Research Report

Genetic Screening in Korean Patients with Frontotemporal Dementia Syndrome

Eun-Joo Kim^a, Duk L. Na^b, Hee-Jin Kim^b, Kyung Won Park^c, Jae-Hong Lee^d, Jee Hoon Roh^{e,f}, Jay C. Kwon^g, Soo Jin Yoon^h, Na-Yeon Jungⁱ, Jee Hyang Jeong^j, Jae-Won Jang^k, Hee-Jin Kim^l, Kee Hyung Park^m, Seong Hye Choiⁿ, Sang Yun Kim^o, Young Ho Park^o, Byeong C. Kim^p, Young Chul Youn^q, Chang-Seok Ki^r, Seung Hyun Kim¹, Sang Won Seo^{b,*,1} and Young-Eun Kim^{s,*,1} ^aDepartment of Neurology, Pusan National University Hospital, Pusan National University School of Medicine and Medical Research Institute, Busan, Republic of Korea ^bDepartment of Neurology, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Republic of Korea ^cDepartment of Neurology, Dong-A Medical Center, Dong-A University College of Medicine, Busan, Republic of Korea ^dDepartment of Neurology, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Republic of Korea ^eDepartment of Biomedical Sciences and Department of Physiology, Korea University College of Medicine, Seoul, Republic of Korea ^fDepartment of Neurology, Korea University Anam Hospital, Seoul, Republic of Korea ^gDepartment of Neurology, Changwon Fatima Hospital, Changwon, Republic of Korea ^hDepartment of Neurology, Eulji University Hospital, Daejeon, Republic of Korea ¹Department of Neurology, Pusan National University Yangsan Hospital, Research Institute for Convergence of Biomedical Science and Technology, Yangsan, Republic of Korea ^JDepartment of Neurology, Ewha Womans University Seoul Hospital, Seoul, Republic of Korea ^kDepartment of Neurology, Kangwon National University Hospital, Kangwon National University College of Medicine, Chuncheon, Republic of Korea ¹Department of Neurology, Hanyang University College of Medicine, Seoul, Republic of Korea ^mDepartment of Neurology, Gachon University Gil Hospital, Incheon, Republic of Korea ⁿDepartment of Neurology, Inha University School of Medicine, Incheon, Republic of Korea ^oDepartment of Neurology, Seoul National University College of Medicine and Clinical Neuroscience Center, Seoul National University Bundang Hospital, Gyeonggi-do, Republic of Korea ^pDepartment of Neurology, Chonnam National University Medical School, Gwangju, Republic of Korea ^qDepartment of Neurology, Chung-Ang University Hospital, Chung-Ang University College of Medicine, Seoul, Republic of Korea ^rGC Genome, Yongin, Gyeonggi-do, Republic of Korea ^sDepartment of Laboratory Medicine, Hanyang University College of Medicine, Seoul, Republic of Korea

Received 5 May 2022 Accepted 4 September 2022 Pre-press 22 September 2022 Published 21 October 2022

*Correspondence to: Young-Eun Kim, MD, Department of Laboratory Medicine, Hanyang university College of Medicine, 222, Wangsimni-ro, Seongdong-gu, Seoul, 04763, Republic of Korea. Tel.: +82 02 2220 1841; Fax: +82 02 2220 0699; E-mail: young0eun@hanyang.ac.kr and Sang Won Seo, MD, Department

of Neurology, Samsung Medical Center, Sungkyunkwan University School of Medicine, Tel.: +82 02 3410 1233; Fax: +82 02 3410 0052; E-mail: sangwonseo@empal.com.

¹These authors contributed equally to this work.

ISSN 2542-4823 © 2022 – The authors. Published by IOS Press. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (CC BY-NC 4.0).

Abstract.

Background: Frontotemporal dementia (FTD) syndrome is a genetically heterogeneous group of diseases. Pathogenic variants in the chromosome 9 open reading frame 72 (*C9orf72*), microtubule-associated protein tau (*MAPT*), and progranulin (*GRN*) genes are mainly associated with genetic FTD in Caucasian populations.

Objective: To understand the genetic background of Korean patients with FTD syndrome.

Methods: We searched for pathogenic variants of 52 genes related to FTD, amyotrophic lateral sclerosis, familial Alzheimer's disease, and other dementias, and hexanucleotide repeats of the *C9orf72* gene in 72 Korean patients with FTD using whole exome sequencing and the repeat-primed polymerase chain reaction, respectively.

Results: One likely pathogenic variant, p.G706R of *MAPT*, in a patient with behavioral variant FTD (bvFTD) and 13 variants of uncertain significance (VUSs) in nine patients with FTD were identified. Of these VUSs, M232R of the *PRNP* gene, whose role in pathogenicity is controversial, was also found in two patients with bvFTD.

Conclusions: These results indicate that known pathogenic variants of the three main FTD genes (*MAPT*, *GRN*, and *C9orf72*) in Western countries are rare in Korean FTD patients.

Keywords: Frontotemporal dementia, MAPT, next-generation sequencing, PRNP

INTRODUCTION

Frontotemporal dementia (FTD) is the second most common type of early onset dementia and is characterized by progressive deterioration of behavior or language associated with frontal or temporal degeneration. It comprises of three clinical phenotypes: behavioral variant FTD (bvFTD), semantic variant primary progressive aphasia (svPPA), and non-fluent/agrammatic variant PPA (nfvPPA), which occasionally overlaps with motor neuron disease or atypical parkinsonian syndromes, such as progressive supranuclear palsy syndrome and corticobasal syndrome [1]. Although FTD is highly heritable in Western countries [2-4], genetic FTD is rare in Asian populations [5, 6]. We previously identified a single known pathogenic variant in the progranulin (GRN) gene and two novel variants in the colony stimulating factor 1 receptor (CSF1R) and alanyl-tRNA synthetase 2 (AARS2) genes in 107 Korean patients with sporadic FTD using next-generation sequencing (NGS) [6]. To elucidate the genetic characteristics of FTD in Asian population, large-scale, unbiased, and in-depth genetic screening using NGS technologies is continuously required.

In this study, we performed whole exome sequencing (WES) of 72 Korean patients with clinical FTD syndrome, without regarding familial status, to identify the presence of pathogenic variants of FTD, amyotrophic lateral sclerosis (ALS), or other dementia-related genes.

MATERIALS AND METHODS

Patients

Patients were prospectively recruited from ten neurology clinics across Korea between June 2016 and January 2018. All patients that were enrolled in this study met the FTD criteria proposed by Knopman et al. [7] or the new consensus diagnostic criteria for bvFTD [8], svPPA, and nfvPPA [9]. Patients with clinical and electrophysiological evidence of ALS were enrolled as having FTD-ALS, regardless of the clinical subtype of FTD. This study was conducted as part of the Clinical Research Center for Dementia of South Korea-FTD (CREDOS-FTD) registry study, which was carried out between 2010 and 2018 [10]. All participants were registered in the CREDOS-FTD registry and evaluated using the CREDOS-FTD protocol. The protocol is composed of a clinical evaluation form (clinical history, neurological examination. Korean version of Mini-Mental State Examination, Hachinski ischemic scale, Global Deterioration Scale, Unified Parkinson's Disease Rating Scale, and Frontotemporal-Clinical Dementia Rating sum of boxes score), caregiver questionnaire form (Korean dementia screening questionnaire, Barthel Activities of Daily Living, Seoul Instrumental Activities of Daily Living, and Caregiver-Administered Neuropsychiatric Inventory and Frontal Behavioral Inventory), and cognitive test battery for FTD, consisting of subdomains assessing attention, language,

visuospatial, memory, and frontal/executive functions [10]. All patients underwent brain magnetic resonance imaging (MRI) to exclude any structural lesions. Patients with current or past neurological or psychiatric illnesses were excluded from this study.

The institutional review boards (IRB) at all participating centers approved the study (IRB No. 2012-12), and informed consent was obtained from each patient and caregiver. As informed consent did not include open access to the data used in the study, our data is not publicly available.

Genetic analysis

Genomic DNA was extracted from peripheral blood leukocytes using a standard procedure. WES was performed using an Agilent SureSelect All Exon kit 50 Mb (Agilent Technologies, Santa Clara, CA, USA) on a NextSeq 500 platform (Illumina Inc., San Diego, CA, USA). Alignment of sequence reads, indexing of the reference genome (GRCh37/hg19), and variant calling were performed using a pipeline based on GATK Best Practices. Variants with allele frequencies >0.001 were filtered out based on public databases, including the Genome Aggregation Database (http://gnomad.broadinstitute.org/) and 1,722 ethnically matched controls from the Korean Reference Genome Database (KRGDB) [11]. Thereafter, 32 genes related to ALS-FTD and 20 genes related to familial Alzheimer's disease and other dementias that have been explored by the authors from OMIM (https://www.omim.org/), GeneReview (http://www.ncbi.nlm.nih.gov/books/NBK1116/),

and high-quality published literature [1, 4] and introduced as causative genes of FTD-ALS spectrum disorders or dementia, were screened for pathogenic variants (Table 1). Missense, frameshift, indel, nonsense, synonymous, and intronic variants within the exon-flaking regions were also evaluated. Splice site and conservation analyses were performed for all synonymous variants using Human Splice Finder and GERP scores. All amino acid variants were confirmed using Sanger sequencing or DeepVariant (https://github.com/google/deepvariant, version 1.2.0). The variants were classified according to the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) guidelines [12]. The BP7 rule of the ACMG/AMP guidelines was only applied in cases that were being predicted as 'no impact and not highly conserved'. Hexanucleotide repeat expansion of chromosome 9 open reading frame

72 (*C9orf72*) was tested for all patients using the triplet repeat-primed polymerase chain reaction, as previously described [13].

RESULTS

Demographic and clinical findings

A total of 72 patients (35 males and 37 females) with FTD were enrolled, including 38 with bvFTD, 26 with svPPA, six with nfvPPA, and two with FTD-ALS. One patient with FTD-ALS presented with abnormal behavior and the other presented with dysarthria followed by behavioral changes. The mean age was 65.8 ± 10.3 years, and the mean age at disease onset was 63.9 ± 11.5 years. The mean interval between disease onset and enrollment into the study was 3.6 ± 2.3 years. Seventeen patients (23.6%) had a history of dementia or neuropsychiatric disease in a first-degree relative. The detailed demographic data is summarized in Table 2. During the preparation of this manuscript, a parallel study on telomere length in FTD syndrome was published by our group [14]. Of the 72 participants, 40 patients with sufficient DNA available for the Southern blotting analysis for telomere length after WES, were included in the parallel study.

Genetic findings

WES yielded an average read depth of 121.81x and the average for $10 \times$ coverage was 99.33%. After variant analysis, one likely pathogenic variant in the MAPT gene, c.2116G>A (p.G706R), was identified in a patient with bvFTD (FTD-18), which has been reported in familial or sporadic FTD [15]. FTD-18 had H1/H1 haplotypes of the MAPT gene and had been enrolled in an FTD modeling study using induced pluripotent stem cell (iPSC) technology and had been reported elsewhere [16]. The four variants of uncertain significance (VUSs) from ALS-FTD-related genes were also detected in four patients with FTD: PRNP gene, c.695T>G (p.M232R) in two patients with bvFTD (FTD-13, FTD-39), VCP gene, c.278G>T (p.R93L) in a patient with svPPA, and UBQLN2 gene, c.20G>A (p.S7N) in a patient with bvFTD (Table 3). Of these, p.M232R of the PRNP gene has been reported previously in Creutzfeldt-Jacob disease (CJD) [17], but its pathogenicity has been controversial. p.R93L of the VCP gene and p.S7N of the UBQLN2 gene were novel variants. None of the patients had abnormal repeat

Gene symbol	RefSeq	Gene description	Chromosomal location	Inheritance
FTD and ALS-relate	ed genes			
ALS2	NM_020919.3	Amyotrophic lateral sclerosis 2	2q33.1	AD
ANG	NM_001145.4	Angiogenin, ribonuclease, RNase A family, 5	14q11.1-q11.2	AD
CCNF	NM_001761.3	Cyclin F	16p13.3	AD
CHCHD10	NM_213720.3	Coiled-coil-helix-coiled-coil-helix domain-containing protein 10	22q11.23	AD
CHMP2B	NM_014043.3	Chromatin modifying protein 2B	3p11.2	AD
CHRNA4	NM_000744.6	Acetylcholine receptor, neuronal nicotinic, alpha-4 subunit	20q13.2-q13.3	AD
CYLD	NM_015247.3	CYLD lysine 63 deubiquitinase	16q12.1	AD
DAO	NM_001917.4	D-amino-acid oxidase	12q24	AD
DCTN1	NM_004082.4	Dynactin 1	2p13	AD
FIG4	NM_014845.5	FIG4 phosphoinositide 5-phosphatase	6q21	AR
FUS	NM_004960.3	FUS RNA binding protein	16p11.2	AD
GRN	NM_002087.2	Granulin	17q21.32	AD
HNRNPA1	NM_031157.2	Heterogeneous nuclear ribonucleoprotein A1	12q13.1	AD
HNRNPA2B1	NM_031243.2	Heterogeneous nuclear ribonucleoprotein A2/B1	7p15	AD
MAPT	NM_005910.5	Microtubule-associated protein tau	17q21.1	AD
MATR3	NM_199189.2	Matrin3	5q31.2	AD
OPTN	NM_021980.4	Optineurin	10p13	AD
PRNP	NM_000311.3	Prion protein	20p13	AD
SETX	NM_015046.5	Senataxin	9q34.13	AD/AR
SIGMAR1	NM_005866.2	Sigma non-opioid intracellular receptor 1	9p13.3	AD/AR
SOD1	NM_000454.4	Superoxide dismutase 1	21q22.11	AD
SPG11	NM_025137.3	SPG11, spatacsin vesicle trafficking associated	15q14	AR
SQSTM1	NM_003900.4	Sequestosome 1	5q35	AD
TAF15	NM_139215.2	TATA-box binding protein associated factor 15	17q11.1-q11.2	AD
TARDBP	NM_007375.3	TAR DNA binding protein	1p36.22	AD
TBK1	NM_013254.3	Tank-binding kinase 1	12q14.2	AD
TIA1	NM_022173.4	TIA1 cytotoxic granule-associated RNA-binding protein	2p13.3	AD
TREM2	NM_018965.2	Triggering receptor expressed on myeloid cells 2	6p21.1	AR

 Table 1

 List of frontotemporal dementia (FTD), amyotrophic lateral sclerosis (ALS), familial Alzheimer's disease and other dementia-related genes

TUBA4A	NM_006000.3	Tubulin alpha 4a	2q35	AD
UBQLN2	NM_013444.3	Ubiquilin 2	Xp11.21	XL
VAPB	NM_004738.4	VAMP (vesicle-associated membrane protein)-associated protein B and C	20q13.33	AD
VCP	NM_007126.3	Valosin-containing protein	9p13.3	AD
Genes of familia	al Alzheimer's disease			
APP	NM_000484.3	Amyloid beta precursor protein	21q21.2	AD
PSEN1	NM_000021.3	Presenilin-1 (alzheimer disease 3)	14q24.3	AD
PSEN2	NM_000447.2	Presenilin-2 (alzheimer disease 4)	1q31-q42	AD
Other dementia-	-related genes			
AARS2	NM_020745.3	Alanyl-tRNA synthetase 2, mitochondrial	6p21.1	AR
ABCD1	NM_000033.3	ATP binding cassette subfamily D member 1	Xq28	XL
ARSA	NM_000487.5	Arylsulfatase A	22q13.33	AR
CSF1R	NM_005211.3	Colony-stimulating factor 1 receptor	5q32	AD
DARS2	NM_018122.4	Aspartyl-tRNA synthetase 2, mitochondrial	1q25.1	AR
EIF2B1	NM_001414.3	Eukaryotic translation initiation factor 2B subunit alpha	12q24.31	AR
EIF2B2	NM_014239.3	Eukaryotic translation initiation factor 2B subunit beta	14q24.3	AR
EIF2B3	NM_020365.4	Eukaryotic translation initiation factor 2B subunit gamma	1p34.1	AR
EIF2B4	NM_015636.3	Eukaryotic translation initiation factor 2B subunit delta	2p23.3	AR
EIF2B5	NM_003907.2	Eukaryotic translation initiation factor 2B subunit epsilon	3q27.1	AR
GALC	NM_000153.3	Galactosylceramidase	14q31.3	AR
GBA	NM_001005741.2	Glucosidase, beta acid	1q21	AD, susceptibility
GLA	NM_000169.2	Galactosidase alpha	Xq22.1	XL
ITM2B	NM_021999.5	Integral membrane protein 2B	13q14.2	AD
NOTCH3	NM_000435.2	Notch 3	19q13.12	AD
SNCB	NM_001001502.1	Synuclein, beta	5q35	AD
TYROBP	NM_003332.3	TYRO protein tyrosine kinase binding protein	19q13.12	AR

AD, autosomal dominant; AR, autosomal recessive; XL, X-linked.

Demographics of patients								
	Total	bvFTD	svPPA	nfvPPA	FTD-ALS			
Number	72	38	26	6	2			
Age (y)	65.8 ± 10.3	64.3 ± 11.5	67.3 ± 7.1	69.8 ± 11.9	61.0 ± 21.2			
Onset Age (y)	61.9 ± 10.6	60.5 ± 12.0	63.3 ± 7.3	66.2 ± 13.4	60.5 ± 21.9			
Sex (M:F)	35:37	20:18	12:14	2:4	1:1			
Interval between onset and enrollment (y)	3.6 ± 2.3	3.6 ± 2.0	4.0 ± 2.8	2.7 ± 2.0	0.6 ± 0.6			
FTD-CDR (SB)	7.5 ± 5.2	8.0 ± 5.5	7.5 ± 5.2	6.0 ± 3.9	4.3 ± 3.2			
MMSE	17.2 ± 9.1	18.5 ± 8.0	15.0 ± 10.7	16.2 ± 8.9	25.5 ± 4.9			
Family history*	23.6% (17/72)	23.7% (9/38)	23.1% (6/26)	33.3% (2/6)	0% (0/2)			

Table	2
Demographics	of patients

ALS, amyotrophic lateral sclerosis; bvFTD, behavioral variant frontotemporal dementia; CDR, Clinical Dementia Rating; F, female; MMSE, Mini-Mental State Examination; M, male; nfvPPA, non-fluent/agrammatic variant primary progressive aphasia; SB, sum of boxes; svPPA, semantic variant primary progressive aphasia. *Family history of dementia or neuropsychiatric disease in first-degree relatives.

expansions of C9orf72. Regarding genes related to familial Alzheimer's disease and other dementias, nine VUSs were identified from seven patients and one of them, p.L136S of the APP gene, was novel (Table 3).

Case carrying p.G706R in the MAPT gene

FTD-18 was a 32-year-old man with a three-year history of progressive behavioral changes and cognitive deficits. His symptoms included impatience, aggression, and emotional blunting. The patient became more inert and depressed with time. He also exhibited hypersexuality and an obsession with sweet foods and computer games. Episodic memory impairment, visuospatial dysfunction, language difficulties, and decline in personal hygiene and activities of daily living developed around the time of his first visit to the neurology clinic. His maternal grandfather had dementia at the age of 60. Neurological examinations were unremarkable. Neuropsychological evaluation revealed severely impaired memory, visuospatial, language, and executive functions. The MMSE score was 20 and the clinical dementia rating was 1. Brain MRI demonstrated bilateral and symmetric frontal atrophy (Fig. 1A). Florbetaben and flortaucipir PET revealed negative findings. The clinical syndromic diagnosis was bvFTD. His symptoms progressed rapidly, and two years after the initial evaluation, he was transferred to another hospital.

Cases carrying p.M232R in the PRNP gene

FTD-13 was a 54-year-old man who worked as a construction manager. He was admitted to the psychiatric ward with rapidly progressive behavioral changes and memory impairment for six months. He repeatedly called friends to borrow money, although he was financially stable; he could not wait his turn at restaurants for food to be put on his plate; he obsessively called his son, until he convinced him to have lunch together; he woke his son up at 2 am to visit his parents' grave and repeatedly asked the same questions. However, he was able to maintain normal daily living activities. Neurological examinations showed normal findings. The MMSE score was 26 and the global deterioration score (GDS) was 3. Neuropsychological test results showed impaired naming, memory, and frontal function. The patient had no family history of dementia. Initial brain MRI revealed prominent atrophy in bilateral frontal lobes. Diffusion-restricted areas were not detected on diffusion-weighted imaging (DWI). The follow-up MRIs, taken at one-month intervals, showed the same findings as the previous ones. Fluorodeoxyglucose-positron emission tomography (FDG-PET) demonstrated severe glucose hypometabolism in the bilateral frontal areas, which worsened on the right side (Fig. 1B). He was diagnosed with bvFTD and transferred to another hospital after six months of follow-ups.

FTD-39 was a 51-year-old man with a two-year history of behavioral changes. He was eating 15 meals per day due to increased appetite and gained 22 kg in a year; he went to the bathroom frequently, over 15 times a day; he provided short answers to questions and refused to take a bath; he showed poor spontaneity and sometimes talked to himself. Memory impairment, visuospatial dysfunction, language difficulties, and disorientation also developed in the patient. There was no family history of dementia. Neurological examinations did not reveal any significant changes. Initial MMSE score was 24, and the GDS was 4. However, brain MRI revealed bilateral frontotemporal atrophy, which worsened on the left side. Initially, he was diagnosed with schizophrenia

Patient	Gene	Reference	Nucleotide	Amino acid	ClinVar	rs number	Allele frequency		In-silico analysis			
ID		sequence	change	change			$gnomAD^{\dagger}$	KRGDB [‡]	Poly Phen-2	SIFT	Mutation Taster	CADD§
ALS-FTD re	lated genes											
FTD-13	PRNP	NM_000311.3	c.695T>G	p.Met232Arg	VUS	rs74315409	0.0009	0.0040	В	D	Р	<20
FTD-39	PRNP	NM_000311.3	c.695T>G	p.Met232Arg	VUS	rs74315409	0.0009	0.0040	В	D	Р	<20
FTD-36	VCP	NM_007126.3	c.278G>T	p.Arg93Leu	N/A	N/A	0	0	D	D	DC	29.90
FTD-48	UBQLN2	NM_013444.3	c.20G>A	p.Ser7Asn	N/A	N/A	0	0	В	Т	Р	<20
Genes of fan	nilial Alzheime	er's disease										
FTD-48	PSEN2	NM_000447.3	c.1262C>T	p.Thr421Met	VUS	rs756609078	0.0001	0	PD	Т	DC	34.00
FTD-61	APP	NM_000484.4	c.407T>C	p.Leu136Ser	N/A	N/A	0	0	PD	D	DC	25.10
Other demen	tia related gen	es										
FTD-41	CSF1R	NM_005211.3	c.110C>T	p.Thr37Met	B*	rs139635308	0.0004	0	PD	Т	Р	<20
FTD-19	GBA	NM_000157.4	c.902G>A	p.Arg301His	N/A	rs140955685	0.0003	0.0009	В	Т	Р	<20
FTD-39	ITB2B	NM_021999.4	c.454-3del	-	N/A	rs747826043	< 0.0001	0.0009	N/A	N/A	N/A	N/A
FTD-71	ITB2B	NM_021999.4	c.454-3del	-	N/A	rs747826043	< 0.0001	0.0009	N/A	N/A	N/A	N/A
FTD-32	NOTCH3	NM_000435.2	c.5336G>T	p.Gly1779Val	N/A	rs771041592	0.0001	0.0009	В	Т	DC	24.80
FTD-39	NOTCH3	NM_000435.2	c.6097C>A	p.Pro2033Thr	VUS	rs375213868	<0.0001	0	PD	D	DC	23.80
FTD-48	GALC	NM_000153.4	c.1912G>A	p.Gly638Ser	VUS	rs769851272	0.0009	0.0009	PD	D	DC	25.10

 Table 3

 Variants of uncertain significance in ALS-FTD, familial Alzheimer's disease, and other dementia related genes

N/A, not applicable; VUS, variant of uncertain significance; PD, probably damaging; B, benign; D, deleterious; T, tolerable; DC, disease causing; P, polymorphism; CADD, Combined Annotation Dependent Depletion. *The variant was submitted by single institute without criteria (accessed on 28 July 2022). †gnomAD, gnome Aggregation Database (http://gnomad.broadinstitute.org/). ‡KRGDB, the Korean Reference Genome Database [11]. [§]The variants over the CADD score 20 are presented in bold. The variants were classified according to the guideline of ACMG [12].



Fig. 1. A) Brain MRIs of FTD-18 who carried variant p.G706R of the *MAPT* gene, revealing bilateral and symmetric frontal atrophy. B) Brain MRIs and FDG-PET images of FTD-13 who carried variant M232R of the *PRNP* gene, showing cortical atrophy and severe glucose hypometabolism in the bifrontal areas. C) Brain MRIs of FTD-39 who carried variant M232R of the *PRNP* gene, demonstrating prominent bilateral frontotemporal atrophy, worse on the left.

in a psychiatric clinic and was referred to a neurology clinic where he was diagnosed with bvFTD. The 9month follow-up MMSE score was 17, and the GDS was 5. Follow-up MRIs showed aggravated cortical atrophy in both frontal and temporal areas (Fig. 1C). After almost three years of follow-ups, the patient stopped visiting the hospital.

Cases carrying novel variants (p.R93L of the VCP gene, p.S7N of the UBQLN2 gene, and p.L136S of the APP gene)

FTD-36 with p.R93L of the VCP gene was a 50year-old man, whose illness began at the age of 45; he presented memory impairment, followed by fluent aphasia, disinhibition, and myoclonic seizures. Brain MRI showed severe bi-frontotemporal atrophy, worse on the left side, and left parietal atrophy (Fig. 2A). Electroencephalography revealed partial seizures arising in both frontal areas. A clinical syndromic diagnosis of svPPA was made. His mother had a stroke. FTD-48 with p.S7N of the *UBQLN2* gene was a 59-year-old woman, whose personality changes including apathy, indifference, loss of empathy, obsession, and disinhibition started at the age of 57. In contrast to the behavioral changes, her memory, language, and visuospatial functions were relatively preserved and her *APOE* genotype was $\varepsilon 3/\varepsilon 4$. Brain



Fig. 2. A) Brain MRIs of FTD-36 who carried variant p.R93L of the VCP gene, showing bi-frontotemporal atrophy (worse on the left) and left parietal atrophy. B) Brain MRIs of FTD-48 who carried variant p.S7N of the UBQLN2 gene, variant p.T421M of the PSEN2 gene and variant p.G638S of the GALC gene, revealing right asymmetric frontotemporal atrophy. C) Brain MRIs of FTD-61 who carried variant p.L136S of the APP gene, demonstrating prominent atrophy in the bifrontal and left temporal area.

MRI revealed right asymmetric frontotemporal atrophy (Fig. 2B). The clinical syndromic diagnosis was bvFTD. The patient's family history was unremarkable. This patient carried two more VUS: p.T421M of the PSEN2 gene and p.G638S of the GALC gene. The p.T421M of *PSEN2* has been reported in an early onset sporadic AD patient, carrying APOE $\varepsilon 4/\varepsilon 4$, from Japan [18]. FTD-61 with p.L136S of the APP gene was a 66-year-old man whose illness began at the age of 58; he presented with apathy, disinhibition, and hyperorality, followed by right-side rigidity and gait disturbances at the age of 60. A brain MRI demonstrated prominent atrophy in the bifrontal and left temporal areas (Fig. 2C). A clinical syndromic diagnosis of bvFTD was made. Notably, his father had Parkinson's disease.

DISCUSSION

Pathogenic variants detected in common FTD genes, such as *MAPT*, *GRN*, and *C9orf72*, are gen-

erally rare in Asian populations. Our first study screening these in 75 Korean patients with sporadic FTD and the subsequent testing on multiple genes using NGS in 107 Korean patients with FTD confirmed the ethnic or geographical variability of the mutations in known FTD genes [5, 6]. In the present study, which is in accordance with previous studies, we identified only one patient with bvFTD harboring p.G706R in the *MAPT* gene.

The p.G706R variant, traditionally known as the G389R mutation in the *MAPT* gene, has been associated with rapidly progressive young-onset FTD, which was similar to those of our FTD-18 [15, 19–21]. This variant revealed the possibility of incomplete penetrance based on a lack of autosomal dominant inheritance patterns or unaffected mutation carriers, which was also observed in FTD-18, whose parents were clinically normal [19–22]. *In vitro* study, the p.G706R variant altered the affinity of tau to microtubules and decreased its ability to enhance microtubule assembly [15, 23]. *MAPT* p.P513A and p.L266V variants have recently been observed in two

Korean patients with early onset Alzheimer's disease and nfvPPA, respectively [24, 25].

Of the 13 VUSs we found in this study, the p.M232R variant of the PRNP gene is one of the five most frequent CJD mutations in Japan [26] and has mostly been reported in Asian populations [17, 27, 28]. The clinical characteristics of p.M232R are similar to those of sporadic CJD, including progressive dementia, 14-3-3 protein positivity, DWI hyperintensity, and no family history [26]. Previously, reported Korean cases of p.M232R were all suspected to be sporadic CJD based on their clinical symptoms at the time of diagnosis [29]. Notably, FTD-13 and FTD-39 in this study presented with frontal behavioral abnormalities associated with prominent bifrontal atrophy or hypometabolism, leading to the clinical diagnosis of bvFTD. Although the clinical course of FTD-13 and FTD-39 progressed somewhat rapidly, all DWIs repeatedly performed at one-month intervals were negative. Since the DWI negative p.M232R cases were presented recently [30], it is possible that both patients might have developed myoclonus, which is a typical feature of CJD, or demonstrated periodic sharp and wave complexes on EEG or positive DWIs later on.

As mentioned earlier, some researchers are concerned about the questionable pathogenicity of p.M232R since it has also been detected in healthy controls or non-CJD patients [31]. In addition to this, the allele frequency of the variant was 0.0008 in the East Asian cohort of gnomAD and 0.0040 in the ethnic-matched control of KRGDB, which is higher than the annual incidence rate of prion disease (0.85 per million population) [26]. Thus, pathological confirmation is required to resolve whether the p.M232R variant found in our patients with bvFTD is pathogenic or rare.

A recently published large international study of genetic FTD showed that approximately 25–30% of patients with FTD syndrome harbored pathogenic variants of 40% in *C9orf72*, 35% in *GRN*, 25% in *MAPT*, and only 1-2% in other genes [32]. Another study from the North American FTD cohort found 31 different pathogenic variants within the three main FTD genes in 223 of 302 sporadic and 390 familial participants (32.2%) [3]. However, our group has only identified two pathogenic variants in the three main FTD genes (one for *MAPT* and one for *GRN*) through CREDOS-FTD genetic studies (approximately 1%) [5, 6]. Given the extreme rarity of genetic FTD in Korea, genetic screening of sporadic and familial FTD through a longitudinal Korean cohort is needed

to better understand the geographical or ethnic variability of genetic FTD. Furthermore, collaborating with worldwide genetic FTD cohorts would encourage the development of powerful biomarkers and construct trial-ready cohorts for new FTD syndrome therapies, eventually leading to its prevention.

ACKNOWLEDGMENTS

The authors have no acknowledgments to report.

FUNDING

This research was supported by a fund (2018-ER6203-02) by Research of Korea Disease Control and Prevention Agency and the "National Institute of Health" research project (project No. 2021-ER1004-00, 2021-ER1004-01).

CONFLICTS OF INTEREST

The authors have no conflict of interest to report.

REFERENCES

- [1] Bang J, Spina S, Miller BL (2015) Frontotemporal dementia. *Lancet* **386**, 1672-1682.
- [2] Rohrer JD, Guerreiro R, Vandrovcova J, Uphill J, Reiman D, Beck J, Isaacs AM, Authier A, Ferrari R, Fox NC, Mackenzie IR, Warren JD, de Silva R, Holton J, Revesz T, Hardy J, Mead S, Rossor MN (2009) The heritability and genetics of frontotemporal lobar degeneration. *Neurology* **73**, 1451-1456.
- [3] Ramos EM, Dokuru DR, Van Berlo V, Wojta K, Wang Q, Huang AY, Miller ZA, Karydas AM, Bigio EH, Rogalski E, Weintraub S, Rader B, Miller BL, Gorno-Tempini ML, Mesulam MM, Coppola G (2020) Genetic screening of a large series of North American sporadic and familial frontotemporal dementia cases. *Alzheimers Dement* 16, 118-130.
- [4] Greaves CV, Rohrer JD (2019) An update on genetic frontotemporal dementia. J Neurol 266, 2075-2086.
- [5] Kim EJ, Kwon JC, Park KH, Park KW, Lee JH, Choi SH, Jeong JH, Kim BC, Yoon SJ, Yoon YC, Kim S, Park KC, Choi BO, Na DL, Ki CS, Kim SH (2014) Clinical and genetic analysis of MAPT, GRN, and C9orf72 genes in Korean patients with frontotemporal dementia. *Neurobiol Aging* 35, 1213.e13-17.
- [6] Kim EJ, Kim YE, Jang JH, Cho EH, Na DL, Seo SW, Jung NY, Jeong JH, Kwon JC, Park KH, Park KW, Lee JH, Roh JH, Kim HJ, Yoon SJ, Choi SH, Jang JW, Ki CS, Kim SH (2018) Analysis of frontotemporal dementia, amyotrophic lateral sclerosis, and other dementia-related genes in 107 Korean patients with frontotemporal dementia. *Neurobiol Aging* **72**, 186.e1-.e7.
- [7] Knopman DS, Kramer JH, Boeve BF, Caselli RJ, Graff-Radford NR, Mendez MF, Miller BL, Mercaldo N (2008) Development of methodology for conducting clinical trials

in frontotemporal lobar degeneration. Brain 131, 2957-2968.

- [8] Rascovsky K, Hodges JR, Knopman D, Mendez MF, Kramer JH, Neuhaus J, van Swieten JC, Seelaar H, Dopper EG, Onyike CU, Hillis AE, Josephs KA, Boeve BF, Kertesz A, Seeley WW, Rankin KP, Johnson JK, Gorno-Tempini ML, Rosen H, Prioleau-Latham CE, Lee A, Kipps CM, Lillo P, Piguet O, Rohrer JD, Rossor MN, Warren JD, Fox NC, Galasko D, Salmon DP, Black SE, Mesulam M, Weintraub S, Dickerson BC, Diehl-Schmid J, Pasquier F, Deramecourt V, Lebert F, Pijnenburg Y, Chow TW, Manes F, Grafman J, Cappa SF, Freedman M, Grossman M, Miller BL (2011) Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia. *Brain* 134, 2456-2477.
- [9] Gorno-Tempini ML, Hillis AE, Weintraub S, Kertesz A, Mendez M, Cappa SF, Ogar JM, Rohrer JD, Black S, Boeve BF, Manes F, Dronkers NF, Vandenberghe R, Rascovsky K, Patterson K, Miller BL, Knopman DS, Hodges JR, Mesulam MM, Grossman M (2011) Classification of primary progressive aphasia and its variants. *Neurology* **76**, 1006-1014.
- [10] Kim EJ, Park KW, Lee JH, Choi S, Jeong JH, Yoon SJ, Kim BC, Kwon JC, Ku BD, Kim SH, Choi BO, Na DL (2014) Clinical and neuropsychological characteristics of a nationwide hospital-based registry of frontotemporal dementia patients in Korea: A CREDOS-FTD Study. *Dement Geriatr Cogn Dis Extra* 4, 242-251.
- [11] Jung KS, Hong KW, Jo HY, Choi J, Ban HJ, Cho SB, Chung M (2020) KRGDB: The large-scale variant database of 1722 Koreans based on whole genome sequencing. *Database* (*Oxford*) 2020, baz146.
- [12] Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL; ACMG Laboratory Quality Assurance Committee (2015) Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 17, 405-424.
- [13] Jang JH, Kwon MJ, Choi WJ, Oh KW, Koh SH, Ki CS, Kim SH (2013) Analysis of the C9orf72 hexanucleotide repeat expansion in Korean patients with familial and sporadic amyotrophic lateral sclerosis. *Neurobiol Aging* 34, 1311.e7-e9.
- [14] Kim EJ, Koh SH, Ha J, Na DL, Seo SW, Kim HJ, Park KW, Lee JH, Roh JH, Kwon JC, Yoon SJ, Jung NY, Jeong JH, Jang JW, Kim HJ, Park KH, Choi SH, Kim S, Park YH, Kim BC, Kim YE, Kwon HS, Park HH, Jin JH (2021) Increased telomere length in patients with frontotemporal dementia syndrome. *J Neurol Sci* **428**, 117565.
- [15] Murrell JR, Spillantini MG, Zolo P, Guazzelli M, Smith MJ, Hasegawa M, Redi F, Crowther RA, Pietrini P, Ghetti B, Goedert M (1999) Tau gene mutation G389R causes a tauopathy with abundant pick body-like inclusions and axonal deposits. *J Neuropathol Exp Neurol* 58, 1207-1226.
- [16] Kim M, Kim HJ, Koh W, Li L, Heo H, Cho H, Lyoo CH, Seo SW, Kim EJ, Nakanishi M, Na DL, Song J (2020) Modeling of frontotemporal dementia using iPSC technology. *Int J Mol Sci* 21, 5319.
- [17] Kitamoto T, Ohta M, Doh-ura K, Hitoshi S, Terao Y, Tateishi J (1993) Novel missense variants of prion protein in Creutzfeldt-Jakob disease or Gerstmann-Sträussler syndrome. *Biochem Biophys Res Commun* 191, 709-714.
- [18] Yagi R, Miyamoto R, Morino H, Izumi Y, Kuramochi M, Kurashige T, Maruyama H, Mizuno N, Kurihara H, Kawakami H (2014) Detecting gene mutations in Japanese

Alzheimer's patients by semiconductor sequencing. *Neurobiol Aging* **35**, 1780.e1-e5.

- [19] Bermingham N, Cowie TF, Paine M, Storey E, McLean C (2008) Frontotemporal dementia and Parkinsonism linked to chromosome 17 in a young Australian patient with the G389R Tau mutation. *Neuropathol Appl Neurobiol* 34, 366-370.
- [20] Chaunu MP, Deramecourt V, Buée-Scherrer V, Le Ber I, Brice A, Ehrle N, El Hachimi K, Pluot M, Maurage CA, Bakchine S, Buée L (2013) Juvenile frontotemporal dementia with parkinsonism associated with tau mutation G389R. J Alzheimers Dis 37, 769-776.
- [21] Sun L, Chen K, Li X, Xiao S (2017) Rapidly progressive frontotemporal dementia associated with MAPT mutation G389R. J Alzheimers Dis 55, 777-785.
- [22] Rossi G, Marelli C, Farina L, Laurà M, Maria Basile A, Ciano C, Tagliavini F, Pareyson D (2008) The G389R mutation in the MAPT gene presenting as sporadic corticobasal syndrome. *Mov Disord* 23, 892-895.
- [23] Niewidok B, Igaev M, Sündermann F, Janning D, Bakota L, Brandt R (2016) Presence of a carboxy-terminal pseudorepeat and disease-like pseudohyperphosphorylation critically influence tau's interaction with microtubules in axon-like processes. *Mol Biol Cell* 27, 3537-3549.
- [24] Giau VV, Senanarong V, Bagyinszky E, An SSA, Kim S (2019) Analysis of 50 neurodegenerative genes in clinically diagnosed early-onset Alzheimer's disease. *Int J Mol Sci* 20, 1514.
- [25] Sung W, Kim YE, Kim SH (2021) Hereditary frontotemporal dementia linked to the pathogenic p.L266V variant of the MAPT gene in Korea. J Clin Neurol 17, 478-480.
- [26] Nozaki I, Hamaguchi T, Sanjo N, Noguchi-Shinohara M, Sakai K, Nakamura Y, Sato T, Kitamoto T, Mizusawa H, Moriwaka F, Shiga Y, Kuroiwa Y, Nishizawa M, Kuzuhara S, Inuzuka T, Takeda M, Kuroda S, Abe K, Murai H, Murayama S, Tateishi J, Takumi I, Shirabe S, Harada M, Sadakane A, Yamada M (2010) Prospective 10-year surveillance of human prion diseases in Japan. *Brain* 133, 3043-3057.
- [27] Takada LT, Kim MO, Cleveland RW, Wong K, Forner SA, Gala II, Fong JC, Geschwind MD (2017) Genetic prion disease: Experience of a rapidly progressive dementia center in the United States and a review of the literature. Am J Med Genet B Neuropsychiatr Genet 174, 36-69.
- [28] Bagyinszky E, Giau VV, Youn YC, An SSA, Kim S (2018) Characterization of mutations in PRNP (prion) gene and their possible roles in neurodegenerative diseases. *Neuropsychiatr Dis Treat* 14, 2067-2085.
- [29] Choi BY, Kim SY, Seo SY, An SS, Kim S, Park SE, Lee SH, Choi YJ, Kim SJ, Kim CK, Park JS, Ju YR (2009) Mutations at codons 178, 200-129, and 232 contributed to the inherited prion diseases in Korean patients. *BMC Infect Dis* 9, 132.
- [30] Kang YJ, Kim KH, Jang SH, Lee GH, Lee YJ, Kim YS, Kim EJ (2019) Diffusion-weighted imaging negative M232R familial Creutzfeldt-Jakob disease. *J Clin Neurosci* 64, 47-49.
- [31] Beck J, Collinge J, Mead S (2012) Prion protein gene M232R variation is probably an uncommon polymorphism rather than a pathogenic mutation. *Brain* 135, e209.
- [32] Moore KM, Nicholas J, Grossman M, McMillan CT, Irwin DJ, Massimo L, Van Deerlin VM, Warren JD, Fox NC, Rossor MN, Mead S, Bocchetta M, Boeve BF, Knopman DS, Graff-Radford NR, Forsberg LK, Rademakers R, Wszolek ZK, van Swieten JC, Jiskoot LC, Meeter LH, Dopper EG, Papma JM, Snowden JS, Saxon J, Jones M, Pickering-

Brown S, Le Ber I, Camuzat A, Brice A, Caroppo P, Ghidoni R, Pievani M, Benussi L, Binetti G, Dickerson BC, Lucente D, Krivensky S, Graff C, Öijerstedt L, Fallström M, Thonberg H, Ghoshal N, Morris JC, Borroni B, Benussi A, Padovani A, Galimberti D, Scarpini E, Fumagalli GG, Mackenzie IR, Hsiung GR, Sengdy P, Boxer AL, Rosen H, Taylor JB, Synofzik M, Wilke C, Sulzer P, Hodges JR, Halliday G, Kwok J, Sanchez-Valle R, Lladó A, Borrego-Ecija S, Santana I, Almeida MR, Tábuas-Pereira M, Moreno F, Barandiaran M, Indakoetxea B, Levin J, Danek A, Rowe JB, Cope TE, Otto M, Anderl-Straub S, de Mendonça A, Maruta C, Masellis M, Black SE, Couratier P, Lautrette G, Huey ED, Sorbi S, Nacmias B, Laforce R Jr, Tremblay ML, Vandenberghe R, Damme PV, Rogalski EJ, Weintraub S, Gerhard A, Onyike CU, Ducharme S, Papageorgiou SG, Ng ASL, Brodtmann A, Finger E, Guerreiro R, Bras J, Rohrer JD; FTD Prevention Initiative (2020) Age at symptom onset and death and disease duration in genetic frontotemporal dementia: An international retrospective cohort study. *Lancet Neurol* **19**, 145-156.