


 Cite this: *RSC Adv.*, 2023, **13**, 31962

A colorimetric and fluorescent signaling probe for assaying Pd²⁺ in practical samples†

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We developed an optical signaling probe to detect Pd²⁺ ions in Pd-containing catalyst and drug candidate. The Pd²⁺ signaling probe (Res-DT) was readily prepared by reacting the versatile fluorochrome resorufin with phenyl chlorodithioformate. In a phosphate-buffered saline solution (pH 7.4) containing sodium dodecyl sulfate (SDS) as a signal-boosting surfactant, Res-DT exhibited a pronounced colorimetric response with a chromogenic yellow to magenta shift, leading to a substantial increase in the fluorescence intensity. The Pd²⁺ signaling performance of Res-DT was attributed to the Pd²⁺-promoted hydrolysis of the dithioate moiety. The probe displayed high selectivity toward Pd²⁺ ions and remained unaffected by commonly encountered coexisting components. Moreover, the detection limit of Res-DT for Pd²⁺ ions was 10 nM, and the signaling was achieved within 7 min. Furthermore, to demonstrate the real-world applicability of Res-DT, a Pd²⁺ assay was performed in Pd-containing catalyst and drug candidate using an office scanner as an easily accessible measurement device. Our results highlight the prospects of Res-DT as a tool to detect Pd²⁺ ions in various practical samples, with potential applications in catalysis, medicine, and environmental science.

 Received 15th August 2023
 Accepted 19th October 2023

DOI: 10.1039/d3ra05549c

rsc.li/rsc-advances

1. Introduction

Palladium (Pd) is a versatile and valuable Pt-group metal with diverse practical applications in industrial arenas such as dentistry, electronics, chemical synthesis, groundwater treatment, and exhaust gas treatment.^{1,2} Additionally, it is employed in therapeutics owing to its antiviral, antifungal, antimicrobial, anticancer, and cardioprotective properties.³ Furthermore, it is a vital component of fuel cells that generate energy through chemical reactions involving hydrogen and oxygen.⁴

Pd-based catalysts are essential for producing myriad fine chemicals that are used to manufacture pharmaceuticals and agricultural products.⁵ In particular, several Pd-catalyzed organic reactions, including the Mizoroki–Heck reaction,⁶ Suzuki–Miyaura reaction,⁷ Sonogashira–Hagihara reaction,⁸ Buchwald–Hartwig amination,⁹ carbonylation,¹⁰ and cyanation,¹¹ are performed in pharmaceutical preparations.¹² Moreover, Pd complexes can be used as anticancer agents owing to their similar chemical and physical properties to those of the widely employed Pt complexes.¹³ However, the inherent toxicity of Pd has made Pd contamination a matter of considerable concern.¹⁴ In particular, Pd species can bind to important biological materials, such as amino acids, DNA, RNA, and

proteins,^{15,16} thereby disrupting cellular processes and leading to severe health problems including weight loss, muscle weakness, seizures, and heart disease.¹⁷ Consequently, the determination of residual Pd in commercially available drug chemicals, agricultural products, and foods is crucial.

Various conventional analytical techniques have been employed to detect Pd in different analytes using sophisticated, specialized analytical instruments.¹⁸ However, these techniques typically require complex sample preparation protocols, stringent experimental conditions, and highly skilled operators.¹⁹ In contrast, colorimetric or fluorescent chemosensors and reaction-based probes hold greater promise for selective and sensitive metal ion/anion detection, including Pd. These methods offer straightforward operability, high sensitivity, no need for heavy instruments, and widespread applicability.²⁰

To meet the increasing need for simpler and more convenient Pd analysis methods, several colorimetry or fluorescence-based chemosensors and reaction-based probes have been developed.^{19,21} Pd signaling sensors have been obtained using diverse ligands including pyridine-2,6-dicarboxamide,²² 2-chloroethyl methyl sulfide,²³ purine derivative,²⁴ and 2-picolyamine.²⁵ Additionally, several Pd-selective signaling probes have been developed by leveraging their Pd-selective reactions, signal-accumulating ability, and ease of design.²⁶ For instance, deallylation of allyl carbamates,²⁷ allyl carbonates,²⁸ allyl ethers,²⁹ and allyl ester,³⁰ and depropargylation of propargyl ethers³¹ and propargyl carbamates³² have been extensively performed to design Pd signaling probes. In addition, probes that detect Pd *via* metal-induced organic transformations have been

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† Electronic supplementary information (ESI) available: Results of Pd²⁺-selective UV-vis and fluorescence signaling experiments; ¹H, ¹³C NMR, and mass spectra of the Res-DT. See DOI: <https://doi.org/10.1039/d3ra05549c>



prepared using the Claisen rearrangement,³³ dimerization,³⁴ and oxidative cyclization reactions.³⁵ Furthermore, several Pd signaling probes that exploit the hydrolysis of thiocarbamate and hydrazones of resorufin and rhodamine fluorochromes have been engineered.³⁶ There have also been various studies exploring Pd²⁺ sensing probes, each contributing valuable insights into their development and applications.³⁷ Representative optical Pd specific reaction-based probes have been summarized in Table S1 (ESI).†

In this study, we introduce a novel dual-mode probe (**Res-DT**) designed for the highly sensitive detection of Pd²⁺ ions through both colorimetric and fluorescent responses, facilitated by the hydrolysis of dithioate-modified resorufin. The probe ensures rapid, convenient, and naked-eye detectable responses, obviating the need for complex instrumentation. **Res-DT** demonstrates efficiency in swiftly and precisely assaying residual Pd²⁺ in a Pd-containing catalyst and a Pd-containing drug candidate noted for its antimicrobial and anticancer activities.

2. Experimental

2.1 Synthesis of Res-DT

Resorufin dithioate (**Res-DT**) was synthesized using a previously reported method with slight modifications.³⁸ In a 100 mL round bottom flask, resorufin (0.43 g, 2.0 mmol) was dissolved in 30 mL of *N,N*-dimethylformamide (DMF). The solution was then mixed with triethylamine (TEA; 0.56 mL, 4.0 mmol) and stirred for 30 min at room temperature. Phenyl chlorodithioformate (0.45 mL, 3.0 mmol) was then added carefully to the solution, and the reaction was allowed to continue for 12 h with constant stirring. DMF was then removed by passing air over the system, and the remaining solid was dissolved in dichloromethane (50 mL). The resulting solution was washed with distilled water and brine and then evaporated. The obtained residue was purified by column chromatography (CH₂Cl₂ : CH₃OH = 49 : 1, v/v). **Res-DT**. 0.52 g, 71% yield as a vermilion-colored powder. ¹H NMR (600 MHz, CDCl₃) δ 7.81 (dt, *J* = 8.9, 1.2 Hz, 1H), 7.64–7.59 (m, 2H), 7.54–7.46 (m, 3H), 7.42 (d, *J* = 9.8 Hz, 1H), 7.13–7.10 (m, 2H), 6.86 (dd, *J* = 9.8, 2.0 Hz, 1H), 6.32 (d, *J* = 2.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃): δ 212.4, 186.2, 156.6, 149.1, 148.7, 144.4, 135.3, 135.2, 134.8, 131.8, 131.2, 130.7, 129.8, 129.7, 119.9, 110.6, 107.3; HRMS (EI⁺, *m/z*): calcd. for C₁₉H₁₁NO₃S₂⁺ [M]⁺: 365.0180, found 365.0179.

2.2 Stock solution preparation

A stock solution containing probe **Res-DT** (0.5 mM) was made by dissolving **Res-DT** in dimethyl sulfoxide (DMSO). Pd-containing solutions (5.0 mM) were prepared by dissolving Pd(OAc)₂, PdCl₂, and Pd(PPh₃)₄ in DMSO, and K₂PdCl₆ in deionized (DI) water. Solutions of metal ions and anions (5.0 mM) were prepared using metal perchlorate salts and sodium salts of the anions, respectively, in DI water. Oxidant solutions (5.0 mM), including H₂O₂, HOCl, peracetic acid, O₂[−], perborate, percarbonate, and ammonium persulfate, were prepared and standardized as described previously.³⁹

2.3 Investigation of Pd²⁺ signaling with Res-DT

We investigated the Pd²⁺ sensing behavior of **Res-DT** in a pH 7.4 phosphate-buffered saline (PBS) solution, using 2% DMSO as a solubilizer. To that end, the analyte stock solution (15 μL, 5.0 mM) was first added to a sample tube and diluted with DI water (2.34 mL) and a predetermined amount of DMSO. Then, a PBS solution (0.30 mL, 100 mM) and SDS (0.30 mL, 100 mM) were added to the mixture, followed by **Res-DT** (30 μL, 0.50 mM). The final concentrations of **Res-DT**, the analyte, PBS, and SDS were 5.0 μM, 25 μM, 10.0 mM, and 10.0 mM respectively. Error bars were determined based on the standard deviation derived from three sets of experiments.

2.4 Mechanism study of Pd²⁺ signaling

In a 100 mL round bottom flask, **Res-DT** (37 mg, 100 μmol) was dissolved in CH₃CN (20 mL). The resulting solution was then mixed with palladium acetate (56 mg, 250 μmol). Upon verifying the completion of the reaction by thin-layer chromatography, the precipitate was collected using a centrifuge (4000 rpm) and washed with CH₃CN. The precipitate was dried and analyzed using field-emission scanning electron microscopy (FE-SEM) equipped with energy-dispersive X-ray spectroscopy (EDX). The remaining solution was evaporated under reduced pressure and the residue was purified by column chromatography. The purified product of the Pd²⁺ signaling of **Res-DT** was scrutinized by ¹H NMR and mass spectrometry measurements.

2.5 Office scanner-based determination of Pd²⁺ concentration in Pd-containing catalyst and drug candidate

To determine the Pd²⁺ concentration in Pd-containing catalyst and drug candidate, five samples with varying Pd²⁺ levels were tested. These samples were prepared by mixing a predetermined amount of the Pd-containing catalyst or drug (ranging from 0 to 6 μL; 1.0 mM), DI water (2.34 mL), DMSO (15 μL), PBS (0.30 mL, 0.10 M), SDS (0.30 mL, 0.10 M), and **Res-DT** (30 μL, 0.50 mM) in a sample vial. The resulting solutions (0.30 mL) were then individually transferred into a 96-well plate. A calibration plot of the color channel levels (RGB) against the Pd species concentration was constructed thereafter using an office scanner (V550, Epson) in transmittance mode.

3. Results and discussion

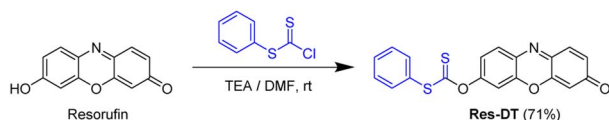
Sulfur-based chemosignaling probes have been extensively used to detect thiophilic metal ions and common oxidants through the desulfurization and oxidative hydrolysis reactions.^{38,40} In the present study, the dithioate moiety, which is prone to desulfurization-induced hydrolysis in the presence of hypochlorite ions,³⁸ was targeted in this study to develop a colorimetric, fluorescent signaling probe for analyzing Pd²⁺ in catalysts and drugs. The undesired response toward hypochlorite ions can be readily suppressed using hypochlorite-scavenging DMSO.⁴¹ Based on this rationale, a simple but optically vibrant dithioate-based Pd²⁺ signaling probe, called **Res-DT**, was developed. Probe **Res-DT** was synthesized by reacting resorufin with phenyl chlorodithioformate (Scheme 1)



and then characterized by NMR spectroscopy and mass spectrometry.

First, the Pd^{2+} signaling condition of **Res-DT** was optimized by measuring the changes in absorbance at 572 nm. Preliminary results indicated that **Res-DT** exhibited moderate Pd^{2+} signaling activity in pH 7.4 PBS with 2% DMSO which acted both as a solubilizer and a scavenger for potential hypochlorite interferants (Fig. S1, ESI[†]). However, the signaling performance was relatively sluggish for practical applications, given that the reaction was incomplete even after 30 min. Therefore, to improve the reaction rate of **Res-DT** for Pd^{2+} ions, three types of surfactants were employed: anionic (SDS), cationic (cetyltrimethylammonium bromide (CTAB)), and nonionic (Tween 20). According to the results (Fig. S2a, ESI[†]), the Pd^{2+} signaling behavior of **Res-DT** was substantially improved upon using SDS (10.0 mM). The other surfactant solutions (CTAB and Tween 20) also enhanced the Pd^{2+} signaling speed of **Res-DT** (Fig. S2b and c, ESI[†]). However, **Res-DT** was slightly hydrolyzed when the CTAB solution (1.0 mM) was used, and undesirable precipitates were formed when the Tween 20-containing solution (0.09 mM) was employed (Fig. S3, ESI[†]). Therefore, subsequent Pd^{2+} signaling experiments with **Res-DT** were performed using the SDS surfactant solution as a signal booster.

The colorimetric and fluorescence signaling properties of **Res-DT** for representative metal ions and anions were investigated under optimized conditions. First, the colorimetric signaling response of **Res-DT** was examined upon exposure to various common metal ions (Fig. 1). **Res-DT** exhibited a weak absorption band at 452 nm with pale-yellow coloration in the measurement solution. However, after treatment with Pd^{2+} ions, it showed a strong increase and moderate decrease in the absorbance at 572 and 452 nm, respectively. Concomitantly, the solution turned magenta from pale yellow (Fig. 1, inset). Furthermore, the other tested metal ions did not induce noticeable changes in the UV-vis spectra or color. Because the signaling ability of **Res-DT** for Pd^{2+} was related to the absorbance variation at two widely separated wavelengths, 572 nm and 452 nm, the selectivity for Pd^{2+} was assessed by ratiometry using the absorbance ratio (A_{572}/A_{452}) (Fig. 1). The A_{572}/A_{452} value of **Res-DT** with Pd^{2+} was 10.2, whereas those of the other tested metal ions were remarkably low and similar to that of **Res-DT** alone (values ranging from 0.04 (Mg^{2+}) to 0.11 (Pt^{2+})). Additionally, the Pd^{2+} selectivity of **Res-DT** was also confirmed in the presence of several representative anions (Fig. S4, ESI[†]) by showing the low A_{572}/A_{452} values of **Res-DT** for anions (ranging from 0.06 (Cl^-) to 0.08 (N_3^-)). Furthermore, taking into the fact that the dithioate-based probe has been used for hypochlorite sensing,³⁸ the changes in the absorbance ratio of **Res-DT** upon treatment with representative oxidants were



Scheme 1 Preparation of the Pd^{2+} signaling probe (**Res-DT**).

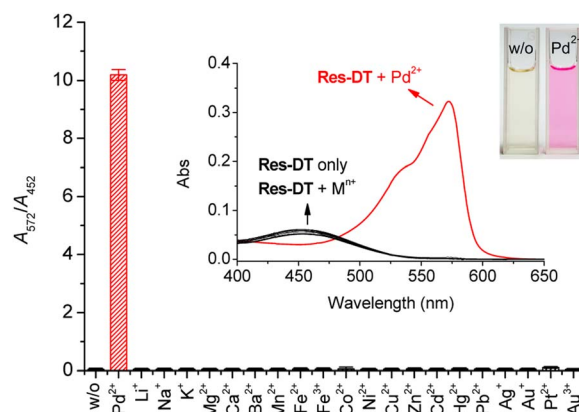


Fig. 1 Changes in the absorbance ratio of **Res-DT** (A_{572}/A_{452}) with the incorporation of common metal ions. Inset: UV-vis spectra and naked-eye photographs of **Res-DT**. [**Res-DT**] = 5.0 μM , [Pd^{2+}] = [M^{n+}] = 25 μM , [SDS] = 10.0 mM, [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO.

measured. The results (Fig. S5, ESI[†]) indicated that the tested oxidants, including hypochlorite ions, negligibly altered the A_{572}/A_{452} ratio (ranging from 0.05 for perborate (PB) to 0.06 for *tert*-butyl hydroperoxide (TBHP)). The photophysical properties of **Res-DT**, both pre- and post- Pd^{2+} signaling, are detailed in Table S2 (ESI[†]).

Next, to confirm the influence of background ions on the Pd^{2+} signaling tendency of **Res-DT**, sensing experiments with coexisting metal ions and anions were performed. According to the results (Fig. 2), the Pd^{2+} signaling behavior of **Res-DT** was unaffected by the presence of coexisting metal ions. Essentially, the A_{572}/A_{452} value of the samples after the Pd^{2+} signaling experiments varied only slightly, ranging from 93.6% (for Fe^{2+}) to 107.3% (for Ba^{2+}) of the control result. In addition, the Pd^{2+} signaling performance of **Res-DT** was not altered by the

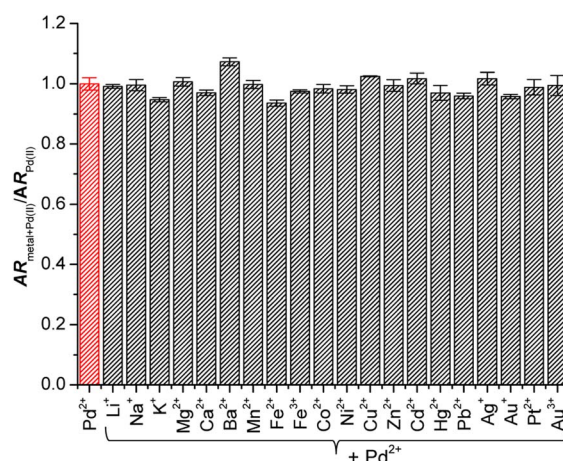


Fig. 2 Effect of coexisting metal ions on the Pd^{2+} signaling activity of **Res-DT**. [**Res-DT**] = 5.0 μM , [Pd^{2+}] = [M^{n+}] = 25 μM , [SDS] = 10.0 mM, [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO. AR denotes the absorbance ratio A_{572}/A_{452} .



presence of different anions, given the narrow range of the A_{572}/A_{452} values (92.5% for N_3^- to 103.0% for F^-) (Fig. S6, ESI†).

Resorufin-based sensing probes typically exhibit significant fluorescence features as well as remarkable colorimetric signaling behavior.⁴² Therefore, the Pd^{2+} signaling performance of **Res-DT** was evaluated based on the changes in the fluorescence response occurring under the same sensing conditions. Based on the results (Fig. 3, inset), **Res-DT** exhibited faint fluorescence emission at approximately 586 nm ($\Phi_{Res-DT} = 0.009$) owing to the photophysical characteristics of the phenolic moiety-protected resorufin fluorophore.⁴³ However, upon exposure to Pd^{2+} ions, **Res-DT** revealed strong fluorescence emission ($\Phi_{Res-DT+Pd(n)} = 0.58$), exhibiting over a 90-fold fluorescence enhancement at 591 nm (Fig. 3). All other tested metal ions showed insignificant fluorescence responses except for Au^{3+} ions ($I/I_0 = 5.88$), with I/I_0 generally fluctuating between 0.97 (for Li^+) and 1.32 (for Pt^{2+}). Furthermore, no measurable changes were observed in the fluorescence emission of **Res-DT** toward the encountered anions, with I/I_0 varying from 0.99 (for Cl^-) to 1.50 (for N_3^-) (Fig. S7, ESI†). These results highlight the fluorometric potential of **Res-DT** to sense Pd^{2+} ions in chemical and industrial applications. However, the Pd^{2+} sensing behavior of **Res-DT** was investigated using colorimetric measurements which allowed the ratiometric analysis, rather than the method relying on a simple turn-on type fluorescence enhancement at a single wavelength. Ratiometry offers several advantages including increased sensitivity, improved selectivity, and reduced interference from the effects of interfering substances in the sample matrix.⁴⁴

The Pd^{2+} signaling was hypothesized to be caused by the generation of the resorufin fluorochrome *via* Pd^{2+} -mediated hydrolysis of the dithioate moiety of **Res-DT** (Scheme 2). In the proposed sensing mechanism of **Res-DT**, the initial stage involves complex formation between the sulfur atom of the C=S bond and thiophilic Pd^{2+} ions.⁴⁵ The complex is then hydrolyzed, yielding resorufin dye with its distinctive magenta color and strong fluorescence signals. To confirm the sensing

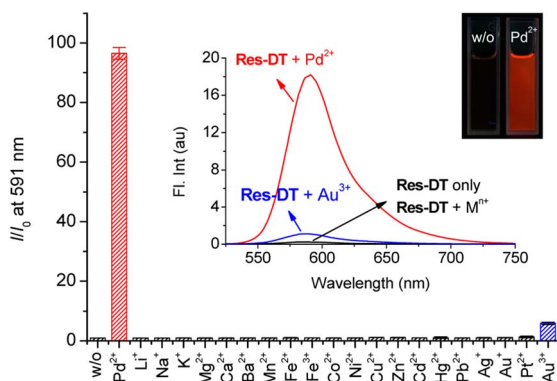
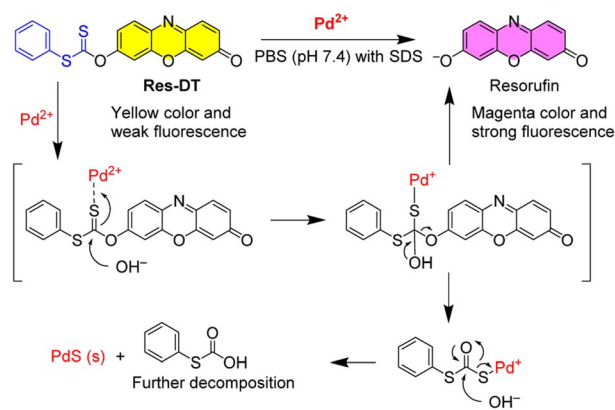


Fig. 3 Changes in the fluorescence enhancement of **Res-DT** at 591 nm with the incorporation of common metal ions. Inset: fluorescence spectra and photographs with hand-held UV-lamp irradiation of **Res-DT**. [**Res-DT**] = 5.0 μ M, [Pd^{2+}] = [M^{n+}] = 25 μ M, [SDS] = 10.0 mM, [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO. λ_{ex} = 485 nm.



Scheme 2 Suggested Pd^{2+} sensing mechanism of **Res-DT**.

mechanism of **Res-DT**, the Pd^{2+} signaling product was scrutinized using 1H NMR measurements. The results indicated that the NMR pattern of **Res-DT** was similar to that of typical phenol-protected resorufin compounds (Fig. 4).⁴⁰ Moreover, the purified Pd^{2+} sensing product (**Res-DT** + Pd^{2+}) exhibited three well-defined resonances in the 1H NMR spectrum that were likely associated with resorufin. Furthermore, our investigation using thin-layer chromatography of the Pd^{2+} signaling solution of **Res-DT** revealed that resorufin is produced (Fig. S8, ESI†). Through FAB mass spectrometry, a diagnostic peak at $m/z = 214$ was identified in the Pd^{2+} signaling product, which matches the calculated mass of resorufin ($C_{12}H_8NO_3^+ [M + H] = 214$) (Fig. S9, ESI†). Additionally, we isolated a greenish-black colored precipitate from the Pd^{2+} signaling solution and confirmed it to be PdS using FE-SEM equipped with EDX (Fig. S10, ESI†).

The influence of pH on the Pd^{2+} signaling performance of **Res-DT** was tested to assess its practical usability. The results showed that the A_{572}/A_{452} value of pristine **Res-DT** remained constant across the pH range 4.0–10.0 (Fig. 5). In contrast, that of the Pd^{2+} signaling sample (**Res-DT** + Pd^{2+}) increased significantly from pH 6.0 onward and stabilized at around pH 8; this tendency was mirrored by that of the reference compound resorufin in the presence of coexisting Pd^{2+} ions. This finding

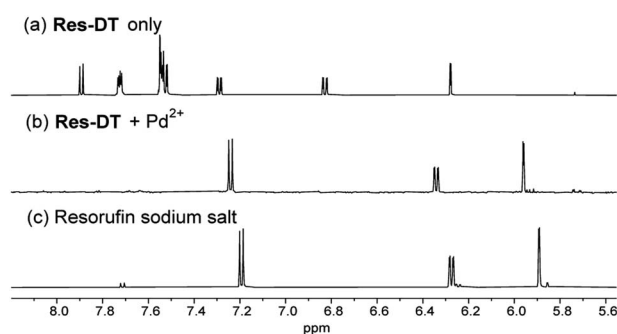


Fig. 4 Partial 1H NMR spectra of (a) **Res-DT**, (b) purified Pd^{2+} signaling product of **Res-DT** (**Res-DT** + Pd^{2+}), and (c) resorufin sodium salt in DMSO- d_6 . [**Res-DT**] = [resorufin sodium salt] = 10.0 mM. The spectrum of **Res-DT** + Pd^{2+} (b) was acquired after purifying the signaling product of **Res-DT** (10.0 mM) and $Pd(OAc)_2$ (25.0 mM) in CH_3CN .



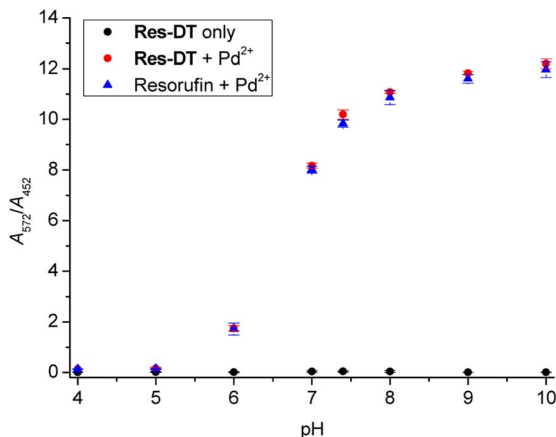


Fig. 5 Effect of pH on the Pd^{2+} signaling of **Res-DT**, represented by the absorbance ratio A_{572}/A_{452} . [**Res-DT**] = [resorufin] = $5.0 \mu\text{M}$, [Pd^{2+}] = $25 \mu\text{M}$, [SDS] = 10.0 mM , [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO. The pH was adjusted using 0.1 N HCl and NaOH.

suggested that the pH profile of the Pd^{2+} signaling was due to the pH dependency of the spectroscopic properties of resorufin. Next, to verify the signaling performance of **Res-DT** for Pd species with different oxidation states, the response of the probe to $\text{Pd}(\text{II})\text{OAc}_2$, $\text{Pd}(\text{II})\text{Cl}_2$, $\text{Pd}(\text{0})(\text{PPh}_3)_4$, and $\text{K}_2\text{Pd}(\text{IV})\text{Cl}_6$ was monitored. The results indicated that **Res-DT** exhibited similar signaling behavior, as evidenced by the absorbance ratios, for the different Pd species in the employed pH 7.4 PBS solution (Fig. S11, ESI[†]).

UV-vis titration of **Res-DT** with Pd^{2+} was subsequently performed to determine the minimum Pd^{2+} ion concentration detected using **Res-DT**. The absorbance ratio A_{572}/A_{452} increased linearly up to a Pd^{2+} concentration of $5.0 \mu\text{M}$ ($R^2 = 0.9987$) (Fig. 6). Using the titration plot and IUPAC recommended equation ($3s_{\text{blank}}/m$), where s_{blank} and m denote standard deviation of the blank signal and analytical sensitivity, respectively, the detection limit was

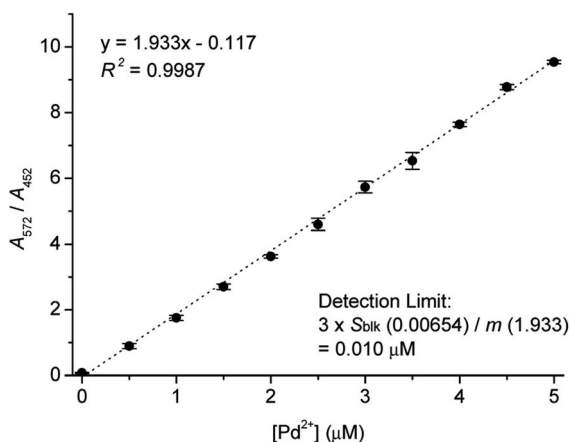


Fig. 6 Calibration curve for Pd^{2+} determination using the absorbance ratio A_{572}/A_{452} . [**Res-DT**] = $5.0 \mu\text{M}$, [Pd^{2+}] = $0-5.0 \mu\text{M}$, [SDS] = 10.0 mM , [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO.

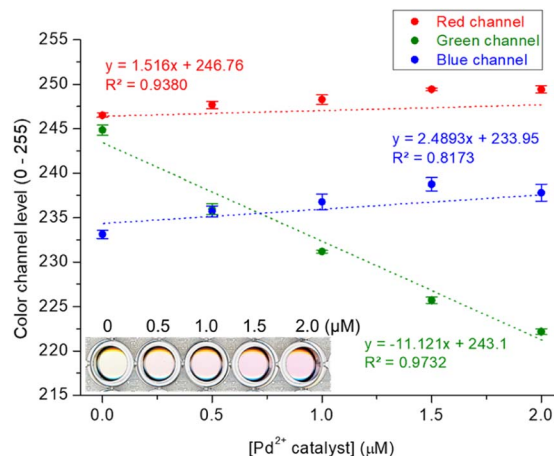


Fig. 7 Plot illustrating the color channel level changes (RGB) in response to a Pd-containing catalyst. Inset: images of solutions with different Pd^{2+} concentrations captured using a scanner. [**Res-DT**] = $5.0 \mu\text{M}$, [Pd^{2+} catalyst] = $0-2.0 \mu\text{M}$, [SDS] = 10.0 mM , [PBS pH 7.4] = 10.0 mM in an aqueous solution containing 2% (v/v) DMSO.

calculated to be 10 nM .⁴⁶ Additionally, we studied the quantitative analytical behavior of **Res-DT** for Pd^{2+} sensing by fluorescence titration (Fig. S12, ESI[†]). The fluorescence emission changes at 591 nm demonstrated a linear correlation with increasing concentrations of Pd^{2+} ($R^2 = 0.9870$). From these results, the detection limit for Pd^{2+} ions was calculated to be 7.3 nM .⁴⁶ Furthermore, we estimated the detection time of **Res-DT** for Pd^{2+} ions by observing the time-dependent change in the absorbance ratio (A_{572}/A_{452}), and it was found to be 7 min (Fig. S13, ESI[†]).

Finally, to assess the practical applicability of the devised probe, Pd assay in Pd-containing catalyst and drug candidate was performed using an office scanner as a readily accessible device for detection.⁴⁷ We used the white catalyst as the Pd-containing catalyst, and a Pd^{2+} -2-picolinic acid complex as the Pd-containing drug candidate.⁴⁸ The **Res-DT** with the tested catalyst and drug candidate exhibited a noticeable color shift from yellow to magenta, and the variation in color could be conveniently characterized through analysis of the RGB color channel levels of the scanned image (Fig. 7 and S14, ESI[†]). Consequently, acceptable calibration curves based on the green channel, rather than the red and blue channels, were obtained for the catalyst and drug candidate, which yielded satisfactory R^2 values of 0.9732 and 0.9887 for the catalyst and drug candidate, respectively. The results of the colorimetric signaling-based Pd^{2+} assay for the catalyst and drug candidate performed using the office scanner (recovery = $94.1-107.2\%$) were consistent with those of UV-vis spectrometry (recovery = $91.0-101.3\%$) (Table S3, ESI[†]). This result implies that the **Res-DT** could be successfully applied to the analysis of Pd^{2+} ions in palladium relevant catalysts and drugs.

4. Conclusions

A simple colorimetric and fluorescent probe (**Res-DT**) was developed for the convenient determination of Pd species in Pd-



containing catalyst and drug candidate. **Res-DT** exhibited selective, sensitive signaling behavior toward Pd species without interference from common metal ions, anions, and oxidants. The signaling was achieved by Pd-induced hydrolysis of the dithioate moiety of **Res-DT**, generating resorufin dye that exhibits a magenta color, visible to the naked eye, and intense fluorescence. The probe was immune to interference from several other metal ions and anions when detecting Pd ions. Additionally, the Pd signaling was achieved within 7 min, and the detection limit of the probe was determined to be 10 nM. Finally, **Res-DT** was successfully employed to analyze Pd²⁺ ions in Pd-containing catalyst and drug candidate. The designed **Res-DT** can potentially be applied in various practical and industrial settings featuring Pd-related systems.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

This research was supported by the Chung-Ang University Graduate Research Scholarship in 2022 (JH) and the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (No. 2022R1F1A1072643).

Notes and references

- 1 C. M. Crudden, M. Sateesh and R. Lewis, *J. Am. Chem. Soc.*, 2005, **127**, 10045–10050.
- 2 (a) X. F. Wu, P. Anbarasan, H. Neumann and M. Beller, *Angew. Chem., Int. Ed.*, 2010, **49**, 9047–9050; (b) C. C. C. J. Seechurn, M. O. Kitching, T. J. Colacot and V. Snieckus, *Angew. Chem., Int. Ed.*, 2012, **51**, 5062–5085.
- 3 N. P. Sudheesh, T. A. Ajith, K. K. Janardhanan and C. V. Krishnan, *Food Chem. Toxicol.*, 2010, **48**, 1858–1862.
- 4 E. Antolini, *Energy Environ. Sci.*, 2009, **2**, 915–931.
- 5 S. Bhaskaran, M. S. A. Padusha and A. M. Sajith, *ChemistrySelect*, 2020, **5**, 9005–9016.
- 6 I. Shinkai, A. O. King and R. D. Larsen, *Pure Appl. Chem.*, 1994, **66**, 1551–1556.
- 7 S. Haber, in *Aqueous-Phase Organometallic Catalysis*, ed. B. Cornils and W. A. Herrmann, Wiley-VCH, Weinheim, 1998.
- 8 U. Beutler, J. Mazacek, G. Penn, B. Schenkel and D. Wasmuth, *Chimia*, 1996, **50**, 154–156.
- 9 D. B. Damon, R. W. Dugger, S. E. Hubbs, J. M. Scott and R. W. Scott, *Org. Process Res. Dev.*, 2006, **10**, 472–480.
- 10 E. J. Lang, K. H. Lee, J. S. Lee and Y. G. Kim, *J. Mol. Catal. A: Chem.*, 1999, **138**, 25–36.
- 11 L. S. Lin, T. J. Lanza, J. J. P. Jewell, P. Liu, S. K. Shah, H. Qi, X. Tong, J. Wang, S. S. Xu, T. M. Fong, C.-P. Shen, J. Lao, J. C. Xiao, L. P. Shearman, D. S. Stribling, K. Rosko, A. Strack, D. J. Marsh, Y. Feng, S. Kumar, K. Samuel, W. Yin, L. V. der Ploeg, S. G. Mills, M. MacCoss, M. T. Goulet and W. K. Hagmann, *J. Med. Chem.*, 2006, **49**, 7584–7587.
- 12 C. Torborg and M. Beller, *Adv. Synth. Catal.*, 2009, **351**, 3027–3043.
- 13 A. S. Abu-Surrah, H. H. Al-Sa'doni and M. Y. Abdalla, *Cancer Ther.*, 2008, **6**, 1–10.
- 14 F. Zereini, C. Wiseman and W. Püttmann, *Environ. Sci. Technol.*, 2007, **41**, 451–456.
- 15 C. D. Spicer, T. Triemer and B. G. Davis, *J. Am. Chem. Soc.*, 2012, **134**, 800–803.
- 16 R. M. Yusop, A. Unciti-Broceta, E. M. V. Johansson, R. M. Sánchez-Martín and M. Bradley, *Nat. Chem.*, 2011, **3**, 239–243.
- 17 J. Kielhorna, C. Melberb, D. Kellerb and I. Mangelsdorfa, *Int. J. Hyg. Environ. Health*, 2002, **205**, 417–432.
- 18 H. Li, J. Fan and X. Peng, *Chem. Soc. Rev.*, 2013, **42**, 7943–7962.
- 19 R. Balamurugan, J.-H. Liu and B.-T. Liu, *Coord. Chem. Rev.*, 2018, **376**, 196–224.
- 20 X. Jin, J. Gao, T. Wang, W. Feng, R. Li, P. Xie, L. Si, H. Zhou and X. Zhang, *Spectrochim. Acta, Part A*, 2020, **224**, 117467.
- 21 M. P. Tracey, D. Pham and K. Koide, *Chem. Soc. Rev.*, 2015, **44**, 4769–4791.
- 22 P. Kumar, V. Kumar and R. Gupta, *RSC Adv.*, 2017, **7**, 7734–7741.
- 23 X. Chen, H. Wang, X. Ma, M. Wang, Y. Zhang, G. Gao, J. Liu and S. Hou, *Dyes Pigm.*, 2018, **148**, 286–291.
- 24 G. Wu, Z. Wang, W. Zhang, W. Chen, X. Jin and H. Lu, *Inorg. Chem. Commun.*, 2019, **102**, 233–239.
- 25 X. Fang, Y. Zhang, M. Li, Z. Zhang, Y. Qi, X. Zhang, X. Zhang, Y. Liu, J. Li and H. Yu, *Dyes Pigm.*, 2023, **209**, 110929.
- 26 M. E. Jun, B. Roy and K. H. Ahn, *Chem. Commun.*, 2011, **47**, 7583–7601.
- 27 (a) M. Du, Y. Zhang, Y. Yu, H. Zhao, Y. Guo and Y. Yang, *Anal. Methods*, 2019, **11**, 6053–6061; (b) Y. Zhang, M. Yang and M. Ji, *New J. Chem.*, 2020, **44**, 20434.
- 28 (a) W. Luo, J. Li and W. Liu, *Org. Biomol. Chem.*, 2017, **15**, 5846–5850; (b) Q. Xia, S. Feng, D. Liu and G. Feng, *Sens. Actuators, B*, 2018, **258**, 98–104.
- 29 M. Kumar, N. Kumar and V. Bhalla, *RSC Adv.*, 2013, **3**, 1097–1102.
- 30 J. Zhou, S. Xu, Z. Yu, X. Ye, X. Dong and W. Zhao, *Dyes Pigm.*, 2019, **170**, 107656.
- 31 T. Chen, T. Wei, Z. Zhang, Y. Chen, J. Qiang, F. Wang and X. Chen, *Dyes Pigm.*, 2017, **140**, 392–398.
- 32 W. Liu, J. Jiang, C. Chen, X. Tang, J. Shi, P. Zhang, K. Zhang, Z. Li, W. Dou, L. Yang and W. Liu, *Inorg. Chem.*, 2014, **53**, 12590–12594.
- 33 X. Li, H. Huang, Y. Zhu, H. Zhao and Z. Wang, *RSC Adv.*, 2015, **5**, 105810–105813.
- 34 A. Higashi, N. Kishikawa, K. Ohyama and N. Kuroda, *Tetrahedron Lett.*, 2017, **58**, 2774–2778.
- 35 M. E. Jun and K. H. Ahn, *Org. Lett.*, 2010, **12**, 2790–2793.
- 36 (a) M. G. Choi, J.-Y. Seo, E. J. Cho and S.-K. Chang, *J. Photochem. Photobiol., A*, 2022, **429**, 113920; (b) A. Ghosh, S. Nandi, A. Sengupta, A. Chattopadhyay, S. Lohar and D. Das, *Inorg. Chim. Acta*, 2015, **436**, 52–56.
- 37 (a) W. Feng, L. Bai, S. Jia and G. Feng, *Sens. Actuators, B*, 2018, **260**, 554–562; (b) L. Wang, M. Ren, Z. Li, L. Dai and



- W. Lin, *New J. Chem.*, 2019, **43**, 552–555; (c) S. Mondal, S. K. Manna, S. Pathak, A. Al Masum and S. Mukhopadhyay, *New J. Chem.*, 2019, **43**, 3513–3519; (d) F.-K. Tang, S.-M. Chan, T. Wang, C.-S. Kwan, R. Huang, Z. Cai and K. C.-F. Leung, *Talanta*, 2020, **210**, 120634.
- 38 M. G. Choi, Y. J. Lee, K. M. Lee, K. Y. Park, T. J. Park and S.-K. Chang, *Analyst*, 2019, **144**, 7263–7269.
- 39 (a) L. Qiao, H. Nie, Y. Wu, F. Xin, C. Gao, J. Jing and X. Zhang, *J. Mater. Chem. B*, 2017, **5**, 525–530; (b) L. Wu, Q. Yang, L. Liu, A. C. Sedgwick, A. J. Cresswell, S. D. Bull, C. Huang and T. D. James, *Chem. Commun.*, 2018, **54**, 8522–8525.
- 40 M. G. Choi, S. Y. Park, K. Y. Park and S.-K. Chang, *Sci. Rep.*, 2019, **9**, 3348.
- 41 F. C. Lopez, A. Shankar, M. Thompson, B. Shealy, D. Locklear, T. Rawalpally, T. Cleary and C. Gagliardi, *Org. Process Res. Dev.*, 2005, **9**, 1003–1008.
- 42 L. Tian, H. Feng, Z. Dai and R. Zhang, *J. Mater. Chem. B*, 2021, **9**, 53–79.
- 43 M. G. Choi, S. Kwon and S.-K. Chang, *Dyes Pigm.*, 2021, **192**, 109394.
- 44 J. S. Kim, M. G. Choi, K. C. Song, K. T. No, S. Ahn and S.-K. Chang, *Org. Lett.*, 2007, **9**, 1129–1132.
- 45 B. J. Stenton, B. L. Oliveira, M. J. Matos, L. Sinatra and G. J. L. Bernardes, *Chem. Sci.*, 2018, **9**, 4185–4189.
- 46 D. C. Harris, in *Quantitative Chemical Analysis*, W.H. Freeman and Company, New York, 8th edn, 2010, pp. 103–105.
- 47 D. C. Christodouleas, A. Nemiroski, A. A. Kumar and G. M. Whitesides, *Anal. Chem.*, 2015, **87**, 9170–9178.
- 48 F. A. Al-Saif, J. Y. Al-Humaidi, D. N. Binjawhar and M. S. Refat, *J. Mol. Struct.*, 2020, **1218**, 128547.

