R. L., and Porter Papke, J. K. (2002) *Br. J. Pharmacol.* **137**, 49–61
- Levitin, F., Stern, O., Weiss, M., Gil-Henn, C., Ziv, R., Prokocimer, Z., Smorodinsky, N. I., Rubinstein, D. B., and Wreschner, D. H. (2005) *J. Biol. Chem.* **280**, 33374–33386
- Soler-Garcia, A. A., Maitra, R., Kumar, V., Ise, T., Nagata, S., Beers, R., Bera, T. K., and Pastan, I. (2005) *Reproduction (Camb.)* **129**, 515–524
- Herr, J. C., Wright, R. M., John, E., Foster, J., Kays, T., and Flickinger, C. J. (1990) *Biol. Reprod.* **42**, 377–382
- Luo, C. W., Lin, H. J., and Chen, Y. H. (2001) *J. Biol. Chem.* **276**, 6913–6921
- Coronel, C. E., Winnica, D. E., Novella, M. L., and Lardy, H. A. (1992) *J. Biol. Chem.* **267**, 20909–20915
- Southan, C., Cutler, P., Birrell, H., Connell, J., Fantom, K. G., Sims, M., Shaikh, N., and Schneider, K. (2002) *Proteomics* **2**, 187–196
- Satomi, Y., Shimonishi, Y., and Takao, T. (2004) *FEBS Lett.* **576**, 51–56
- Arredondo, J., Chernyavsky, A. I., Jolkovsky, D. L., Webber, R. J., and Grando, S. A. (2006) *J. Cell. Physiol.* **208**, 238–245
- Foster, J. A., Klotz, K. L., Flickinger, C. J., Thomas, T. S., Wright, R. M., Castillo, J. R., and Herr, J. C. (1994) *Biol. Reprod.* **51**, 1222–1231
- Menez, A. (1998) *Toxicol.* **36**, 1557–1572
- Tsetlin, V. (1999) *Eur. J. Biochem.* **264**, 281–286
- Ibanez-Tallon, I., Miwa, J. M., Wang, H. L., Adams, N. C., Crabtree, G. W., Sine, S. M., and Heintz, N. (2002) *Neuron* **33**, 893–903
- Nirthanan, S., and Gwee, M. C. (2004) *J. Pharmacol. Sci.* **94**, 1–17
- Bray, C., Son, J. H., Kumar, P., and Meizel, S. (2005) *Biol. Reprod.* **73**, 807–814
- Kumar, P., and Meizel, S. (2005) *J. Biol. Chem.* **280**, 25928–25935
- Bray, C., Son, J. H., and Meizel, S. (2002) *Biol. Reprod.* **67**, 782–788
- Son, J. H., and Meizel, S. (2003) *Biol. Reprod.* **68**, 1348–1353
- Bray, C., Son, J. H., and Meizel, S. (2005) *Mol. Hum. Reprod.* **11**, 881–889
- Meizel, S. (2004) *Biol. Rev. Camb. Philos. Soc.* **79**, 713–732

***PATE* Gene Clusters Code for Multiple, Secreted TFP/Ly-6/uPAR Proteins That Are Expressed in Reproductive and Neuron-rich Tissues and Possess Neuromodulatory Activity**

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