# Rescue Effect of Exercise on Impaired Arteriolar Myogenic Response with Advancing Age

Kwang-Seok Hong<sup>1</sup>, Man-Gyoon Lee<sup>2</sup>

<sup>1</sup>School of Medicine, University of Virginia, Charlottesville, United States; <sup>2</sup>Graduate School of Physical Education, Kyung Hee University, Yongin, Korea

**PURPOSE**: Mechanosensitive vascular smooth muscle cells (VSMCs) of resistance arteries crucially regulate blood flow by constricting or dilating over fluctuation of blood pressure (the myogenic response). This review was aimed at introducing cellular signaling that is relevant to arterial myogenic response and briefly describing how the arterial autoregulation is impaired by aging and exercise intervention restores the diminished myogenic responsiveness.

METHODS: A systemic literature research was conducted through PUBMED to comprehend previous studies that explore molecular mechanisms underlying arterial myogenic response, impaired pressure-induced vasoconstriction with advancing age, and effect of exercise training on the arterial autoregulation.

RESULTS: The myogenic response generally consists of three steps: 1) detection of mechanical stress (e.g. stretch, tension) exerted on VSMCs, 2) biological transduction pathways (e.g., depolarization, Ca2+ entry, phosphorylation of myosin light chain, Ca2+ sensitization, actin polymerization), and 3) adjustment of vascular tone (e.g. vasoconstriction or dilation). Aging induces vascular aging that is coupled to increased risks of development of cardiovascular diseases. The intrinsic ability of VSMCs to maintain appropriate blood flow in response to changes in intravascular pressure has been reported to be impaired with advancing age. In contrast, exercise intervention has been demonstrated to rescue aging-induced attenuation of arterial myogenic responsiveness.

**CONCLUSIONS:** Abnormal myogenic response of resistance arteries leads to vascular rupture, vasospasms, hypertension, or hypotension. Therefore, it will be valuable to investigate the exact mechanisms underlying the contribution of exercise training to arterial myogenic response to prevent and treat impaired arterial autoregulation-induced cardiovascular disorders.

Key words: Aging, Myogenic reactivity, Pressure-induced vasoconstriction, Protein kinase C, Vascular smooth muscle cells

#### **INTRODUCTION**

Elevated intravascular pressure increases the radius of blood vessels, which leads to considerable increase in blood flow. Contrarily, decreased intraluminal pressure results in insufficient blood perfusion. As the transitions in intraluminal pressure and subsequent surplus/lack of blood flow cause vascular rupture, edema, or ischemia, small arteries or arterioles (generally  $<\!200~\mu m$  of internal diameter) exhibit an intriguing autoregulatory mechanism in response to change in intraluminal pressure to regulate local blood perfusion, minimize capillary hydrostatic pressure, and modulate peripheral vascular resistance [1]. This arterial auto-

regulation, called the 'myogenic response', is defined as intrinsic vascular behavior which elicits vasoconstriction or vasodilation when intraluminal pressure increases or decreases, respectively [2]. The myogenic response is referred to as the inherent properties of vascular smooth muscle cells (VSMCs) since myogenic responsiveness is independent of endothelial cells (ECs) or neurohumoral modulation [1,3]. Abnormal regulation of the myogenic response has been observed in various cardiovascular or metabolic disorders such as subarachnoid hemorrhage, diabetes, congestive heart failure [4]. In this context, as impaired myogenic responsiveness is closely linked to cardiovascular diseases (e.g., hypertension, ischemic stroke, vasospasm), the intrinsic autoregulation has been paid

Corresponding author: Man-Gyoon Lee Tel +82-31-201-3753 Fax +82-31-201-3743 E-mail mlee@khu.ac.kr
Received 4 Jan 2017 Revised 18 Jan 2017 Accepted 13 Feb 2017

@ This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.



attention as drug targets.

A major culprit of morbidity and mortality in older individuals is largely related to cardiovascular diseases [5]. Advancing age has been demonstrated to cause functional and structural alteration in vascular beds including vascular wall remodeling or excessive vascular stiffness [6]. It has been established along with aging that increased reactive oxygen species (ROS) in the VSMCs and ECs lead to impairment of nitric oxide signaling, increased inflammatory responses, up- or down-regulation of transcriptional factors regulating VSMC proliferation [7]. In contrast to advancing age, physical activity has a plethora of beneficial effects on the cardiovascular system [8]. It has been investigated that exercise training markedly reduces coronary artery diseases, hypertension, and heart failure-mediated morbidity and mortality [9,10]. Thus, exercise intervention may act as a primary or secondary prevention and treatment for cardiovascular diseases. In regard to the arterial autoregulaton that is crucial for the regulation of local blood flow, specific mechanisms by which myogenic responsiveness is impaired with advancing age have not yet fully defined. In addition, whether and how regular physical activity restores the impaired myogenic reactivity in elderly individuals still remains poorly understood. This review has focused on briefly describing intracellular signaling pathways for the myogenic response and how the autoregulation is affected by aging and exercise training.

## **MOLECULAR MECHANISMS UNDERLYING THE MYOGENIC RESPONSE**

For over 100 years, numerous studies in vascular biology have continually sought to investigate intracellular signaling for the myogenic response. Despite these efforts, how the intriguing myogenic responsiveness operates in physiological or pathological circumstances is still unclear. As far as is known, myogenic reactivity is comprised of several processes as follows: mechanosensitive ion channels, receptors, extracellular proteins, and cytoskeletal proteins sense mechanical stresses on vascular wall following changes in intraluminal pressure. The detected mechanical forces are converted to biological signals (i.e., mechanotransduction) such as alteration in membrane potential, Ca2+ influx, myosin light chain phosphorylation-dependent vasoconstriction, Ca<sup>2+</sup> sensitization, or cytoskeletal rearrangement for myogenic vasoconstriction [1,4,11].

Increased intraluminal pressure leads the sensory machineries of resistance arteries or arterioles to detect tension or stretch of the VSMCs and then initiate intracellular signaling events for pressure-induced vasoconstriction. Accordingly, the identification of mechanosensors in the VSMCs has been drawing keen attention of leading investigators. To date, integrins and G protein-coupled receptors (GPCRs) have been reported to contribute to detection of change in intraluminal pressure and in turn evoke the myogenic response. Integrins consisting of a non-covalent interaction of  $\alpha$ - and  $\beta$ -subunit heterodimers are involved in divergent vasomotor reactivity by regulating Ca2+ dynamics (e.g., extracellular Ca<sup>2+</sup> influx) [12]. In addition to the role of integrins in vascular reactivity, it has been elucidated that inhibition of  $\alpha_v \beta_3$  or  $\alpha_5 \beta_1$  integrin using specific antibodies abolishes myogenic vasoconstriction in skeletal muscle arterioles [13]. Further, more recently, activation of  $\alpha_{\nu}\beta_{3}$  integrin has been delineated to regulate pressure-induced vasoconstriction in cerebral arteries by modulating intracellular Ca<sup>2+</sup> waves [14]. Next, since the novel finding that membrane stretch evokes conformational changes in angiotensin II type 1 receptor (AT<sub>1</sub>R) and provoke downstream signaling in the absence of its ligand, angiotensin II [15], there is growing evidence that deformation of vascular wall following increased intraluminal pressure causes ligand-independent activation of GPCRs in the VSMCs. Specifically, numerous studies have been undertaken to demonstrate using pharmacological inhibition or genetic manipulation (e.g., knockout or knockdown of target GPCRs) that mechanoactivation of purinergic receptors [16], cysteinyl leukotriene 1 receptors [17], and AT<sub>1</sub>R [18,19] takes part in myogenic vasoconstriction of mesenteric, cerebral, and skeletal muscle arterioles.

Membrane depolarization following an acute elevation in intravascular pressure is a major determinant for myogenic responsiveness of small arteries and arterioles. Transient receptor potential (TRP) channels have been suggested to regulate membrane potential and Ca<sup>2+</sup> signaling [20]. Sub-families of TRP channels are comprised of canonical (TRPC), melastatin (TRPM), polycystin (TRPP), and vanilloid (TRPV), akyrin (TRPA), and musolipin (TRPML) channels. Based on biophysical properties of the channels, cations (e.g., Ca<sup>2+</sup>, K<sup>+</sup>, Na<sup>+</sup>) are selectively or non-selectively allowed to be permeable for alteration in membrane potential [20]. With respect to myogenic vasoconstriction, blockade of TRPC6 channel by antisense oligodeoxynucleotides results in significant reductions in membrane potential and myogenic reactivity in pressurized cerebral arteries [21]. Activation of TRPM4 channels being permeable to monovalent cations (e.g., Na+) and regulated by intracellular Ca2+ level and protein kinase C (PKC) has also been identified to induce membrane depolarization and contribute to myogenic reactivity of cerebral arteries [22]. Thus, it is suggested that TRP channels play a critical role in membrane poten-



tial regulation and pressure-induced vasoconstriction. Further, epithelial Na<sup>+</sup> channel (ENaC) has been thought to be a mechano-gated channels in the VSMCs [23,24]. It is implicated that pressure-induced stretch of the VSMCs may be sensed by extracellular components of ENaC and then the detection may cause the opening of pore-forming components of ENaC [24]. Na+ entry through ENaC is coupled to membrane depolarization and subsequent Ca<sup>2+</sup> influx that is required for myogenic responsiveness. Indeed, it has been found that amiloride/benzamil-dependent pharmacological inhibition of ENaC profoundly abolishes myogenic vasoconstriction [25,26].

In contrast to the role of cation entry through TRP channels and ENaC in membrane depolarization following distension of the VSMCs, K<sup>+</sup> channels serve as negative feedback regulators that prevent exaggerated myogenic reactivity in resistance arteries. Specifically, the large (big) conductance Ca<sup>2+</sup>-activated K<sup>+</sup> (BK<sub>Ca</sub>) channels and voltage-dependent K<sup>+</sup> (K<sub>v</sub>) channels participate in modulation of pressure-induced vasoconstriction through K<sup>+</sup> efflux-mediated decrease in membrane potential of the VSMCs. Stimulation of  $BK_{Ca}$  channels by  $Ca^{2+}$  sparks (a vigorous local  $Ca^{2+}$  increase by up to 1-100 µM) exerts spontaneous transient outward currents and in turn hyperpolarization-mediated vasodilation [27]. It has been revealed that increased intraluminal pressure augments the frequency of Ca2+ sparks and knockdown of subunits of BKca channels markedly increase myogenic responsiveness [28,29]. Along with BK<sub>Ca</sub> channels, K<sub>v</sub> channels negatively regulate pressure-induced vasoconstriction. Isolated mesenteric or cerebral arteries treated with selective inhibitors of K<sub>v</sub>1, K<sub>v</sub>2, or K<sub>v</sub>7 channel have shown a significant myogenic vasoconstriction [30,31].

As described above, the activation of diverse ion channels evokes membrane depolarization or hyperpolarization of the VSMCs once intraluminal pressure is altered. Pressure-mediated membrane depolarization has been identified to stimulate voltage-operated Ca<sup>2+</sup> channels (VOCCs) [1]. Moreover, inositol trisphosphate (IP<sub>3</sub>) generated from activation of GPCRs and ryanodine activate IP<sub>3</sub> receptors (IP<sub>3</sub>Rs) and ryanodine receptors (RyRs) in sarcoplasmic reticulum (SR). These events cause Ca<sup>2+</sup> release from the SR by activating IP<sub>3</sub>Rs and RyRs [32]. It has been welldefined that interaction of intracellular Ca2+ with calmodulin (i.e., Ca2+ -calmodulin complex) provokes activation of myosin light chain kinase (MLCK) and MLCK-induced phosphorylation of 20 kDa myosin light chain (LC-20) leads to VSMC contraction by stimulating the MgATPase of actomyosin cross-bridge [33,34]. Thus, Ca<sup>2+</sup> influx through VOCCs and Ca<sup>2+</sup> release from the SR are key determinants for myogenic vasoconstriction.

However, intracellular Ca<sup>2+</sup> signaling in the VSMCs may not be sufficient to account for pressure-induced vasoconstriction since a slight elevation in intracellular Ca2+ has been found in response to increase in intraluminal pressure [35]. Ca<sup>2+</sup> sensitization, one of Ca<sup>2+</sup>-independent mechanisms underlying the myogenic response, is defined as increase or maintenance of vascular contractility in the absence of increased intracellular Ca<sup>2+</sup> in the VSMCs. This interesting phenomenon is associated with regulation of myosin light chain phosphatase (MLCP) suppressing the phosphorylation of LC-20 that is essential for VSMC contraction. Myosin phosphatase targeting subunit 1 (MYPT1) and 17 kDa protein kinase C-potentiated inhibitory protein (CPI-17) are phosphorylated by Rhoassociated kinase (ROCK) and PKC, respectively [36,37]. The phosphorylation of those subunits attenuates MLCP activity, which enhances MLCK activity and augments or maintain vascular contractility without a significant increase in Ca<sup>2+</sup> level. Suppression of phosphorylation of MLCP considerably reduces myogenic vasoconstriction in cerebral and skeletal muscle arterioles [38]. Next, as for pressure-induced cytoskeletal reorganization, dynamics of actin thin filament has been focused as another Ca<sup>2+</sup>-independent mechanism underlying the myogenic response. Contractile  $\alpha$ -actin filaments have been known to anchor to focal adhesion complexes under integrins embedded in the plasma membrane [39,40]. Actin cytoskeleton reorganization (i.e. transition from globular  $\alpha$ -actin to filamentous  $\alpha$ -actin) following alteration in intraluminal pressure is obligatory for myogenic reactivity as globular  $\alpha$ -actin level is found to decrease along with increase in intraluminal pressure and actin polymerization inhibitors (e.g., cytochalasins, latrunculin) significantly diminish pressure-induced vasoconstriction in cerebral or skeletal muscle arterioles [18,41].

Collectively, the myogenic autoregulation of small arteries and arterioles for the regulation of local flood flow and peripheral resistance is an intriguing outcome of integrity of diverse signaling pathways (Fig. 1).

## AGING-MEDIATED ATTENUATION IN THE **MYOGENIC RESPONSE**

Appropriate blood supply to skeletal muscle is required to perform daily activities and sustained muscle contraction [42]. Impaired physical performance with advancing age may be accompanied by dysregulation of local blood flow and inadequate oxygen supply [42]. Skeletal muscle in old rats has shown a reduction in blood flow capacity following elec-



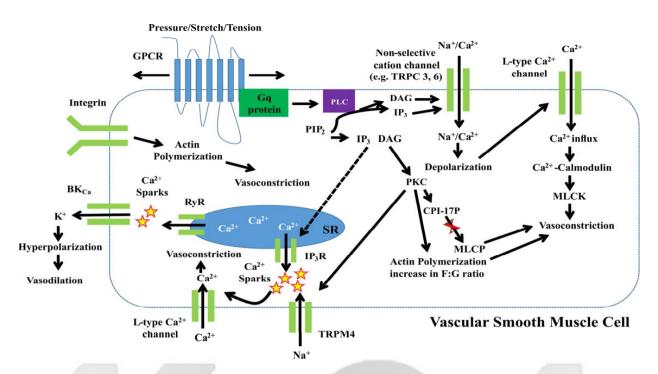


Fig. 1. Schematic diagram showing molecular mechanisms underlying arteriolar myogenic vasoconstriction. Abbreviations: BK<sub>ca</sub>, large conductance Ca<sup>2+</sup>activated K<sup>+</sup> channel; CPI-17, 17-kDa protein kinase C-potentiated inhibitory protein; DAG, diacylglycerol; GPCR, G protein-coupled receptor; IP<sub>3</sub>, inositol trisphosphate; IP<sub>3</sub>R, inositol trisphosphate receptor; MLCK, myosin light chain kinase; MLCP, myosin light chain phosphatase; MYPT1, myosin phosphatase targeting subunit; PIP2, phosphatidylinositol bisphosphate; PKC; protein kinase C; PLC, phospholipase C; RyR, ryanodine receptor, SR, sarcoplasmic reticulum; TRPM4, transient receptor potential melastatin 4.

trical stimulation [43]. Further, it has been demonstrated in human study that elderly individuals have lower leg blood flow and vascular conductance during submaximal-intensity exercise, compared with those of young subjects [44]. In regard to this, a decline in physical performance has been suggested to be due, in part, to age-dependent impairment in myogenic responsiveness [45]. Indeed, myogenic reactivity has been reported to decrease with advancing age in mesenteric, cerebral, and skeletal muscle arterioles [46,47].

Muller-Delp et al. [45] have found that skeletal muscle arterioles isolated from rat soleus muscle (predominantly composed of oxidative muscle fibers) and gastrocnemius muscle (referred to as glycolytic muscle fibers) have a greater myogenic vasoconstriction in young rats compared to those in old rats. The impaired myogenic reactivity with advancing age is not limited to the microcirculation of rodents. Diminished pressure-induced vasoconstriction with aging has been shown in human skeletal muscle arteries [48]. Retinal arteriole autoregulation during exercise (i.e. lifting weights) has been investigated and suggested to be less in elderly individuals [49]. With respect to the decline in myogenic constriction, it has been hypothesized that aging-mediated attenuation of myogenic response is related to alteration in activity of voltage-dependent (K<sub>v</sub>) and/or BK<sub>Ca</sub> channels [50]. The activation of K<sup>+</sup> channels expressed in the VSMCs typically leads to membrane hyperpolarization that suppresses Ca2+ influx through VOCCs and prevents exaggerated myogenic vasoconstriction [11,31]. Thus, pharmacological inhibition of K<sub>v</sub> or BK<sub>Ca</sub> channels with 4-aminopyridine (4-AP) or iberiotoxin, respectively, enhanced myogenic responsiveness in skeletal muscle arterioles in both young and old rats [50]. However, the significant difference in myogenic reactivity of soleus or gastrocnemius muscle arterioles between young and old rats was completely abolished in the presence of 4-AP or iberiotoxin. It is indicated that aging-mediated decline in myogenic vasoconstriction may result from the augmented K+ channel activity in both skeletal muscle arterioles [50].

Beyond small arteries or arterioles in skeletal muscle, the myogenic autoregulation of resistance arterioles in brain such as the circle of Willis and pial vascular bed protects cerebral microcirculation from pressureinduced injury and edema [51]. Even though the importance of the myogenic response has been well-investigated clinically and experimentally [52,53], the question as to how myogenic responsiveness is affected by advanced aging has not fully demonstrated. Middle cerebral arteries isolated from young (3-month-old) and old (24-month-old) mice have shown



**Table 1.** Spontaneous tone and pressure-induced vasoconstriction in young and old arterioles

Animal or human subject	Age	Vessels studied	Vascular responses (Spontaneous tone & Pressure-induced vasoconstriction)	References
Fischer 344 rats	Young: 4 months; Old: 24 months	Soleus and gastrocne- mius muscle arterioles	<ol> <li>Spontaneous tone (young vs. old): 52±3 vs. 25±4% (soleus), 20±4 vs. 11±3% (gastrocnemius)</li> <li>Reduced pressure-induced vasoconstriction in old arterioles (both soleus and gastrocnemius)</li> </ol>	Muller-Delp et al. [45]
Human subjects	Young: $25 \pm 1$ year; Old: $65 \pm 1$ year	Brachial artery	In response to increased intravascular pressure, lower peak mean blood velocity in old subject group (12.43 $\pm$ 1.16 vs. 17.97 $\pm$ 2.01 cm/s)	Lott et al. [48]
Fischer 344 rats	Young: 4 months; Old: 24 months	Soleus and gastrocne- mius muscle arterioles	<ol> <li>Spontaneous tone (young vs. old): 52±4 vs. 44±3% (soleus), 34±2 vs. 30±2% (gastrocnemius)</li> <li>Reduced pressure-induced vasoconstriction in old arterioles (both soleus and gastrocnemius)</li> </ol>	Kang et al. [50]
Fischer 344 rats	Young: 4-6 months; Old: 22-24 months	Soleus muscle arterioles	1) Spontaneous tone (young vs. old): 28±3 vs. 19.4±3.6% 2) Reduced pressure-induced vasoconstriction in old arterioles	Ghosh et al. [46]
C57BL/6 mice	Young: 3 months; Old: 24 months	Middle cerebral arteries	Static intravascular pressure (over 140 mmHg): diminished myogenic constriction in old arteries     Pulsatile intravascular pressure (pulse pressure amplitude: 40 mmHg, frequency: 450/min): lower pressure-induced vaso-constriction in old arteries	Springo et al. [47]

similar myogenic vasoconstriction in response to increases static intraluminal pressure (i.e., 20-100 mmHg) [47]. However, at the relatively high intravascular pressure (i.e., 140 mmHg), myogenic responsiveness of the cerebral arteries has shown to be significantly lower in old mice, compared to that in young mice. Springo and colleagues [47] applied pulsatile intravascular pressure (pulse pressure frequency: 450/min, pulse pressure amplitude: 40 mmHg) to mimic physiological circumstance. In contrast to static intravascular pressure, it was found that pulsatile pressureinduced vasoconstriction is markedly impaired with advanced age. It is suggested that the inappropriate autoregulation of proximal resistance arterioles causes distal cerebral microcirculation to be exposed high intraluminal pressure and subsequent vascular injury [47]. Previous studies showing the negative effects of advancing age on arterial myogenic response are summarized in Table 1.

Meanwhile, there may be some debate of whether advancing age exclusively diminishes pressure-induced vasoconstriction. There are several subtypes of voltage-dependent Ca<sup>2+</sup> channels including L-type, P/Q-type, R-type, and T-type Ca<sup>2+</sup> channels in the VSMCs and/or ECs [54]. In contrast to L-type channels, Ca<sub>v</sub>3.2 T-type channels allow local Ca<sup>2+</sup> entry in the VSMCs, stimulating BK<sub>Ca</sub> channels and eliciting hyperpolarizationinduced decrease in Ca<sup>2+</sup> influx and vasodilation [55]. In this context, mouse mesenteric arteries with a deficiency of Ca<sub>v</sub>3.2 T-type channels display augmented pressure-induced vasoconstriction [56] that has been consistently observed with NiCl2, an inhibitor of Ca<sub>v</sub>3.2 T-type channels, in mesenteric arterioles of young mice (8-17 weeks old) [54]. It is indicated that the T-type channels in the VSMCs appear to limit myogenic vasoconstriction. Interestingly, Mikkelsen & Colleagues [54] have demonstrated that the opposite effect of Ca<sub>v</sub>3.2 T-type channels on myogenic vasoconstriction is markedly abrogated in resistance arterioles of adult mice (28-56 weeks old). It is implicated that myogenic autoregulation can be enhanced with advancing age. Thus, further investigation is required for a better understanding of impact of aging on myogenic responsiveness of small arteries and arterioles.

## **EXERCISE-INDUCED RESTORATION OF** IMPAIRED MYOGENIC REACTIVITY WITH ADVANCING AGE

Intrinsic autoregulation of resistance arteries and arterioles in response to changes in blood pressure plays crucial roles in regulation of local blood flow and peripheral resistance in animals and humans [1]. It has been delineated that the ability of resistance arteries and arterioles to modulate vascular contractility for the satisfaction of appropriate local blood perfusion is enhanced by exercise training [57]. To examine intracellular mechanisms by which exercise ameliorates the myogenic response in porcine coronary resistance arteries, PKC-mediated signaling pathways for myogenic vasoconstriction has been paid attention [57]. PKC has been well-established to regulate the myogenic response through L-type Ca<sup>2+</sup> channel activation, Ca<sup>2+</sup> sensitization, and actin polymerization [38,58]. In this study, coronary arteries isolated from animals involved in



a training program (6 miles per hour, 60-minute, 16 weeks) showed greater pressure-induced vasoconstriction than that of control animals [57]. Further, the trained animals exhibited greater attenuation of myogenic vasoconstriction in the presence of PKC inhibitor chelerythrine, suggesting that the enhanced myogenic response in coronary arteries of trained animals is attributed to increased PKC signaling pathways. This is supported by their additional findings showing that PKC-dependent Ca<sup>2+</sup> influx through L-type Ca<sup>2+</sup> channels was substantially greater in coronary arteries from trained animals. Additionally, decreased Ca<sup>2+</sup> entry by PKC inhibition and PKC- $\alpha$  expression at a protein level were considerably higher in the exercise group [57]. Thus, exercise training may enhance the myogenic vasoconstriction of coronary arteries by modulating PKC-related intracellular signaling.

Based on previous work [57], it may be assumed that regular physical activity improves age-related impairment of myogenic response. Indeed, it has been found that treadmill exercise training (15 meter/minute, 15 incline, 20-60 min/day, 5 days/week, 10-12 weeks) consistently enhances myogenic responsiveness in skeletal muscle arterioles isolated from young (4-6 month old) rats and interestingly restores the attenuation of pressureinduced vasoconstriction with advancing age [46]. The restored myogenic vasoconstriction in old (22-24 months old) rats was largely similar to that in young control rats. It has been demonstrated that an increase in K<sub>v</sub> channel activity is responsible for a reduction in myogenic vasoconstriction with advancing age [50]. As previously described, K+ efflux through K<sub>v</sub> channels leads to membrane hyperpolarization and vasodilation. Thus, it is suggested that aging-induced increase in K<sub>v</sub> channel activity diminishes myogenic vasoconstriction of skeletal muscle arterioles in old rats [50]. Consistent with this interesting findings, Ghosh and colleagues have elucidated that exercise training elicits K<sub>v</sub>1 channel adaptation (i.e. presumably decreases in channel activity and/or expression), thereby restoring arterial myogenic responsiveness from old rats [46]. However, despite those novel studies [46,57], the mechanisms underlying exercise-mediated enhancement of the myogenic response remain poorly understood. Therefore, further investigations into the role of exercise training in the myogenic response are needed. In addition, it may be worth investigating whether physical inactivity (e.g., bed rest) deteriorates activities of biological machineries that are involved in the myogenic autoregulation and whether the impaired myogenic response by physical inactivity could be restored by exercise training.

#### CONCLUSION

Small arteries and arterioles are considered as mechanosensitive blood vessels that control their diameter to regulate local microvascular hemodynamics by sensing alteration in intraluminal pressure. Exaggerated or attenuated myogenic responsiveness in resistance arteries has been reported to cause pathophysiological conditions such as hypertension, vasospasm, ischemic stroke, or orthostatic hypotension. The prevalence of vascular diseases is associated with advancing age, which may be partly attributed to age-dependent impairment of myogenic responsiveness in resistance arteries. In view of prevention and treatment of cardiovascular disorders, a decline in the myogenic response with advancing age has been a therapeutic target. Taken together, greater insight of specific mechanisms underlying the impact of regular physical activity on arterial myogenic responsiveness has to be further made to reduce cardiovascular diseases-related mortality in elderly individuals.

### **ACKNOWLEDGEMENT**

We cordially thank Dr. Michael Hill, University of Missouri-Columbia, for constructive review of the manuscript prior to submission.

#### REFERENCES

- 1. Davis MJ, Hill MA. Signaling mechanisms underlying the vascular myogenic response. Physiological Reviews 1999;79(2):387-423.
- 2. Bayliss WM. On the local reactions of the arterial wall to changes of internal pressure. The Journal of Physiology 1902;28(3):220-231.
- 3. Murphy TV, Spurrell BE, Hill MA. Cellular signalling in arteriolar myogenic constriction: involvement of tyrosine phosphorylation pathways. Clinical and Experimental Pharmacology and Physiology 2002;29(7): 612-619.
- 4. Hill MA, Meininger GA, Davis MJ, Laher I. Therapeutic potential of pharmacologically targeting arteriolar myogenic tone. Trends in Pharmacological Sciences 2009;30(7):363-374.
- 5. Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a "set up" for vascular disease. Circulation 2003;107(1):139-146.
- 6. Harvey A, Montezano AC, Lopes RA, Rios F, Touyz RM. Vascular fibrosis in aging and hypertension: Molecular mechanisms and clinical implications. Canadian Journal of Cardiology 2016;32(5):659-668.



- 7. Mikhed Y, Daiber A, Steven S. Mitochondrial oxidative stress, mitochondrial DNA damage and their role in age-related vascular dysfunction. International Journal of Molecular Sciences 2015;16(7):15918-15953.
- 8. Kojda G, Hambrecht R. Molecular mechanisms of vascular adaptations to exercise. Physical activity as an effective antioxidant therapy?. Cardiovascular Research 2005;67(2):187-197.
- 9. Cleroux J, Feldman RD, Petrella RJ. Lifestyle modifications to prevent and control hypertension. 4. Recommendations on physical exercise training. Canadian Hypertension Society, Canadian Coalition for High Blood Pressure Prevention and Control, Laboratory Centre for Disease Control at Health Canada, Heart and Stroke Foundation of Canada. Canadian Medical Association Journal 1999;160(9):S21-S28.
- 10. Piepoli MF, Davos C, Francis DP, Coats AJ, ExTra MC. Exercise training meta-analysis of trials in patients with chronic heart failure (Ex-TraMATCH). British Medical Journal 2004;328(7433):189.
- 11. Hill MA, Yang Y, Ella SR, Davis MJ, Braun AP. Large conductance, Ca<sup>2+</sup>-activated K<sup>+</sup> channels (BK<sub>Ca</sub>) and arteriolar myogenic signaling. FEBS Letters 2010;584(10):2033-2042.
- 12. Wu X, Mogford JE, Platts SH, Davis GE, Meininger GA, et al. Modulation of calcium current in arteriolar smooth muscle by alphay beta3 and alpha5 beta1 integrin ligands. The Journal of Cell Biology 1998; 143(1):241-252.
- 13. Martinez-Lemus LA, Crow T, Davis MJ, Meininger GA.  $\alpha_{v}\beta_{3}$  and  $\alpha_{5}\beta_{1}$ integrin blockade inhibits myogenic constriction of skeletal muscle resistance arterioles. American Journal of Physiology-Heart and Circulatory Physiology 2005;289(1):H322-H329.
- 14. Mufti RE, Zechariah A, Sancho M, Mazumdar N, Brett SE, et al. Implications of alphavbeta3 integrin signaling in the regulation of Ca<sup>2+</sup> waves and myogenic tone in cerebral arteries. Arteriosclerosis, Thrombosis, and Vascular Biology 2015;35(12):2571-2578.
- 15. Yasuda N, Miura S, Akazawa H, Tanaka T, Qin Y, et al. Conformational switch of angiotensin II type 1 receptor underlying mechanical stress-induced activation. EMBO Reports 2008;9(2):179-186.
- 16. Li Y, Baylie RL, Tavares MJ, Brayden JE. TRPM4 channels couple purinergic receptor mechanoactivation and myogenic tone development in cerebral parenchymal arterioles. Journal of Cerebral Blood Flow & Metabolism 2014;34(10):1706-1714.
- 17. Storch U, Blodow S, Gudermann T, Mederos YSM. Cysteinyl leukotriene 1 receptors as novel mechanosensors mediating myogenic tone together with angiotensin II type 1 receptors-brief report. Arterioscle-

- rosis, Thrombosis, and Vascular Biology 2015;35(1):121-126.
- 18. Hong K, Zhao G, Hong Z, Sun Z, Yang Y, et al. Mechanical activation of angiotensin II type 1 receptors causes actin remodelling and myogenic responsiveness in skeletal muscle arterioles. The Journal of Physiology 2016;594(23):7027-7047.
- 19. Schleifenbaum J, Kassmann M, Szijarto IA, Hercule HC, Tano JY, et al. Stretch-activation of angiotensin II type 1a receptors contributes to the myogenic response of mouse mesenteric and renal arteries. Circulation Research 2014;115(2):263-272.
- 20. Earley S, Brayden JE. Transient receptor potential channels in the vasculature. Physiological Reviews 2015;95(2):645-690.
- 21. Welsh DG, Morielli AD, Nelson MT, Brayden JE. Transient receptor potential channels regulate myogenic tone of resistance arteries. Circulation Research 2002;90(3):248-250.
- 22. Gonzales AL, Yang Y, Sullivan MN, Sanders L, Dabertrand F, et al. A PLCgamma1-dependent, force-sensitive signaling network in the myogenic constriction of cerebral arteries. Science Signaling 2014;7:ra49 (DOI: 10.1126/scisignal.2004732).
- 23. Drummond HA, Grifoni SC, Jernigan NL. A new trick for an old dogma: ENaC proteins as mechanotransducers in vascular smooth muscle. Physiology 2008a;23(1):23-31.
- 24. Drummond HA, Jernigan NL, Grifoni SC. Sensing tension: epithelial sodium channel/acid-sensing ion channel proteins in cardiovascular homeostasis. Hypertension 2008b;51(5):1265-1271.
- 25. Jernigan NL, Drummond HA. Vascular ENaC proteins are required for renal myogenic constriction. American Journal of Physiology-Renal Physiology 2005;289(4):F891-F901.
- 26. Kim EC, Choi SK, Lim M, Yeon SI, Lee YH. Role of endogenous ENaC and TRP channels in the myogenic response of rat posterior cerebral arteries. PLoS One 2013;8(12):e84194.
- 27. Jaggar JH, Porter VA, Lederer WJ, Nelson MT. Calcium sparks in smooth muscle. American Journal of Physiology-Cell Physiology 2000;278(2): C235-C256.
- 28. Evanson KW, Bannister JP, Leo MD, Jaggar JH. LRRC26 is a functional BK channel auxiliary gamma subunit in arterial smooth muscle cells. Circulation Research 2014;115(4):423-431.
- 29. Yang Y, Sohma Y, Nourian Z, Ella SR, Li M, et al. Mechanisms underlying regional differences in the Ca<sup>2+</sup> sensitivity of BK(Ca) current in arteriolar smooth muscle. The Journal of Physiology 2013;591(5):1277-1293.
- 30. Mackie AR, Brueggemann LI, Henderson KK, Shiels AJ, Cribbs LL, et



- al. Vascular KCNQ potassium channels as novel targets for the control of mesenteric artery constriction by vasopressin, based on studies in single cells, pressurized arteries, and in vivo measurements of mesenteric vascular resistance. The Journal of Pharmacology and Experimental Therapeutics 2008;325(2):475-483.
- 31. Zhong XZ, Harhun MI, Olesen SP, Ohya S, Moffatt JD, et al. Participation of KCNQ (Kv7) potassium channels in myogenic control of cerebral arterial diameter. The Journal of Physiology 2010;588(17):3277-
- 32. Wray S, Burdyga T. Sarcoplasmic reticulum function in smooth muscle. Physiological Reviews 2010;90(1):113-178.
- 33. Kamm KE, Stull JT. Dedicated myosin light chain kinases with diverse cellular functions. The Journal of Biological Chemistry 2001;276(7): 4527-4530.
- 34. Wilson DP, Sutherland C, Walsh MP. Ca<sup>2+</sup> activation of smooth muscle contraction: evidence for the involvement of calmodulin that is bound to the triton insoluble fraction even in the absence of Ca<sup>2+</sup>. The Journal of Biological Chemistry 2002;277(3):2186-2192.
- 35. Osol G, Brekke JF, McElroy-Yaggy K, Gokina NI. Myogenic tone, reactivity, and forced dilatation: a three-phase model of in vitro arterial myogenic behavior. American Journal of Physiology-Heart and Circulatory Physiology 2002;283(6):H2260-2267.
- 36. Eto M. Regulation of cellular protein phosphatase-1 (PP1) by phosphorylation of the CPI-17 family, C-kinase-activated PP1 inhibitors. The Journal of Biological Chemistry 2009;284(51):35273-35277.
- 37. Muranyi A, Derkach D, Erdodi F, Kiss A, Ito M, et al. Phosphorylation of Thr695 and Thr850 on the myosin phosphatase target subunit: inhibitory effects and occurrence in A7r5 cells. FEBS Letters 2005;579(29): 6611-6615.
- 38. Moreno-Dominguez A, Colinas O, El-Yazbi A, Walsh EJ, Hill MA, et al. Ca<sup>2+</sup> sensitization due to myosin light chain phosphatase inhibition and cytoskeletal reorganization in the myogenic response of skeletal muscle resistance arteries. The Journal of Physiology 2013;591(5):1235-1250.
- 39. Walsh MP, Cole WC. The role of actin filament dynamics in the myogenic response of cerebral resistance arteries. Journal of Cerebral Blood Flow & Metabolism 2013;33(1):1-12.
- 40. Yamin R, Morgan KG. Deciphering actin cytoskeletal function in the contractile vascular smooth muscle cell. The Journal of Physiology 2012;590(17):4145-4154.
- 41. Moreno-Dominguez A, El-Yazbi AF, Zhu HL, Colinas O, Zhong XZ,

- et al. Cytoskeletal reorganization evoked by Rho-associated kinaseand protein kinase C-catalyzed phosphorylation of cofilin and heat shock protein 27, respectively, contributes to myogenic constriction of rat cerebral arteries. The Journal of Biological Chemistry 2014;289(30): 20939-20952.
- 42. Proctor DN, Parker BA. Vasodilation and vascular control in contracting muscle of the aging human. Microcirculation 2006;13(4):315-327.
- 43. Irion GL, Vasthare US, Tuma RF. Age-related change in skeletal muscle blood flow in the rat. The Journal of Gerontology 1987;42(6):660-665.
- 44. Proctor DN, Shen PH, Dietz NM, Eickhoff TJ, Lawler LA, et al. Reduced leg blood flow during dynamic exercise in older endurance-trained men. Journal of Applied Physiology 1998;85(1):68-75.
- 45. Muller-Delp JM, Spier SA, Ramsey MW, Lesniewski LA, Papadopoulos A, et al. Effects of aging on vasoconstrictor and mechanical properties of rat skeletal muscle arterioles. American Journal of Physiology-Heart and Circulatory Physiology 2002;282(5):H1843-H1854.
- 46. Ghosh P, Mora Solis FR, Dominguez JM, Spier SA, Donato AJ, et al. Exercise training reverses aging-induced impairment of myogenic constriction in skeletal muscle arterioles. Journal of Applied Physiology 2015;118(7):904-911.
- 47. Springo Z, Toth P, Tarantini S, Ashpole NM, Tucsek Z, et al. Aging impairs myogenic adaptation to pulsatile pressure in mouse cerebral arteries. Journal of Cerebral Blood Flow & Metabolism 2015;35(4):527-530.
- 48. Lott ME, Herr MD, Sinoway LI. Effects of age on brachial artery myogenic responses in humans. American Journal of Physiology-Regulatory, Integrative and Comparative Physiology 2004;287(3):R586-R591.
- 49. Jeppesen P, Gregersen PA, Bek T. The age-dependent decrease in the myogenic response of retinal arterioles as studied with the retinal vesel analyzer. Graefe's Archive for Clinical and Experimental Ophthalmology 2004;242(11):914-919.
- 50. Kang LS, Kim S, Dominguez JM, Sindler AL, Dick GM, et al. Aging and muscle fiber type alter K+ channel contributions to the myogenic response in skeletal muscle arterioles. Journal of Applied Physiology 2009;107(2):389-398.
- 51. Vrselja Z, Brkic H, Mrdenovic S, Radic R, Curic G. Function of circle of willis. Journal of Cerebral Blood Flow & Metabolism 2014;34(4): 578-584
- 52. Toth P, Csiszar A, Tucsek Z, Sosnowska D, Gautam T, et al. Role of 20-HETE, C channels, and BK<sub>Ca</sub> in dysregulation of pressure-induced Ca<sup>2+</sup>



- signaling and myogenic constriction of cerebral arteries in aged hypertensive mice. American Journal of Physiology-Heart and Circulatory Physiology 2013a;305(12):H1698-H1708.
- 53. Toth P, Tucsek Z, Sosnowska D, Gautam T, Mitschelen M, et al. Agerelated autoregulatory dysfunction and cerebromicrovascular injury in mice with angiotensin II-induced hypertension. Journal of Cerebral Blood Flow & Metabolism 2013b;33(11):1732-1742.
- 54. Mikkelsen MF, Bjorling K, Jensen LJ. Age-dependent impact of Ca 3.2 T-type calcium channel deletion on myogenic tone and flow-mediated vasodilatation in small arteries. The Journal of Physiology 2016;594(20): 5881-5898.
- 55. Harraz OF, Abd El-Rahman RR, Bigdely-Shamloo K, Wilson SM, Brett SE, et al. Ca(v)3.2 channels and the induction of negative feedback in

- cerebral arteries. Circulation Research 2014;115(7):650-661.
- 56. Harraz OF, Brett SE, Zechariah A, Romero M, Puglisi JL, et al. Genetic ablation of Ca<sub>V</sub>3.2 channels enhances the arterial myogenic response by modulating the RyR-BK<sub>Ca</sub> axis. Arteriosclerosis, Thrombosis, and Vascular Biology 2015;35(8):1843-1851.
- 57. Korzick DH, Laughlin MH, Bowles DK. Alterations in PKC signaling underlie enhanced myogenic tone in exercise-trained porcine coronary resistance arteries. Journal of Applied Physiology 2004;96(4):1425-1432.
- 58. Nakayama K, Obara K, Tanabe Y, Saito M, Ishikawa T, et al. Interactive role of tyrosine kinase, protein kinase C, and Rho/Rho kinase systems in the mechanotransduction of vascular smooth muscles. Biorheology 2003;40(1,2,3):307-314.

